



2018

Using Crosslinked Hyaluronic Acid (HA) and Collagen Scaffolds with Sustained Brain-Derived Neurotrophic Factor (BDNF) Release for Post-SCI Nerve Regeneration

Panth Doshi

Virginia Commonwealth University

Follow this and additional works at: <https://scholarscompass.vcu.edu/uressposters>

 Part of the [Molecular, Cellular, and Tissue Engineering Commons](#)

© The Author(s)

Downloaded from

Doshi, Panth, "Using Crosslinked Hyaluronic Acid (HA) and Collagen Scaffolds with Sustained Brain-Derived Neurotrophic Factor (BDNF) Release for Post-SCI Nerve Regeneration" (2018). *Undergraduate Research Posters*. Poster 265.
<https://scholarscompass.vcu.edu/uressposters/265>

This Book is brought to you for free and open access by the Undergraduate Research Opportunities Program at VCU Scholars Compass. It has been accepted for inclusion in Undergraduate Research Posters by an authorized administrator of VCU Scholars Compass. For more information, please contact libcompass@vcu.edu.



VCU

Using Crosslinked Hyaluronic Acid (HA) and Collagen Scaffolds with Sustained Brain-Derived Neurotrophic Factor (BDNF) Release for Post-SCI Nerve Regeneration



Panth Doshi
Virginia Commonwealth University
Honors College



BACKGROUND

- Traumatic events resulting in spinal cord injuries (SCIs) often leave people paralyzed or with partial loss of motor function below the site of injury. The physical disabilities arising from traumatic events **prevent people from functioning at the same level as pre-injury.**

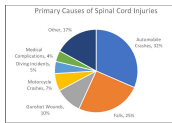


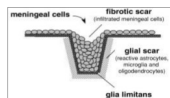
Figure 1: Most common traumatic events that result in spinal cord injury.

- The immune response after SCIs protects the blood brain barrier. **Astrocyte hypertrophy** and release of **chondroitin sulfate proteoglycans (CSPGs)** are chemical barriers to regeneration. Meanwhile, the formation of the **glia limitans** is a physical barrier to regeneration.

Cell	Proteoglycan	Core	Location	Effect on Regeneration
Astrocyte	Hyaluronan	HA	BBB	BBB
	Chondroitin sulfate	CS	BBB	BBB
	Heparan sulfate	HS	BBB	BBB
Microglia	Hyaluronan	HA	BBB	BBB
	Chondroitin sulfate	CS	BBB	BBB
	Heparan sulfate	HS	BBB	BBB
Ependymal cell	Hyaluronan	HA	BBB	BBB
	Chondroitin sulfate	CS	BBB	BBB
	Heparan sulfate	HS	BBB	BBB
Oligodendrocyte	Hyaluronan	HA	BBB	BBB
	Chondroitin sulfate	CS	BBB	BBB
	Heparan sulfate	HS	BBB	BBB
Astrocyte	Hyaluronan	HA	BBB	BBB
	Chondroitin sulfate	CS	BBB	BBB
	Heparan sulfate	HS	BBB	BBB

Figure 2: Common CSPGs act as chemical inhibitors to nerve regeneration

Figure 3: Rupture of the blood brain barrier results in a physical barrier to nerve regeneration



METHODS

A literature review on the inhibitory post-SCI environment and the efficacy of currently used hyaluronic acid (HA) and collagen scaffolds was completed to propose a viable scaffold polymer to regenerate tissue and restore neural function.

RESULTS

- HA scaffolds and collagen scaffolds reduce leukocyte extravasation, decreasing inflammation, to counteract the inhibitory post-SCI environment. HA scaffold implants resulted in less inflammation, lower GFAP intensity, and level of CSPGs
- Immune response, signified by inflammatory cells and different leukocyte types, shifted away from the center of injury to the periphery
- Implanting HA scaffolds and collagen scaffolds into injured spinal cords resulted in more neurofilaments than the natural healing process
- Regenerating neurons had more linear longitudinal growth and more myelinated axons
- Linear aligned nerve regeneration is correlated with better functional recovery than random regeneration

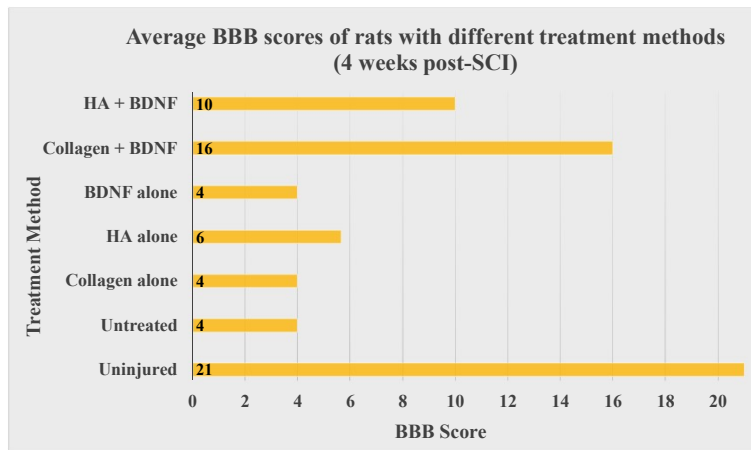
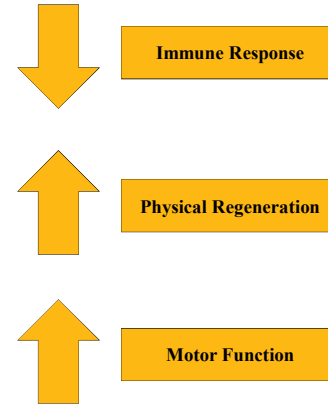
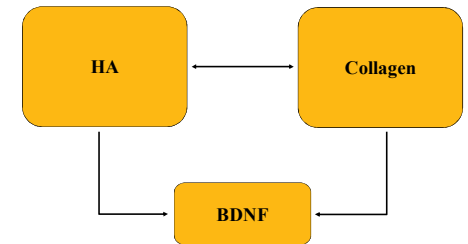


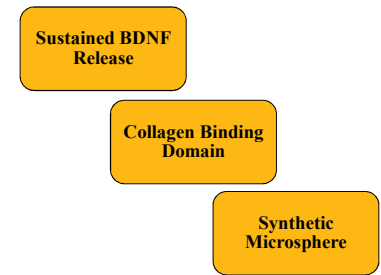
Figure 4: On average, scaffolds alone or growth factors alone are unable to restore much motor function in rats. However, by combining a scaffold polymer with BDNF, functional recovery increases greatly

PROPOSAL

- HA scaffold with sustained BDNF release through synthetic microspheres and linear ordered collagen scaffold with sustained BDNF release through collagen binding domain both have high levels of functional recovery
- Combine effective HA + BDNF scaffold with effective collagen + BDNF scaffold



- Crosslink HA and collagen for base scaffold polymer, then adding in collagen binding domain and embedding in synthetic microspheres for sustained BDNF release



- Further research needs to be completed on:
 - Possible crosslinkers
 - Microsphere polymers
 - Effectiveness of proposed scaffold

REFERENCES

Chen, Y., Tang, Y., Vogel, L. C., & DeVivo, M. J. (2013). Causes of spinal cord injury. *Topics in Spinal Cord Injury Rehabilitation*, 19(1), 1-8. doi:10.1310/sci1901-1

Fuhrmann, T., Obermeyer, J.M., Tator, C.H., & Shoichet, M.S. (2015). Click-crosslinked injectable hyaluronic acid hydrogel is safe and biocompatible in the intrathecal space for ultimate use in regenerative strategies of the injured spinal cord. *Methods*, 84, 60-69. doi:10.1016/j.ymeth.2015.03.023

Han, Q., Sun, W., Lin, H., Zhao, W., Gao, Y., Zhao, Y., Chen, B., ... Dai, J. (2009). Linear ordered collagen scaffolds loaded with collagen-binding brain-derived neurotrophic factor improve the recovery of spinal cord injury in rats. *Tissue Engineering Part A*, 15(10), 2927-2935. doi:10.1089/ten.tea.2008.0506

Park, J., Lim, E., Back, S., Na, H., Park, Y., & Sun, K. (2009). Nerve regeneration following spinal cord injury using matrix metalloproteinase-sensitive, hyaluronic acid-based biomimetic hydrogel scaffold containing brain-derived neurotrophic factor. *Journal of Biomedical Materials Research Part A*, 93(3), 1091-1099. doi:10.1002/jbma.32519

Shearer, M. C., & Favcett, J. W. (2001). The astrocyte/meningeal cell interface — A barrier to successful nerve regeneration? *Cell and Tissue Research*, 305(2), 267-273. doi:10.1007/s004410100384

Siebert, J. R., Steencken, A. C., & Osterhout, D. J. (2014). Chondroitin sulfate proteoglycans in the nervous system: Inhibitors to repair. *Biomed Research International*, 2014, 1-15. doi:10.1155/2014/845323

Wen, Y., Yu, S., Wu, Y., Ju, R., Wang, H., Liu, Y., Wang, Y., & Xu, Q. (2016). Spinal cord injury repair by implantation of structured hyaluronic acid scaffold with PLGA microspheres in the rat. *Cell and Tissue Research*, 364(1), 17-28. doi:10.1007/s00441-015-2298-1

ACKNOWLEDGMENTS

I would like to thank Professor Mary Boyes, Jennifer Mak, and the HONR 200 teaching assistants for their guidance throughout my research project. Special thanks to Herb Hill and UROP for printing this poster.

FURTHER INFORMATION

Panth Doshi is a Biomedical Engineering major in the Honors College at VCU.

Email: doship@vcu.edu

LinkedIn: linkedin.com/in/doship