ERalpha isoforms modulate the tumorigenicity of 24R,25(OH)2D3 in estrogen-responsive cancer

Anjali Verma
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ERα isoforms modulate the tumorigenicity of 24R,25(OH)₂D₃

in estrogen-responsive cancer

A thesis submitted in partial fulfillment of the requirements for the degree of Doctor

of Philosophy at Virginia Commonwealth University

by

ANJALI VERMA

B.Sc. Biomedical Engineering, Case Western Reserve University 2013

April 2019

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Results
Discussion

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<td>1α,25(OH)D₃</td>
<td>1α,25-dihydroxyvitamin D₃</td>
</tr>
<tr>
<td>24S,25(OH)D₃</td>
<td>24S,25-dihydroxyvitamin D₃</td>
</tr>
<tr>
<td>25(OH)D₃</td>
<td>25-hydroxyvitamin D₃</td>
</tr>
<tr>
<td>2BP</td>
<td>2-bromopalmitate</td>
</tr>
<tr>
<td>AA</td>
<td>African American</td>
</tr>
<tr>
<td>AF1</td>
<td>Transcriptional activation domain 1</td>
</tr>
<tr>
<td>AF2</td>
<td>Transcriptional activation domain 2</td>
</tr>
<tr>
<td>ANOVA</td>
<td>Analysis of variance</td>
</tr>
<tr>
<td>AR</td>
<td>Androgen receptor</td>
</tr>
<tr>
<td>BAX</td>
<td>B cell lymphoma protein 2-associated X protein</td>
</tr>
<tr>
<td>BCL2</td>
<td>B cell lymphoma protein 2</td>
</tr>
<tr>
<td>BME</td>
<td>Basement membrane extract</td>
</tr>
<tr>
<td>CA</td>
<td>Caucasian American</td>
</tr>
<tr>
<td>CD-FBS</td>
<td>Charcoal-dextran stripped fetal bovine serum</td>
</tr>
<tr>
<td>CXCL12</td>
<td>C-X-C- motif chemokine 12</td>
</tr>
<tr>
<td>CXCR4</td>
<td>Chemokine receptor type 4</td>
</tr>
<tr>
<td>DAG</td>
<td>Diacylglycerol production</td>
</tr>
<tr>
<td>DPBS</td>
<td>Distilled phosphate buffered saline</td>
</tr>
<tr>
<td>E₂</td>
<td>17β-estradiol</td>
</tr>
<tr>
<td>ECM</td>
<td>Extracellular matrix</td>
</tr>
<tr>
<td>EdU</td>
<td>5-ethynyl-2'-deoxyuridine</td>
</tr>
<tr>
<td>EGF</td>
<td>Epidermal-like growth factor</td>
</tr>
<tr>
<td>ELISA</td>
<td>Enzyme-linked immunosorbent assay</td>
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ER  Estrogen receptor
ER-  ER negative
ER+  ER positive
ERBB2  Erythroblastic oncogene B
Erk  Extracellular signal-regulated kinases
ERs  Estrogen receptors
ERα36  Estrogen receptor α 36
ERα66  Estrogen receptor α 66
ERα66-  ERα66 negative
ERα66+  ERα66 positive
ERβ  Estrogen receptor β
FBS  Fetal bovine serum
FGF  Fibroblast growth factor
GAPDH  Glyceralde-3-phosphate dehydrogenase
GFP  Green fluorescent protein
GPER1 a.k.a. GPR30  G-protein coupled estrogen receptor
H&E  Hematoxylin and eosin
HG DMEM  High-glucose Dulbecco’s minimum essential medium
I3K  Phosphor-inositol-3-kinase
IGF-1  Insulin-like growth factor
IP₂  Inositol triphosphate
IRB  Institutional review board
KD  Knockdown
LPA  Lysophosphatidic acid
LSCC  Laryngeal squamous cell carcinoma
<table>
<thead>
<tr>
<th>Abbreviation</th>
<th>Definition</th>
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<tbody>
<tr>
<td>MAPK</td>
<td>Mitogen-activated protein kinase</td>
</tr>
<tr>
<td>MBC</td>
<td>Methyl β-cyclodextrin</td>
</tr>
<tr>
<td>M-CSF</td>
<td>Macrophage colony stimulating factor</td>
</tr>
<tr>
<td>MEM</td>
<td>Minimum essential growth media</td>
</tr>
<tr>
<td>MMP1</td>
<td>Matrix metalloproteinase 1</td>
</tr>
<tr>
<td>MMPs</td>
<td>Matrix metalloproteinases</td>
</tr>
<tr>
<td>NP-40</td>
<td>Nonidet p40</td>
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<tr>
<td>NSG</td>
<td>NOD.Cg-Prkd&lt;sup&gt;scid&lt;/sup&gt;I&lt;sup&gt;II&lt;/i&gt;rg&lt;sup&gt;tm1Wjl&lt;/sup&gt;/SzJ</td>
</tr>
<tr>
<td>OC</td>
<td>Osteoclast</td>
</tr>
<tr>
<td>OCGM</td>
<td>Osteoclast precursor growth medium</td>
</tr>
<tr>
<td>OCPs</td>
<td>Osteoclast precursors</td>
</tr>
<tr>
<td>OCT</td>
<td>Optimal cutting temperature compound</td>
</tr>
<tr>
<td>OPG</td>
<td>Osteoprotegerin</td>
</tr>
<tr>
<td>ORF</td>
<td>Open-reading-frame</td>
</tr>
<tr>
<td>OS</td>
<td>Overall survival</td>
</tr>
<tr>
<td>OV</td>
<td>Overexpressed</td>
</tr>
<tr>
<td>RPMI</td>
<td>Roswell Park Memorial Institute 1640 media</td>
</tr>
<tr>
<td>PBS</td>
<td>Phosphate buffered saline</td>
</tr>
<tr>
<td>PGE&lt;sub&gt;2&lt;/sub&gt;</td>
<td>Prostaglandin E&lt;sub&gt;2&lt;/sub&gt;</td>
</tr>
<tr>
<td>PKB/Akt</td>
<td>Protein kinase B</td>
</tr>
<tr>
<td>PKC</td>
<td>Protein kinase C</td>
</tr>
<tr>
<td>PLA&lt;sub&gt;2&lt;/sub&gt;</td>
<td>Phospholipase A&lt;sub&gt;2&lt;/sub&gt;</td>
</tr>
<tr>
<td>PLC</td>
<td>Phospholipase C</td>
</tr>
<tr>
<td>PLD</td>
<td>Phospholipase D</td>
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<tr>
<td>PR</td>
<td>Progesterone receptor</td>
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<tr>
<td>Abbreviation</td>
<td>Definition</td>
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<tr>
<td>PTH</td>
<td>Parathyroid hormone</td>
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<tr>
<td>RANK</td>
<td>Nuclear factor kappa-B</td>
</tr>
<tr>
<td>RANKL</td>
<td>RANK ligand a.k.a. Nuclear factor kappa-B ligand</td>
</tr>
<tr>
<td>RFP</td>
<td>Red fluorescent protein</td>
</tr>
<tr>
<td>RIPA</td>
<td>Radioimmunoprecipitation assay</td>
</tr>
<tr>
<td>SNAI1</td>
<td>Snail family transcriptional repressor</td>
</tr>
<tr>
<td>TCGA</td>
<td>Total Cancer Genome Atlas</td>
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<tr>
<td>TDAAC</td>
<td>Tissue and Data Acquisition and Analysis Core</td>
</tr>
<tr>
<td>TdT</td>
<td>Terminal deoxynucleotidyl transferase</td>
</tr>
<tr>
<td>TNBC</td>
<td>Triple-negative breast cancer</td>
</tr>
<tr>
<td>UCSC</td>
<td>University of California Santa Cruz</td>
</tr>
<tr>
<td>VCU</td>
<td>Virginia commonwealth university</td>
</tr>
<tr>
<td>VDR</td>
<td>Vitamin D receptor</td>
</tr>
<tr>
<td>VDRE</td>
<td>Vitamin D response element</td>
</tr>
<tr>
<td>VEGF</td>
<td>Vascular endothelial growth factor</td>
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Abstract

Over 200,000 cases of breast cancer are diagnosed every year. Nearly 20% of these patients supplement their diets with some form of vitamin D. This high frequency of vitamin D supplement use may be due in part to research suggesting that cancer patients with higher serum vitamin D_3 levels have better prognoses than patients with low serum vitamin D_3. However, double-blind clinical trials on the efficacy of vitamin D_3 supplementation in breast cancer have been inconclusive. A recent meta-analysis showed evidence of reduced cancer recurrence in patients taking vitamin D_3 supplements who had ‘estrogen receptor positive’ (ERα66+) breast cancer, but not those who had estrogen receptor negative’ (ERα66-) breast cancer.

Once ingested, vitamin D_3 is metabolized in the liver into the circulating pre-hormone 25(OH)D_3, which is then further metabolized into 1α,25(OH)_2D_3 and 24R,25(OH)_2D_3. 24R,25(OH)_2D_3 has been shown to activate a number of membrane signaling pathways, some of which overlap with 17β-estradiol (E_2) signaling through ERα36, a membrane isoform of ERα66. The central hypothesis of this thesis was that 24R,25(OH)_2D_3 is tumorigenic in certain cancers and that this tumorigenicity is mediated in part by ERα isoforms.

E_2 signaling through ERα36 has been described in the ERα66-, ERα36+ breast cancer cell line HCC38\(^1\). Specific aim 1 determined whether E_2 signaling through ERα36 was tumorigenic other cancers with different ERα profiles. Specific aim 2 determined how 24R,25(OH)_2D_3 affected tumorigenicity in breast cancer using the common breast cancer cell line MCF7 (ERα66+, ERα36+) as a model. Specific aim 3 investigated the role of ERα isoforms in 24R,25(OH)_2D_3 signaling in breast cancer cell lines by comparing
the tumorigenic effects of 24R,25(OH)_{2}D_{3} in MCF7 cells (ERα66+, ERα36+) and HCC38 cells (ERα66-, ERα36+). To determine whether ERα66 regulates the effects of 24R,25(OH)_{2}D_{3}, ERα66 was expressed in two ERα66- cell lines. The effect of 24R,25(OH)_{2}D_{3} on apoptosis was assessed in wild-type and ERα-expressing cell lines.
Chapter 1.

Introduction

Nearly 1 in 8 women will be diagnosed with breast cancer in her lifetime. Approximately 20% of these women will supplement their diets with some form of vitamin D. This high frequency of vitamin D supplement use may be due in part to reports that suggest that cancer patients with higher serum 25-hydroxyvitamin D (25(OH)D) levels have better prognoses and increased survival rates than patients with low 25(OH)D. However, double-blind clinical trials and some pre-clinical studies investigating the efficacy of vitamin D supplementation on cancer patient outcomes have been inconclusive, and recent large-scale studies do not show any correlation between vitamin D supplement use and cancer incidence. One reason for this may be that most double-blind clinical trials do not account for the heterogeneous nature of breast cancer in their analyses. A recent meta-analysis of vitamin D serum levels in breast cancer patients showed that elevated serum vitamin D is associated with a reduced risk of tumor recurrence in patients with 'estrogen receptor positive' (ER+) disease, but did not show a similar association between vitamin D status and cancer recurrence in patients with 'estrogen receptor negative' (ER-) tumors (Figure 1.1). This suggests that estrogen receptor status may be a factor in the efficacy of vitamin D supplementation in cancer.

Breast cancer is a multifaceted, heterogeneous disease with numerous molecular subtypes that
influence disease severity. These molecular subtypes can be demarcated by the expression of hormone receptors, such as the estrogen receptor. Estrogen receptor status is a discrete identification paradigm that allows clinicians to divide patients into meaningful groups for personalized treatment regimens. Patients with ER+ tumors generally have better outcomes than patients with ER- tumors. This is in part because ER+ breast cancer responds favorably to treatment with anti-estrogen drugs. However, the mechanisms governing the more aggressive pathology of ER- tumors compared to their less aggressive ER+ counterparts have not been fully elucidated.

One reason for this may lie in the intrinsically misleading histological classifications assigned to 'ER+' and 'ER-' tumors. In actuality, ER+ and ER- tumor classification denote the intra-tumoral expression of only one estrogen receptor – the transcriptionally active steroid hormone receptor ERα66. Both ER+ and ER- tumors express many other estrogen receptors, such as the membrane receptor ERα36, an isoform of ERα66; estrogen receptor β (ERβ), another transcriptionally active ER11; and G-protein coupled receptor 30 (GPR30), a membrane estrogen receptor12.

Another reason for different serum vitamin D3 levels in ER+ vs. ER- patients may be because serum vitamin D3 is actually a measure of 25(OH)D3, which is in itself a precursor to the active steroid hormone metabolites 1α,25(OH)2D3 and 24R,25(OH)2D3. Once ingested, vitamin D3 is metabolized in the liver into the circulating pre-hormone 25(OH)D3, which is then further metabolized into 1α,25(OH)2D3 and 24R,25(OH)2D313. Research has shown that 1α,25(OH)2D3 has anti-tumorigenic effects in certain cancers, including both ER+ and ER- breast cancer14. However, little has been done to study the effects of 24R,25(OH)2D3 on cancer tumorigenicity.
The effect of 24R,25(OH)_2D_3 on ER+ vs. ER- breast cancer tumorigenicity is particularly interesting because 24R,25(OH)_2D_3 has been shown to activate a number of membrane signaling pathways, some of which overlap with 17\beta-estradiol (E_2) signaling through the membrane receptor ER\alpha. In chondrocytes, 24R,25(OH)_2D_3 has been shown to rapidly increase the activity of phospholipase-D2 (PLD2)\textsuperscript{15} and subsequently activate protein kinase C (PKC)\textsuperscript{16}. Studies with 24S,25(OH)_2D_3, the enantiomer of 24R,25(OH)_2D_3, have shown that this rapid pathway is unique to the 24R,25(OH)_2D_3 metabolite \textsuperscript{17}. Downstream of PKC, 24R,25(OH)_2D_3 promotes p53 ubiquitination and subsequent degradation, reduces the ratio of B cell lymphoma protein 2-associated X protein to B cell lymphoma protein 2 (BAX/BCL2), inhibits cytochrome C translocation, and blocks caspase-3 cleavage to prevent apoptosis\textsuperscript{18}.

The receptor that mediates the actions of 24R,25(OH)_2D_3 has not yet been definitively identified. However, three candidate receptors have been suggested: a membrane isoform of the vitamin D receptor (mVDR), which binds 24R,25(OH)_2D_3 and can be isolated from tibial chick fracture calluses\textsuperscript{19,20}; catalase-1, an enzyme isolated from chick intestine cell membranes which binds 24R,25(OH)_2D_3\textsuperscript{21}; and family 57 b2 (FAM47B2), a transmembrane protein in articular cartilage cells which synthesizes ceramide sphingolipids in cartilage and binds 24,25(OH)_2D_3 with a higher affinity than 1,25(OH)_2D_3\textsuperscript{22}. All three of these receptors candidates are associated with membrane-signaling complexes. This, coupled with the 24R,25(OH)_2D_3 pathway, suggests that 24R,25(OH)_2D_3 is modulated by a membrane-associated protein signaling complex\textsuperscript{23}.

Many estrogen receptors act through membrane-associated pathways. GPR30, for example, is a G-protein coupled receptor (GPCR) estrogen receptor which signals...
through the epidermal growth factor receptor (EGFR) to activate the phosphoinositol-3-kinase (PI3K) pathway. Downstream of PI3K, GPR30 signaling prevents apoptosis and enhances metastasis. Similarly, ERβ has been described as a potent estrogen receptor with a high affinity for 17β-estradiol (E₂). However, ERβ has primarily been described as a classical genomic nuclear receptor rather than a membrane receptor. It does have some extra-nuclear activity, but this extra-nuclear activity has been most well-described in the mitochondria and cytoplasm or in conjunction with other estrogen receptors such as GPR30 at the membrane.

The membrane isoform of ERα (ERα36), on the other hand, has been shown to signal at the membrane through the second messengers PKC and PLD. Our lab has shown that in both ER+ and ER- cancers, rapid E₂ signaling through ERα36 activates PKC, which subsequently activates PLD. This is a similar pathway to that described by 24R,25(OH)₂D₃ in chondrocytes. However, as stated above, 24R,25(OH)₂D₃ activates PLD, then PKC; while the E₂-ERα36 signaling cascade activates PKC before PLD. The similarity in these two pathways suggests crosstalk between the 24R,25(OH)₂D₃ pathway and E₂-ERα36 pathway may be partially regulating the effects of 24R,25(OH)₂D₃ on ER+ and ER- breast cancer.

It is important to note that while our work has shown that the rapid effects of E₂ on PKC at the membrane are the result of ERα36 signaling, other labs have shown that ERα46 and ERα66, the nuclear and cytosolic classic isoforms of ERα, can be trafficked to the membrane via palmitoylation. Once at the membrane, signaling by E₂-ERα66 may mimic E₂-ERα36 signaling. Therefore, it is necessary to investigate both the ERα36 and ERα66 isoforms for the possibility of crosstalk between 24R,25(OH)₂D₃, and membrane-
Modulation of 24R,25(OH)₂D₃ Tumorigenicity by ERα Isoforms in E₂-Responsive Cancer • Spring 2019

ERα signaling. The parallel mechanisms and potential crosstalk between the signaling mechanisms of 24R,25(OH)₂D₃ and ERα36 led us to investigate the role of 24R,25(OH)₂D₃ in breast cancer.

Our lab has shown that in both ER+ and ER- breast cancer, E₂ binds to ERα36 and activates PKC-PLD mediated signaling pathways at the membrane that act downstream to prevent apoptosis and promote proliferation. In chondrocytes, 24R,25(OH)₂D₃ binds to a membrane receptor to initiate a similar signaling cascade that prevents apoptosis. The overall goal of this thesis was to determine the effect of 24R,25(OH)₂D₃ on tumorigenicity in breast cancer, and to determine whether those effects were modulated by ERα isoforms.

E₂ signaling through ERα36 has been described in the ERα66-, ERα36+ breast cancer cell line HCC38. Specific aim 1 determined whether E₂ signaling through ERα36 was tumorigenic in other cancers that have different ERα profiles. The tumorigenicity of E₂ was assessed in two laryngeal squamous cell carcinoma (LSCC) cell lines with different ERα isoform expression. Cancer stage and metastasis was correlated with ERα isoform expression in primary LSCC tumor samples and matched normal controls to assess correlations between ERα isoform expression and tumor aggression.

Specific aim 2 determined how 24R,25(OH)₂D₃ affected tumorigenicity in breast cancer using the common breast cancer cell line MCF7 (ERα66+, ERα36+) as a model. The tumorigenicity of 24R,25(OH)₂D₃ was assessed in vitro by examining DNA synthesis and markers for apoptosis, epithelial-to-mesenchymal transition and metastasis, and in vivo in a xenograft mammary fat pad murine tumor model. The mechanism of 24R,25(OH)₂D₃ signaling in MCF7 (ERα66+, ERα36+) was identified using chemical and
Specific aim 3 investigated the role of ERα isoforms in 24R,25(OH)_{2}D_{3} signaling in breast cancer cell lines by comparing the tumorigenic effects of 24R,25(OH)_{2}D_{3} in two breast cancer cells: MCF7 (ERα_{66+}, ERα_{36+}) and HCC38 (ERα_{66-}, ERα_{36+}). Tumorigenicity was assessed \textit{in vivo} and \textit{in vitro}, and the mechanism of 24R,25(OH)_{2}D_{3} signaling in HCC38 (ERα_{66-}, ERα_{36+}) was elucidated using chemical and antibody inhibitors and enzyme activity assays. To determine which isoform of ERα regulates the effects of 24R,25(OH)_{2}D_{3} on apoptosis ERα_{66} was overexpressed in HCC38 (ERα_{66-}, ERα_{36+}). To investigate whether ERα_{66} was only tumorigenic in breast cancer cell lines, ERα was overexpressed in an unrelated cell line, C2C12 (ERα_{66-}, ERα_{36+}). The effect of 24R,25(OH)_{2}D_{3} on apoptosis was assessed in wild-type and ERα-overexpressed cell lines. The \textit{central hypothesis} of these aims was that 24R,25(OH)_{2}D_{3} is tumorigenic in certain cancers and that this tumorigenicity is mediated in part by ERα isoforms.

**Specific Aim 1: Determine the estrogen receptor profile and estrogen responsiveness in laryngeal cancer and clinical outcomes.**

There is growing evidence that laryngeal squamous cell carcinomas (LSCC) are responsive to sex hormones, specifically 17β-estradiol (E_{2}), despite the controversy regarding the presence and characterization of E_{2} receptors (ER). Determination of sex hormone responsiveness impacts the prognosis of LSCC patients and the treatment modalities implemented by their clinicians. Discovery of membrane-associated steroid hormone receptors and rapid membrane signaling opened the possibility that cancers previously labeled ‘non-hormone dependent,’ and ‘ER negative’ might be susceptible to
the effects of E₂ via these membrane receptors. The **objective** of this aim was to show that the expression of different membrane ERs in LSCC is not uniform, which may result in differential and antagonistic responses to E₂. This aim **hypothesized** that E₂ signaling through ERα36 was tumorigenic, and that primary LSCC aggression was proportional to ERα36 and inversely proportional to ERα66 expression.

ERα isoform expression was characterized in LSCC cell lines and primary clinical samples. Tumorigenicity of E₂ in LSCC was assessed by examining the effects of E₂ on proliferation and apoptosis *in vitro* and tumor burden in a mouse xenograft model *in vivo*. A possible E₂ signaling mechanism was determined by using chemical signaling inhibitors and an ERα36-blocking antibody. Associations between ERα expression and LSCC aggression were confirmed using data from the Total Cancer Genome Atlas.

**Specific Aim 2: Determine if 24R,25-Dihydroxyvitamin D₃ regulates breast cancer cells in vitro and in vivo.**

Epidemiological studies indicate that high serum 25-hydroxyvitamin D₃ [25(OH)D₃] is associated with improved prognosis and survival in some breast cancer patients. Pre-clinical studies attributed this to the anti-tumorigenic properties of its steroid hormone metabolite 1α,25(OH)₂D₃. However, 1α,25(OH)₂D₃ is highly calcemic and thus has a narrow therapeutic window. Here we investigate the effects of another steroid hormone metabolites of 25(OH)₂D₃, 24R,25(OH)₂D₃, as an alternative non-calcemic vitamin D₃ supplement. The **objective** of this aim was to assess the effect of 24R,25(OH)₂D₃, a steroid hormone metabolite of vitamin D₃, on tumorigenic markers in the MCF7 (ERα66+, ERα36+), an ERα66+, ERα36+ breast cancer cell line. We **hypothesized** that
24R,25(OH)₂D₃ would be anti-tumorigenic in MCF7 and that a rapid membrane-associated mechanism would stimulate this effect.

NOD-SCID-IL2γR null female mice with MCF7 xenografts in the mammary fat pad were treated with 24R,25(OH)₂D₃ and changes in tumor burden and metastases were assessed *in vivo*. Breast cancer cell lines were treated *in vitro* with 24R,25(OH)₂D₃ to determine its effects on proliferation, apoptosis, migration, and metastatic markers. A possible 24R,25(OH)₂D₃ signaling mechanism was determined by using chemical signaling inhibitors and an ERα36-blocking antibody to determine the dependence of ERα isoforms on 24R,25(OH)₂D₃ signaling.

**Specific Aim 3: Determine if 24R,25(OH)₂D₃ differentially regulates tumorigenicity in estrogen receptor α dependent manner.**

Our lab has identified an understudied metabolite of vitamin D₃, 24R,25(OH)₂D₃, which has anti-tumorigenic effects in the ERα66+, ERα36- breast cancer cell line MCF7. However, preliminary evidence suggests that ERα66 and ERα36 may modulate the anti-tumorigenic effects of 24R,25(OH)₂D₃. The objective of this aim was to evaluate the effect of 24R,25(OH)₂D₃ in the breast cancer cell line HCC38 (ERα66-, ERα36+) and determine the role of ERα66 and ERα36 on 24R,25(OH)₂D₃ tumorigenicity. We hypothesized that 24R,25(OH)₂D₃ would be pro-tumorigenic in HCC38 and that this effect would be modulated by ERα66.

NOD-SCID-IL2γR null female mice with HCC38 xenografts in the mammary fat pad were treated with 24R,25(OH)₂D₃ and changes in tumor burden were assessed *in vivo*. Breast cancer cell lines were treated *in vitro* with 24R,25(OH)₂D₃ to determine its effects.
on proliferation, apoptosis, migration, and metastatic markers. A possible 24R,25(OH)₂D₃ signaling mechanism was determined by enzymatic activity assays and by using chemical signaling inhibitors and an ERα36-blocking antibody to determine the dependence of ERα isoforms on 24R,25(OH)₂D₃ signaling. To determine which isoform of ERα regulates the effects of 24R,25(OH)₂D₃ on apoptosis ERα66 was overexpressed in the ERα66-negative HCC38. To investigate whether ERα66 was only tumorigenic in breast cancer cell lines ERα was also overexpressed in an unrelated cell line, C2C12. The effect of 24R,25(OH)₂D₃ on apoptosis was assessed in wild-type and ERα-overexpressed cell lines.
Chapter 2.

Background and Literature Review

Steroid hormones mediate a wide variety of vital developmental and physiological functions in different organs, such as the reproductive system, bone, brain, and fat differentiation and metabolism. When steroid hormones are deficient or abnormal, they have deleterious effects such as promoting the development and progression of hormone-responsive cancers\(^28\). Traditionally, steroid hormones signal by binding to receptors in the cytosol followed by either homodimerization or heterodimerization to form a complex, which is then translocated to the nucleus. In the nucleus, the complex functions as a transcription factor by binding to either specific response elements or to transcription initiation complexes on the DNA to either activating or repressing transcription\(^29\).

This process occurs in a matter of hours to days\(^30,31\). In contrast, it is now well-accepted that most steroid hormones induce a non-nuclear rapid signaling response that occurs in seconds to minutes, which is not delayed by inhibition of transcription or translation\(^31–35\). The signaling response is a result of steroid hormones binding to specific extra-nuclear receptors, which mainly reside in the cell membrane, particularly the caveolae. When activated, these receptors trigger signaling cascades that involve kinases, phospholipases, calcium flux, and other secondary messengers\(^36,37\).

Nature of steroid-hormone membrane receptors

Rapid signaling is mediated through receptors that reside in the cell membrane. Many of these receptors act as G protein-coupled receptors upon ligation, leading to
signaling cascades that regulate existing protein function and downstream gene expression. The nature of these receptors is a matter of controversy. Some have advocated that they are the traditional receptors residing in the cell membrane, particularly concerning receptors for estrogen, progesterone, androgen, and glucocorticoids, and the vitamin D receptor. Others have suggested that they are truncated isoforms of traditional nuclear membrane receptors. For example, the well-described membrane estrogen receptor ERα36 is a truncated isoform of the canonical estrogen receptor ERα66. However, others have identified novel orphan receptors that also mediate rapid membrane signaling.

**Classical nuclear receptor trafficking**

Nuclear receptors for sex hormones have been found in the plasma membranes of hormone-responsive cancers where they activate rapid signaling pathways that enhance cancer cell survival and proliferation. It is thought that the classical receptors exert their effects through transactivation of the epidermal-like growth factor (EGF) or insulin-like growth factor-1 (IGF-1) tyrosine kinase receptors to stimulate kinase cascades. Another theory suggests that classical receptors signal through collaboration with G-protein coupled receptors.

Trafficking of the classical receptors to the plasma membrane occurs through palmitoylation, which involves the attachment of palmitic acid to an internal cysteine residue. Palmitoylation promotes the association of the receptor with caveolin-1, a transporter protein, to caveolae rafts in the plasma membrane. Palmitoylation occurs in a nine amino acid motif, which includes the cysteine palmitoylation site. This
palmitoylation motif is highly conserved in estrogen receptor (ER) isoforms α and β, progesterone receptor (PR) and androgen receptor (AR)\textsuperscript{35,40}. Once the receptor is associated with the caveolae, it may interact with many signaling proteins associated with the caveolae rafts and activate various Gα and Gβ/γ proteins within seconds\textsuperscript{53,54}.

**Estrogen Receptor α and its isoforms**

In the last ten years, several isoforms of ERα have come to light \textsuperscript{43}. The most widely studied of these alternative receptors are ERα46 and ERα36, a membrane-bound variant of the classical ERα, ERα66. Both ERα46 and ERα36 arise from alternative splicing of the ESR1 gene \textsuperscript{42}, with ERα36 including a novel exon of the gene, exon 9 \textsuperscript{42}. ERα36 has been identified in many breast cancer cell lines that were previously characterized as ERα negative \textsuperscript{1}, such as HCC38 and MDA-MB-231 \textsuperscript{55}. In breast cancer, the presence of ERα36 has been associated with increased aggression and metastasis \textit{in vitro}, \textit{in vivo}, and \textit{in situ} \textsuperscript{56,57}. In one study, a continuum of ERα66/ERα36 positive/negative cell line-derived xenografts was examined \textit{in vivo}. ERα66+, ERα36- tumors found to be the least aggressive, ERα66+, ERα36+ slightly more aggressive, ERα66-, ERα36- second-most aggressive, and ERα66-, ERα36+ tumors the most aggressive of all four phenotypes. This suggests that the relative levels of ERα66 and ERα36 within a tumor are more important modulators of breast cancer aggression than the presence of each receptor individually. However, it should be noted that this study did not examine levels of ERα46, which could have further modulated tumor aggression. ERα46 has been found in some breast cancer cell lines, but little is known about its function as an estrogen receptor independent of ERα66 and ERα36 \textsuperscript{27,58}.
In a sampling of breast cancer cells, MCF7, which is widely regarded as a less-aggressive breast cancer cell line\textsuperscript{59}, was found to have high levels of ERα\textsubscript{66+}, low levels of ERα\textsubscript{46}, and moderate levels of ERα\textsubscript{36} (Figure 2.1). HCC38, a moderately aggressive breast cancer cell line\textsuperscript{1}, had no ERα\textsubscript{66}, moderate levels of ERα\textsubscript{46}, and moderate levels of ERα\textsubscript{36} (Figure 2.1). MDA-MB-231, an aggressively metastatic breast cancer cell line\textsuperscript{60}, had no ERα\textsubscript{66}, high levels of ERα\textsubscript{46}, and moderate levels of ERα\textsubscript{36} (Figure 2.1). It is not clear which of these isoforms or what combination of isoforms determines the relative levels of aggression or metastatic potential of each cell line; however, it is crucial to take the expression of each of the ERα isoforms into account when classifying them as ‘ER positive’ or ‘ER negative.’

Figure 2.1 Western blot data of ERα isoforms in breast cancer cells.

[A] ERα\textsubscript{66}, [B] ERα\textsubscript{46}, and [C] ERα\textsubscript{36} normalized to GAPDH and expressed as mean normalized signal intensity ± standard error in MCF7, HCC38, and MDA-MB-231 breast cancer cell lines\textsuperscript{61}.
**Estrogen and estrogen receptors in laryngeal cancer**

Although the importance of estrogen and estrogen receptor signaling has been well-established in breast cancer, estrogen signaling is also important in other hormonally responsive cancers. The larynx is a secondary sex organ that undergoes trophic changes in response to hormonal shifts during puberty, the role of steroid hormones and steroid hormone receptors in laryngeal squamous cell carcinoma (LSCC) has been understudied. Researchers first began to describe functional estrogen receptors (ERs) in LSCC cell lines over three decades ago\(^{62,63}\). These studies led to a small clinical trial on the use of tamoxifen, a potent anti-estrogen, in the treatment of LSCC\(^{64,65}\). However, the trial failed within six months because there was no observed clinical response or reduction in tumor size. This largely ended investigations into ER and the use of anti-estrogens in the treatment of LSCC.

ER\(\alpha\)\(^{36}\) is present in both ER\(\alpha\)\(^{66}\) positive (ER\(\alpha\)\(^{66}+\)) and ER\(\alpha\)\(^{66}\) negative (ER\(\alpha\)\(^{66}-\)) LSCC samples\(^{66–68}\). However, unlike breast cancer\(^{27}\), ER\(\alpha\)\(^{36}\) expression in LSCC is highly variable. This suggests that crosstalk between rapid and genomic signaling pathways mediated by multiple ERs may control the estrogen response in LSCC. This section highlights some of the rapid estrogen signaling pathways previously described in LSCC and suggests that clinical decisions regarding prognosis and treatment of LSCC may be informed by the ER\(\alpha\) profiles of individual LSCC patients.

Previous studies by our laboratory show that loss of nuclear ER staining, but not membrane ER staining (as measured by immunohistochemistry) is associated with reduced patient prognoses and increased lymph node metastases in LSCC\(^{66}\). This is similar to trends observed in ER- breast cancer\(^{66,67,69}\). A loss of estrogen receptor \(\beta\)
expression (ERβ) has been associated with increased aggression in LSCC\textsuperscript{70}. Changes in the non-traditional transmembrane ER G-protein coupled estrogen receptor (GPER1) and ERα\textsubscript{36} have also been associated with malignancies observed in histology of human vocal fold tissue\textsuperscript{71} and \textit{in vitro} experiments on human LSCC cells\textsuperscript{66,67}.

**Estrogen receptor signaling**

Classical estrogen signaling is mediated by the nuclear receptors ERα and ERβ. E\textsubscript{2} binding to these receptors in the cytoplasm leads to hetero-or homo-dimerized ERs and the translocation of the E\textsubscript{2}-ER ligand-receptor complex to the nucleus. Here it acts as a transcription factor for genes associated with proliferation, cell survival, metastasis, invasion, migration, and others\textsuperscript{68,72}. Simultaneously, membrane-associated E\textsubscript{2}-ER binding triggers many rapid signal transduction pathways, such as phosphoinositol-3-kinase (PI3K) / protein kinase B (PKB, also known as Akt), extracellular signal-regulated kinases (ERK), and mitogen-activated protein kinase (MAPK)\textsuperscript{68,73}. Both the canonical and rapid signaling pathways induced by E\textsubscript{2}-ER binding stimulate pro-tumorigenic phenotypes. In this way E\textsubscript{2} is a potent mitogen and EMT stimulus, promoting invasion and metastasis in prostate, ovarian, breast, uterine, and LSCC cells\textsuperscript{67,74–79}.

E\textsubscript{2}-ER interactions encompass a complex network of canonical and rapid responses. In addition to stimulating a number of mitogenic and cell survival pathways in LSCC as described below, E\textsubscript{2} also stimulates the transcription of its own receptors. Canonically, the ESR1 gene, which encodes ERα, has both distal and proximal promoter regions. The E\textsubscript{2}-ER complex directly stimulates the transcription of ESR1 by binding to one or more of these promoters in certain cell types\textsuperscript{80,81} and indirectly stimulates them by
binding to other receptors that can enhance ER transcription downstream (such as EGFR). Treatment with dose-dependent E2 increased gene expression of ERα36 and ERα66 (Figure 2.2).

![Graph](image)

**Figure 2.2** E2 upregulates the expression of ERα36 and ERα66 in UM-SCC-12, a laryngeal cancer cell line.

Rapid Estrogen Signaling

Estrogen, like other steroid hormones, initiates some non-nuclear rapid signaling pathways that initiate a number of mitogenic, anti-apoptotic, invasive, and metastases-promoting signaling mechanisms. Many of these pathways involve phospholipases, kinases and other secondary messengers such as phospholipase D (PLD) and protein kinase C (PKC). These may result in altered mRNA transcripts downstream, but these non-genomic pathways are still distinct from classical signaling in that they do not directly involve the activation or suppression of DNA response elements by the E2-ER complex.

One commonly studied membrane-associated estrogen receptor is ERα36. ERα36 lacks the transcriptional activation domains AF1 and AF2, which implies that ERα36 is incapable of directly stimulating transcription by binding to gene promoter sites. This lack of transcriptional activity distinguishes the actions of ERα36 from the transcriptionally active canonical receptor ERα66. ERα36 does, however, retain the DNA binding, ligand binding, and partial dimerization zones of the canonical protein. Thus ERα36 may act as a co-factor to dimerize with other, possibly transcriptionally active, ERs. ERα36 also has a unique function mediated by 27 amino acids transcribed from the novel exon 9 of the ESR1 gene. Although the specific function of these 27 amino acids has not been fully described, research suggests that they are involved in the membrane-associated signaling actions of ERα36. Breast cancer cells transfected with ERα36 lacking exon 9 are unable to trigger the rapid membrane-associated signaling described for wild-type ERα36, suggesting a unique function for ERα36 independent from its role as a truncated dimerizing ERα66 variant.
ERα36 is highly expressed in many breast cancer tumors that were previously diagnosed as ER negative. In these tumors, E2 signaling through ERα36 stimulates some tumorigenic and cell cycle disrupting pathways. Evidence of E2 signaling in ERα66-,ERα36+ cells has called into question traditional pathological classifications that describe ERα66 negative breast cancers as ‘estrogen non-responsive.’ Furthermore, in addition to promoting E2 signaling, unlike the traditional ERα66, high expression of ERα36 is associated with unfavorable clinical prognosis, suggesting that ERα36 might have value as a prognostic biomarker.

Similar trends have been observed in LSCC, with ERα36-expressing cancers expressing more markers of clinical aggression than their ERα66 positive counterparts. Some studies have shown that E2 increases proliferation in LSCC cells through both genomic and non-genomic mechanisms. Our lab and others have shown that E2 stimulation of ERα36 triggers PKC activation. This mechanism is also present in breast cancer cells. In vitro activation of ERα36 by E2 in HEp-2 and UM-SCC-12 cells inhibits apoptosis and enhances proliferation, and activates PLD and PKC (Figure 2.3). Further investigation into the mechanism of membrane E2 signaling has revealed that the pro-proliferative and anti-apoptotic mechanisms of E2 are dependent upon PLD and PKC activation. However, the specific receptors which control E2 signaling in laryngeal cancer have not been identified (Figure 2.3).
**Figure 2.3 Mechanism of E2 signaling through ERα36.**

E2 activates ERα36 in the caveolae and leads to Gq-mediated phospholipase D and PLC pathways. PLD activates PI3K-mediated anti-apoptotic pathways, and PLC activates PKC-ERK pathways, leading to proliferation and metastasis.
24R,25(OH)\textsubscript{2}D\textsubscript{3} signaling

A similar parallel mechanism has been demonstrated in 24R,25(OH)\textsubscript{2}D\textsubscript{3} signaling in chondrocytes \textsuperscript{18,92,93}, which has been shown to inhibit apoptosis through an as yet unidentified membrane receptor. Once ingested, vitamin D\textsubscript{3} – the most bioavailable form of vitamin D – is hydroxylated in the liver into 25(OH)D\textsubscript{3}, the circulating form of vitamin D\textsubscript{3} commonly measured as ‘serum vitamin D’ \textsuperscript{13}. 25(OH)D\textsubscript{3} is then further hydroxylated in the kidneys to become the steroid hormones 1,25(OH)\textsubscript{2}D\textsubscript{3} or 24,25(OH)\textsubscript{2}D\textsubscript{3} \textsuperscript{13}. 1α,25(OH)\textsubscript{2}D\textsubscript{3} has been widely studied because of its role in regulated calcium homeostasis in concert with parathyroid hormone (PTH) \textsuperscript{94}. 1α,25(OH)\textsubscript{2}D\textsubscript{3} is also an essential hormone for bone health because of its role in sequestering calcium within the bones to maintain bone mineralization and strength. 24R,25(OH)\textsubscript{2}D\textsubscript{3} is not as well studied, partially because it is often regarded as a ‘waste product’ that cycles with 1α,25(OH)\textsubscript{2}D\textsubscript{3} to regulate the production of the latter hormone. However, studies by our lab and others have demonstrated a role for 24R,25-(OH)\textsubscript{2}D\textsubscript{3} in growth plate development and fracture callus healing, where 24R,25-(OH)\textsubscript{2}D\textsubscript{3} has an anti-apoptotic effect and is an essential regulator of chondrocyte sensitivity to 1α,25(OH)\textsubscript{2}D\textsubscript{3}.

24R,25(OH)\textsubscript{2}D\textsubscript{3} plays a key role in fracture healing and chondrocyte differentiation via a membrane-mediated PLD dependent pathway \textsuperscript{93,95}. Unlike 1α,25(OH)\textsubscript{2}D\textsubscript{3}, which helps regulate calcium mineralization of the lower parts of the growth plate, 24R,25(OH)\textsubscript{2}D\textsubscript{3} is essential for cell cycle regulation in earlier stages of the growth plate, particularly in the progression of pre-proliferative chondrocytes to proliferative chondrocytes \textsuperscript{96}. Recent studies have suggested that 24R,25(OH)\textsubscript{2}D\textsubscript{3} or the 24-hydroxylase that produces it, CYP24A1, may modulate cell proliferation and apoptosis in cancer cells \textsuperscript{97–99}. This effect
may be a direct result of the activities of either the hormone or enzyme; or may be the result of modulation of the effects of 1α,25(OH)₂D₃. 24,25(OH)₂D₃ has been shown to regulate the production of the 1-hydroxylase enzyme CYP27B1, which converts 25(OH)D₃ to 1,25(OH)₂D₃. In a similar manner, 24R,25(OH)₂D₃ promotes chondrocyte sensitivity to 1α,25(OH)₂D₃, encouraging a positive feedback loop where each hormone regulates the production of the other, resulting in 1α,25(OH)₂D₃ and 24R,25(OH)₂D₃ cycling within the body.

As described above, 24R,25(OH)₂D₃ has been shown to signal through an as-yet unidentified receptor to stimulate PLD and subsequent anti-apoptosis response in chondrocytes. The identity of this receptor has been a matter of debate for decades. Larsson et al., while studying the lysosomal membranes of chick intestinal cells, reported that catalase-1 specifically binds 24R,25(OH)₂D₃, suggesting a rapid peroxidase-involved signal transduction pathway for 24,25(OH)₂D₃. A number of papers from the lab of Dr. Anthony Norman have reported evidence for a membrane-bound receptor for 24R,25(OH)₂D₃ in chick fracture healing callus that may be involved in rapid or genomic signal transduction. Crystallographic studies of the classical vitamin D receptor (VDR) suggest that conformational changes in the receptor may accommodate the transient binding of 24R,25(OH)₂D₃ to the ligand-binding pocket of VDR. Recently, another 24R,25(OH)₂D₃ receptor, FAM57B2, was identified by Martineau et al., which may be essential to the 24R,25(OH)₂D₃ response in fracture callus repair. Numerous studies in rat growth plate chondrocytes have characterized a specific membrane-associated signal transduction pathway that mediates both classical genomic and rapid responses of
24R,25(OH)₂D₃, strongly suggesting the presence of a membrane receptor or signaling complex that regulates the actions of 24R,25(OH)₂D₃₁⁷,2₃,9₃,1₀₃.

While a consensus receptor for 24R,25(OH)₂D₃ remains elusive, membrane signaling by 24R,25(OH)₂D₃ has been described. In costochondral resting zone chondrocytes, 24R,25(OH)₂D₃ binds to an as yet unidentified membrane-bound receptor and inhibits phospholipase A₂ activity (PLA₂)₁⁰⁴. The resulting signaling pathway alters fatty acid turnover and subsequently inhibits the release of arachidonic acid and prostaglandin E₂ (PGE₂) production₂₃,1₀₃,1₀₅. This inhibition of arachidonic acid turnover alters membrane fluidity and calcium flux within the cell, modulating signal transduction downstream and stimulating PKC activity₂₃,1₀₃,1₀₆,1₀₇. The effect of 24R,25(OH)₂D₃ is stereospecific; 24S,25(OH)₂D₃ does not activate this pathway, providing further evidence that the response to the hormone is receptor mediated.

24R,25(OH)₂D₃ binding to the membrane receptor also stimulates PLD₁, which increases diacylglycerol production (DAG), to in turn activate PKCα₁₅,₁₆,₁₈. This increase in PKCα has been observed in whole cell lysate and cell membrane fractions, while 24R,25(OH)₂D₃ has been shown to increase PKCζ in resting zone matrix vesicles₂₃,1₀₈.

Although 24,25(OH)₂D₃ stimulation of PLD₂ activates DAG to stimulate PKC, this subsequent activation of PKC does not coincide with the translocation of PKC to the plasma membrane₁₆. This suggests that 24R,25(OH)₂D₃ stimulation of PKC activity via PLD may not be a direct result of 24R,25(OH)₂D₃ binding to a membrane receptor and is instead a result of downstream signaling by other secondary messengers. Furthermore, although resting zone cells have active phospholipase C (PLC) isoforms – namely PLCβ₁ and PLCβ₃ – neither PLC is activated by 24R,25(OH)₂D₃, and inhibition of PLC does not
affect 24R,25(OH)2D3 stimulation of PKC15,17. Together with the inhibition of PLA2 and stimulation of DAG act to enhance PKC activity to stimulate the phosphorylation of multiple proteins in multiple anti-apoptotic downstream pathways, including MAPK and ERK, which are also stimulated by 17β-estradiol through both genomic and non-genomic pathways36,47,83,109,110 (Figure 2.4).

**Figure 2.4 24R,25 signaling through PLD.**

24R,25(OH)2D3 binds to an unidentified membrane receptor to stimulate PLD2 in resting zone chondrocytes. Activation of PLD2 stimulates lysophosphatidic acid (LPA), which stimulates PLC and PI3K. Downstream of PI3K, 24R,25(OH)2D3 inhibits p53 ubiquitination in chondrocytes, preventing apoptosis.
Vitamin D3 supplementation in cancer.

Preclinical studies and meta-analyses have suggested that 1,25(OH)\(_2\)D\(_3\) and 25(OH)D\(_3\) may have potential as a preventive therapy for breast cancer. Despite this, clinical studies investigating vitamin D supplementation on breast cancer prognosis have been largely inconclusive \(^8,111–113\). Oral vitamin D3 has been shown to increase cell turnover, autophagy, and reduce tumor growth in the mammary glands of normal mice \(^14,114\), and intravenous 1α,25(OH)\(_2\)D\(_3\) has been shown to reduce tumor growth in ER-positive breast cancer xenografts \(^14\). \textit{In vitro}, 1α,25(OH)\(_2\)D\(_3\) has been shown to reduce angiogenic, metastatic, and apoptotic markers in ER-positive breast cancer cells, but not in normal cultured human mammary epithelial cells \(^115–118\). This could be a function specific to the interactions of 1α,25(OH)\(_2\)D\(_3\) with the VDR, which regulates 1α,25(OH)\(_2\)D\(_3\)’s genomic actions by dimerizing with the retinoic acid receptor to form a transcription complex \(^119\). Dimerized VDR bound to 1α,25(OH)\(_2\)D\(_3\) has been shown to suppress transcription of the ESR1 gene \(^120\), which could inhibit the feed-forward loop of estrogen-ER\(\alpha\) signaling \(^121\). Alternatively, 1α,25(OH)\(_2\)D\(_3\)’s anti-tumorigenicity could be a function of its non-genomic actions, as its analogs 1,24,25-trihydroxyvitamin D3 and 1,25,26-trihydroxyvitamin D3, which are incapable of binding to VDR, have also been shown to inhibit the growth of breast cancer cells \textit{in vitro} \(^122\).

Studies examining the use of 1α,25(OH)\(_2\)D\(_3\) as a safe, easy to deliver supplement against breast cancer tumorigenicity have been promising; however, there is a risk of hypercalcemia with excess 1α,25(OH)\(_2\)D\(_3\). Hypercalcemia is a common co-morbidity of cancer, with up to 30% of all cancer patients suffering severe hypercalcemia \(^123\). It is the leading cause of hospitalization in cancer patients, and who have breast cancer are...
particularly susceptible \textsuperscript{124}. Excess 1\alpha,25(OH)\textsubscript{2}D\textsubscript{3} is a documented cause of hypercalcemia \textsuperscript{125}, and blood serum levels of free 1\alpha,25(OH)\textsubscript{2}D\textsubscript{3} as low as 200pmol/L (with \sim 85 pmol/L approximating normal 1\alpha,25(OH)\textsubscript{2}D\textsubscript{3} levels) can be toxic \textsuperscript{126}. Many \textit{in vitro} studies only observe the anti-tumorigenic effects of 1\alpha,25(OH)\textsubscript{2}D\textsubscript{3} when treating with 1-100 nmol/L, well above the toxicity threshold for this compound \textsuperscript{127–129}.

The search for non-calcemic analogs of vitamin D\textsubscript{3} that can approximate 1\alpha,25(OH)\textsubscript{2}D\textsubscript{3}’s anti-tumorigenic effects has led researchers to look at modifications of 1\alpha,25(OH)\textsubscript{2}D\textsubscript{3} that do not bind to VDR and thus do not have the same hypercalcemic effects. Some of these studies have been successful, but the majority are not approved for the treatment of cancer \textsuperscript{130}.

Although connections between improved breast cancer prognoses and high vitamin D serum levels have been well-established, there is no consensus on the efficacy of vitamin D supplementation in the treatment of breast cancer. Studies examining breast cancer prognosis and vitamin D serum levels could be improved by considering the breast cancer molecular subtypes, particularly the ER status, of the patients involved. 24R,25(OH)\textsubscript{2}D\textsubscript{3}, an understudied vitamin D\textsubscript{3} metabolite, may have potential as a non-calcemic vitamin D\textsubscript{3} metabolite with tumor modulating properties. Further research is needed to understand the connections between ER status and the anti-tumorigenicity of vitamin D supplements, and the parallels between membrane mediated estrogen signaling through ER\alpha\textsubscript{36} and 24R,25(OH)\textsubscript{2}D\textsubscript{3} may provide insight into this connection.
Chapter 3.

Estradiol Receptor Profile and Estrogen Responsiveness in Laryngeal Cancer and Clinical Outcomes

[Introduction]

Laryngeal cancer, despite arising from a secondary sex organ and having a clear gender disposition, is not uniformly accepted as sex hormone-dependent. The determination of sex hormone responsiveness, specifically to 17β-estradiol (E$_2$), impacts the prognosis of the patients and the treatment modalities that are implemented in these types of cancers. E$_2$ plays a key role in the pathogenesis of hormone-responsive cancers, the most well studied being breast cancer and prostate cancer, by enhancing cell proliferation, survivability, and metastasis. Due to its effects on cancer progression and behavior, the presence of E$_2$ receptors (ER) has long been the most important molecular marker used in the diagnosis and as a predictor of prognosis in breast cancer dictating treatment.

Classically, E$_2$ mediates its effects through interaction with its nuclear receptors, ERα and ERβ. These receptors act as transcription factors by binding to specific response elements, or transcription initiation complexes, on the DNA to either activate or repress transcription in a process that occurs in a matter of hours to days. Steroid hormones also initiate non-nuclear, non-genomic rapid signaling within seconds to minutes that...
occurs independently of transcription or translation. This results from steroid binding to specific extra-nuclear receptors associated with the cell membrane, leading to the activation of signaling cascades that involve kinases, phospholipases, calcium flux and other second messengers, and may result in altered mRNA levels. Recent studies indicate that motifs within ER exons encode sequences associated with palmitoylation, thereby linking the receptor to the membrane.

One of the key candidates for the membrane mediated action of E2 is a novel ERα splice variant with a molecular mass of approximately 36 kDa, which has been named ERα36. ERα36 differs from ERα66, the classic nuclear ER receptor, by lacking both transcriptional activation domains (AF1 and AF2) (exons 7 and 8) but retaining the DNA-binding domain and partial dimerization and ligand-binding domains of the nuclear receptor. This suggests that although it may still bind to DNA, ERα36 may not directly activate transcription. Rather, it may affect transcriptional function as a co-factor due to its ability to dimerize with other ERs and bind to DNA. In addition, ERα36 contains a novel exon 9, which encodes 27 amino acids of unknown function. Breast cancer cells transfected with ERα36 lacking exon 9, lack the ability to trigger rapid activation of membrane associated signaling, suggesting that this may be an important key to its role in E2 responsiveness.

ERα36 is gaining special interest particularly in regards to its role in mediating E2 responsiveness in traditionally diagnosed ER negative tumors, and its role in modulating anti-estrogen resistance in ER positive tumors, which have otherwise been clinically regarded as hormone-unresponsive. Inclusion of this receptor in screens of ER negative breast cancers showed that high levels of ERα36 expression were correlated
with an unfavorable clinical prognosis independent of nuclear ER status. Thus ERα36 has been suggested as an important novel marker for breast cancer clinical characterization, suggesting it may serve a valuable function for laryngeal cancers as well.

ERα36 has the ability to mediate the effects of E2 independently of nuclear ERα, as demonstrated in ERα66 null cells and in studies using antibodies against the nuclear and membrane isoforms of ERα. The receptor has been shown to initiate divergent pathways from the plasma membrane that re-converge downstream to affect cancer cell survivability. E2 binding to ERα36 affects proliferation through the activation of G-proteins, which in turn lead to the activation of phospholipase C (PLC). PLC activation leads to the production of diacylglycerol (DAG) and inositol trisphosphate (IP3), which subsequently trigger calcium signaling and protein kinase C (PKC) activation. Together, calcium signaling and PKC activation culminate in downstream activation of mitogen-activated protein kinase (MAPK) to increase DNA synthesis and gene transcription. At the same time, E2 activation of ERα36 reduces apoptosis by blocking effects of common chemotherapeutics, such as Taxol, by activating phospholipase D (PLD) at the membrane, leading to activation of lysophosphatidic acid (LPA) signaling and phosphoinositol-3 kinase (PI3K). This in turn attenuates the caspase cascade that promotes apoptosis. This mechanism of E2’s action was also shown in laryngeal cancer cells.

As laryngeal cancer is not perceived to be a sex hormone-responsive cancer, most studies do not examine tumor responses or tumorigenesis regulation by steroid hormones in laryngeal cancer. Little can be concluded from the sparse data available on the relation...
of E$_2$ with laryngeal cancer. Even the presence of classical cytosolic ERs in laryngeal cancer is controversial. Several studies have argued that laryngeal cancer does not possess any functional ER$^{135}$, or rather implied that the presence of the receptors originates from stromal or mesenchymal tissues in the larynx and not from the epithelial component, which constitutes the origin of laryngeal cancer$^{136}$. Other studies affirm the presence of classical nuclear ERs in laryngeal cancer$^{137,138}$ and even correlate the presence and number of the receptors with local metastasis status$^{139}$. Despite this lack of agreement, there is growing evidence that laryngeal cancer cells display E$_2$ responsiveness, as E$_2$ was found to increase the proliferation of laryngeal cancer cells$^{89-91}$ via a non-genomic mechanism$^{64}$. Furthermore, 4’hydroxytamoxifen has been demonstrated to inhibit the growth of laryngeal cancer, in part, due to its anti-estrogen effect$^{63,140}$; and this effect is eliminated with the addition of E$_2$$^{63,64,137}$.

Previously, we have shown that Hep2 laryngeal carcinoma epithelial cells express both ER$\alpha$66 and ER$\alpha$36. ER$\alpha$36 was found to reside in the plasma membrane, and stimulation of ER$\alpha$36 by E$_2$ stimulated PKC activation via a mechanism comparable to that shown in breast cancer cells$^1$. In vitro, E$_2$ activation of ER$\alpha$36 was shown to cause a PLD-dependent increase in PKC activity, followed by an upregulation of angiogenic and metastatic factors. ER$\alpha$36 signaling enhanced both proliferation and the anti-apoptotic effect of E$_2$ against chemotherapeutics, indicating a role in tumorigenesis. Furthermore, it was demonstrated both in vitro and in vivo that E$_2$ enhanced vascular endothelial growth factor (VEGF) and fibroblast growth factor (FGF) expression. An association has been suggested between high levels of ER$\alpha$36 and VEGF and a more aggressive tumor phenotype, as evidenced by the correlation to lymph node metastasis$^{66}$.
These findings advocate a role for E$_2$ in the tumorigenesis of laryngeal cancer and raise the option of heterogeneity of expression of ERs and responsiveness in the tumor cells that warrants further exploration. In the present study we studied two laryngeal cancer lines with different ER$_\alpha$ expression to examine the signaling pathways activated by E$_2$ in an attempt to clarify the importance of profiling laryngeal cancer E$_2$ responsiveness and the clinical implications ER profiling might have for the treatment of laryngeal cancer.

**Materials and Methods**

**Cell Culture**

UM-SCC-11A and UM-SCC-12 cells were obtained from the Carey Laboratory at the University of Michigan (Ann Arbor, MI) and cultured in high-glucose Dulbecco’s minimum essential medium (HG DMEM) lacking phenol red (Invitrogen, Carlsbad, CA), supplemented with 10% fetal bovine serum (FBS) (ThermoFisher, Waltham, MA, USA), 2mM L-glutamine, and 100U/100U penicillin-streptomycin (ThermoFisher).

**Presence of ER Isoforms**

ER$_\alpha$ expression in laryngeal cancer cells was identified using sequence-specific primers, designed in our lab previously$^{16}$, to identify traditional ER$_\alpha$66 and the spliced variant ER$_\alpha$36. A commercial primer was used to identify ESR2 (ER$\beta$) (Qiagen, Hilden, Germany). UM-SCC-12 and UM-SCC-11A cells were plated in 24-well plates at 20,000 cells/cm$^2$ (n=6 per variable) and cultured to confluence. RNA was extracted with TRIzol and used to synthesize cDNA libraries (Thermo Fisher Scientific, High Capacity cDNA Reverse Transcription kit). Levels of RNA expression were quantified using gene-specific
primers (Table 3.1). RNA expression for ERα46/66 was done with primers that amplified exon 1 of the ESR1 gene, and primers for ERα36 amplified exon 9 of the ESR1 gene. Data were verified with a commercial primer from Qiagen, which amplified exon 3/4 of ESR1. Expression of ERβ was assessed with a commercial primer from Qiagen, which amplified exon 5/6 of ESR2. All gene expression was quantified using a whole-cell standard and normalized to GAPDH.

<table>
<thead>
<tr>
<th>Gene</th>
<th>Forward Primer</th>
<th>Reverse Primer</th>
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<tbody>
<tr>
<td>ERα46/66</td>
<td>TGCCTCGCCTCTAAACCTCG</td>
<td>TCCAGATGCTTTGTTGTTG</td>
</tr>
<tr>
<td>ERα36</td>
<td>TCCTCGTGTCTAAAGCCTCTG</td>
<td>AAAATGTC CCCACGTCACA</td>
</tr>
<tr>
<td>GAPDH</td>
<td>CTATAAATGGAGCCGCAGCC</td>
<td>TGGCAGCGC AAAAAGAAGATG</td>
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Table 3.1 Human primers used in real-time PCR analysis.

Western blots were used to identify and quantify ER protein in the laryngeal cell lines. Cells were plated at 10,000 cells/cm² in T-75 flasks (Corning) and cultured to confluence. Monolayers were washed twice with 1X PBS and ERα66, ERα46, ERα36, and ERβ were assessed using a western blot assay with GAPDH protein as the loading control. UM-SCC-12, UM-SCC-11A, and MCF7 cells were lysed in 200μL radioimmunoprecipitation assay buffer (RIPA) and resolved on 4-20% Tris-glycine extended gels using gel electrophoresis. Proteins were transferred from the gel onto a PVDF membrane using the Bio-rad Mini Trans-Blot® Electrophoretic Transfer Cell. Membranes were blocked and probed with primary and secondary antibodies according
to the LI-COR Western Blot Protocol. Membranes were then imaged on the Odyssey®CLx Infrared Imaging System from LI-COR.

Response to 17β-Estradiol

17β-Estradiol (#BML-DM200-0050) was purchased from Sigma Aldrich (Farmingdale, NY) and reconstituted to 1mM stock solutions in absolute ethanol. Stock solutions were diluted in warm complete media to final treatment concentrations. For all experiments except DNA synthesis, cells were treated at confluence. Media were aspirated and cell layers were treated with media containing 0 (vehicle), $10^{-9}$M, $10^{-8}$M, or $10^{-7}$M $E_2$ for 9 minutes at 37°C, 5%CO$_2$, 100% humidity. Media were then removed by aspiration and cells were harvested (PKC, PLD assays) or incubated with fresh complete media for 24 hours (p53 and VEGF protein expression).

Specific inhibition of membrane associated factors involved in ERα36 signaling was used to examine whether the effects of $E_2$ were mediated via these mechanisms. PLD was inhibited by wortmannin (VWR, Radnor, PA, #80055-508)$^{15,141,142}$. Caveolae were disrupted using methyl β-cyclodextrin, which depletes the membranes of cholesterol (Millipore-Sigma, St. Louis, MO, #C4555-10G)$^1$. Estrogen receptors were inhibited using a neutralizing antibody to ERα36 (Chi Scientific, Maynard, MA)$^1$. Confluent cultures were pre-treated with media containing different concentrations of the inhibitor as indicated below. After 30 minutes, the media were replaced with fresh media containing either 0, $10^{-8}$, or $10^{-7}$ M $E_2$ for 9 minutes. These media were removed by aspiration and cells were harvested immediately for analysis of PKC and PLD activity. Alternatively, fresh media were added to the cultures for 24 hours and protein expression for p53 and VEGF determined by duo-set ELISA as described below.
DNA Synthesis

Cells were cultured to 70% confluence; then serum-starved in phenol-red free HG DMEM supplemented with 1% charcoal-dextran filtered FBS and 100U/100U penicillin-streptomycin. After 48 hours in starvation media, cells were treated with 17β-estradiol or inhibitors (see above) and incubated with fresh complete media for 20 hours. At that time, cells were pulsed with 10µL of a 1:100 dilution of 5-ethynyl-2’-deoxyuridine (EdU) and incubated for an additional 4 hours. Cells were then harvested and assayed for EdU incorporation according to manufacturer’s instructions (ThermoFisher Scientific).

DNA Fragmentation

Apoptosis was examined by assessing cells for DNA fragmentation by colorimetric terminal deoxynucleotidyl transferase (TdT)-mediated dUTP nick end labeling (Trevigen, TiterTAC™ in situ microplate TUNEL assay). After E2 treatment, cells were incubated with fresh media for 24 hours and fixed with formaldehyde containing 3.7% sucrose before being processed for TUNEL staining following manufacturer’s instructions. Final absorbance was detected at 450 nm using a microplate spectrophotometer.

PKC Specific Activity

Cells were cultured to confluence in 24-well plates and treated with E2 as above; media were removed by aspiration and cells were washed twice with 2mL/well of 1X PBS. Cells were lysed in 200µL of PKC lysis buffer as per manufacturer’s instructions and assayed for PKC activity using the PKC Activity Kit from (AbCam, Cambridge, UK). Cell lysates were assessed for total protein content using the Pierce BCA 660 reagent kit (VWR) and PKC activity was normalized to total protein.
PLD Specific Activity

Cells were cultured to confluence in 24-well plates and treated with E2 as above; media were removed by aspiration and cells were washed twice with 2mL/well of 1X PBS. Cells were lysed in PLD lysis buffer as per manufacturer's instructions and assayed for PLD activity using the PLD Activity Kit from Thermo Fisher Scientific. Cell lysates were assessed for total protein content using the Pierce BCA 660 reagent kit and PLD activity was normalized to total protein.

Total p53 Levels

P53 is a tumor suppressor gene that induces apoptosis. Total p53 levels were measured 24 hours post-treatment using a sandwich ELISA (Human Total p53 DuoSet® IC, R&D Systems, Minneapolis, MN). Cells were plated in 24-well plates, treated with E2 for 9 minutes as described above, and incubated for 24 hours. After 24 hours, cells were lysed in 200μL of 1X PBS, 1mM EDTA, 0.5% Triton™ X-100. Lysates were centrifuged at 10,000g for 15 minutes and the supernatant was collected and assayed for total protein content (ThermoFisher, Pierce 660 nm Protein Assay) and total p53 content following manufacturer’s instructions. Data are presented as a ratio of picograms of total p53 to total protein content.

Statistical Analysis

Data are presented as mean ± standard error of six independent cultures per treatment group. Outliers were removed with Grubbs’ test using a P-value of 0.05. Data with one independent variable were analyzed by analysis of variance (ANOVA) with Tukey’s post-test to determine significance between groups. Groups that do not share a letter are considered significant with p-values < 0.05. Experiments with two independent
variables (DNA synthesis, total p53 content, PKC, and PLD assays) were analyzed with one-way ANOVA as described above and two-way ANOVA with Bonferroni’s modification of Student’s t-test post-tests between all group means using Graphpad Prism 7. P-values <0.05 were considered significant and were marked with * denoting significance between 0 vs. E₂ treatments for a given x-axis group.

Results

Western blots of UM-SCC-12 and UM-SCC011A cells showed that different laryngeal carcinoma cell lines express different sex hormone receptors (Figure 3.1A). As expected, the breast cancer MCF7 cell line possessed ERα66, ERα46, ERα36, and ERβ, confirming the gel system was working. UM-SCC-12 cells expressed mRNA for ERα66 whereas none was found in the UM-SCC-11A cells (Figure 3.1B). Both laryngeal cancer cell lines expressed mRNA for ERα36 (Figure 3.1C), but only UM-SCC-11A cells expressed mRNA for ERα (Figure 3.1C).
Figure 3.1 Estrogen receptor expression in whole cell lysates of UM-SCC-12 and UM-SCC-11A cells.

[A] ERα66, ERα36, and ERβ protein were identified via western blot using MCF7 breast cancer cells as controls. mRNA expression of [B] exon 1 of ESR1 (ERα66), [C] exon 9 of ESR1 (ERα36), and [D] ESR2 (ERβ) was measured by real-time qPCR.
E₂ regulated proliferation in the two cell lines in a differential manner. E₂ caused a slight decrease in DNA synthesis in UM-SCC-11A cells, which was significant at 10⁻⁷M (Figure 3.2A). In contrast, E₂ increased DNA synthesis in UM-SCC-12 cells, which was significant at 10⁻⁸M and 10⁻⁷M (Figure 3.2B). Antibodies to ERα36 had no effect on cell proliferation in untreated UM-SCC-11A cells, but slightly increased proliferation in UM-SCC-11A cells treated with E₂ (Figure 3.2C). In contrast, antibodies to ERα36 did not affect the stimulatory effect of E₂ on cell proliferation in the UM-SCC-12 cells (Figure 3.2D). In UM-SCC-11A cells, inhibition of PLD with wortmannin alone did not affect proliferation. However, treatment with 0.1 and 1 M wortmannin in combination with 10⁻⁷M E₂ reduced DNA synthesis as compared to matched-dose wortmannin treatment alone (Figure 3.2E). Similarly, wortmannin alone did not affect DNA synthesis, but it prevented an E₂-dependent increase in DNA synthesis in UM-SCC-12 cells (Figure 3.2F).

Apoptosis was regulated by E₂ in a cell line-specific manner. E₂ did not have a significant effect on DNA fragmentation measured via TUNEL assay in the UM-SCC-11A cells (Figure 3.3A). In contrast, E₂ caused a dose-dependent inhibition in TUNEL in the UM-SCC-12 cells (Figure 3.3B). E₂ did not affect total p53 in the UM-SCC-11A cells (Figure 3.3C) but reduced significantly p53 content of UM-SCC-12 cells (Figure 3.3D). Depletion of membrane cholesterol with methyl β-cyclodextrin reduced p53 levels to the same extent as 10⁻⁷M E₂ in UM-SCC-12 cells but did not further reduce the inhibitory effect of the hormone (Figure 3.4).
Figure 3.2 UM-SCC-12 and UM-SCC-11A cells were serum-starved and treated with E2 for 9 minutes.

Media were changed, and 24 hours later: [A, B] DNA synthesis was measured using a ClickiT EdU Microplate assay. [C, D] A second set of cultures was pre-treated with media or an antibody to ERα36 and then treated with vehicle or 10^{-7} M E2 for 9 minutes. Media were changed and 24 hours later, DNA synthesis was measured. [E, F] A third set of cultures was pre-treated with wortmannin and then treated with vehicle or 10^{-7} M E2 for 9 minutes before media were changed and DNA synthesis was measured 24 hours later.
Figure 3.3 UM-SCC-12 and UM-SCC-11A cells were treated with E2 for 9 minutes.

[A, B] Media were changed, and 24 hours later, DNA fragments were measured in cultures using TUNEL staining. [C, D] A second set of cultures was treated identically and total intracellular p53 content was quantified with a sandwich ELISA assay.
Figure 3.4 UM-SCC-12 cells were treated with methyl-β-cyclodextrin for 30 minutes, then treated with 10^{-8}M E2 for 9 minutes. Media were changed, and 24 hours later total intracellular p53 content was quantified with a sandwich ELISA assay.
PKC activity in the UM-SCC-12 cells was regulated by a membrane-associated mechanism. E₂ caused a dose-dependent increase in PKC specific activity (Figure 3.5A). This stimulatory effect of E₂ required intact caveolae. Depletion of plasma membrane cholesterol prevented the E₂-dependent increase in enzyme activity (Figure 3.5B) in a biphasic manner, with the maximum effect at 0.1mM methyl-β-cyclodextrin (MBC). 1mM MBC slightly reduced basal PKC activity, but 1mM MBC and 10⁻⁷M E₂ together slightly increased PKC activity (Figure 3.5B). A neutralizing antibody specific to ERα36 increased PKC activity, but the addition of 10⁻⁷M E₂ to cultures treated with the ERα36 antibody did not further increase PKC activity, suggesting that the effects of E₂ in UM-SCC-12 cells are at least partially elicited through the activation of membrane ERα36 (Figure 3.5C).

E₂ caused a biphasic increase in PLD specific activity in the UM-SCC-12 cells that was significant at 10⁻⁸M (In the present study, we observed that the expression of ERs in laryngeal cancer cell lines is not uniform, which might explain the controversies in the literature regarding the presence and nature of the ER in laryngeal cancer. UM-SCC-11A expressed ERα36 and ERβ with no apparent ERα66, while UM-SCC-12 expressed both ERα66 and ERα36 with minimal ERβ. This diversity in ER expression translates to a disparate response to E₂. E₂ had a detrimental effect in UM-SCC-12 cells, enhancing proliferation and reducing apoptosis. UM-SCC-11 cells were much less responsive to E₂, inhibiting DNA synthesis at high concentrations, although not affecting the apoptotic rate. UM-SCC-11A cells expressed high levels of ERβ, which might offer an explanation for the protective effect conferred by E₂. ERβ has been postulated to be a dominant-negative regulator of ERα modulating transcriptional responses to estrogens. The ratio of ERα vs. ERβ within a cell may determine the cell sensitivity
to E2 and its biological responses26. This protective effect of ERβ has been observed in breast cancer cells in which the expression of ERβ was found to inhibit proliferation and cell invasion146, and in colorectal cancer in which the decrease of expression of ERβ was hypothesized to have a role in tumorigenesis147 and might demonstrate the same effect in laryngeal cancer cells.

Figure 3.5 E2 stimulates PKC activity in UM-SCC-12 cell layer lysates.

[A] UM-SCC-12 cells were treated with E2 for 9 minutes. [B] A second set of cultures was treated with methyl-β-cyclodextrin for 30 minutes, then treated with 10⁻⁸M E2 for 9 minutes. [C] A third set of cultures was treated with a neutralizing antibody to ERα36 for 30 minutes, then treated with 10⁻⁸M E2 for 9 minutes. For all cultures, media were aspirated and monolayers were washed twice with 1X PBS. PKC activity in cell layer lysates was measured using a commercial kit and normalized to total protein content and reaction incubation time.
Figure 3.6 E2 stimulates PLD activity in UM-SCC-12 cell layer lysates.

[A] UM-SCC-12 cells were treated with E2 for 9 minutes.

[B] A second set of cultures was treated with methyl-β-cyclodextrin for 30 minutes, then treated with $10^{-8}$M E2 for 9 minutes. [C] A third set of cultures was treated with an antibody to ERα36 for 30 minutes, then treated with $10^{-8}$M E2 for 9 minutes. For all cultures, media were aspirated and monolayers were washed twice with 1X PBS. PLD activity in cell layer lysates was measured using a commercial kit and normalized to total protein content and reaction incubation time.
Discussion

E2 initiates multiple rapid signaling pathways from the cell membrane in different types of cells and via different membrane receptors, many of which remain the subject of active investigation. While there is evidence indicating that these receptors are in fact the traditional receptors residing in the cell membrane, novel orphan receptors have been identified and are being implicated in rapid membrane signaling by E2. Previously we showed that Hep2 laryngeal carcinoma epithelial cells express both traditional ERα66 and the recently identified splice variant, ERα36. ERα36, which was found to be a key cellular and transcriptional regulator of proliferation and enhanced aggressiveness in breast cancer, was shown to reside in the plasma membrane of Hep2 cells and to mediate the activation of PKC in response to E2 via a mechanism comparable to that seen in the breast cancer cells. ERα36 signaling had roles in tumorigenesis of laryngeal cancer enhancing proliferation and in the anti-apoptotic effect of E2 against chemotherapeutics such as Taxol.

In the present study, we observed that the expression of ERs in laryngeal cancer cell lines is not uniform, which might explain the controversies in the literature regarding the presence and nature of the ER in laryngeal cancer. UM-SCC-11A expressed ERα36 and ERβ with no apparent ERα66, while UM-SCC-12 expressed both ERα66 and ERα36 with minimal ERβ. This diversity in ER expression translates to a disparate response to E2. E2 had a detrimental effect in UM-SCC-12 cells, enhancing proliferation and reducing apoptosis. UM-SCC-11 cells were much less responsive to E2, inhibiting DNA synthesis at high concentrations, although not affecting the apoptotic rate.
UM-SCC-11A cells expressed high levels of ERβ, which might offer an explanation for the protective effect conferred by E₂. ERβ has been postulated to be a dominant-negative regulator of ERα modulating transcriptional responses to estrogens. The ratio of ERα vs. ERβ within a cell may determine the cell sensitivity to E₂ and its biological responses. This protective effect of ERβ has been observed in breast cancer cells in which the expression of ERβ was found to inhibit proliferation and cell invasion, and in colorectal cancer in which the decrease of expression of ERβ was hypothesized to have a role in tumorigenesis and might demonstrate the same effect in laryngeal cancer cells.

UM-SCC-12 cells demonstrated responses to E₂ similar to those seen in ERα-positive MCF7 and ERα-negative HCC38 breast cancer cells as well as the Hep2 laryngeal cancer cell line. E₂ caused dose dependent enhancement of proliferation and inhibition of apoptosis. These effects were mediated by the stimulation of the known PLD and PKC signaling cascades, as evidenced by the activation of both enzyme activities in UM-SCC-12 cells in response to E₂ and by the inhibition of proliferation when cultures were pre-treated with wortmannin, a specific PLD inhibitor at the concentrations used in this study. Pretreatment with methyl β-cyclodextrin, which depletes the membranes of cholesterol and thereby disrupts the caveolae, inhibited the activation of PKC and PLD and abolished the anti-apoptotic effect, indicating the importance of the specialized membrane compartment for E₂-dependent signaling pathway in UM-SCC-12 laryngeal cancer cells.

Despite the clear similarity to the mechanism and effects of E₂ seen in our previous work, in UM-SCC-12 cells it seems that ERα36 plays only a part in the membrane
signaling pathways of E2. While antibodies against ERα36 prevented the stimulatory effect of E2 on the activation of PKC, they did not inhibit the effects of E2 on proliferation rate or PLD activity. This suggests that E2 causes an increase in PKC activity and proliferation by activating another estrogen receptor, rather than ERα36 as previously described in HCC38 breast cancer cells66. It should also be noted that while the antibody to ERα36 did not affect cell proliferation in UM-SCC-12 cells, it stimulated both PKC and PLD activity in these cells, suggesting that neutralizing ERα36 stimulates an unidentified mechanism which stimulates PKC and PLD in a compensatory pro-survival pathway. Furthermore, it is possible that the antibody to ERα36 simply stimulated PKC activity to a saturated point, preventing further increases in PKC activity by any stimulant rather than specifically blocking the effects of E2.

The mechanism of E2’s action in the UM-SCC-12 laryngeal cancer cell line is intricate and not yet clarified completely. One hypothesis for the incongruity with previous results is that in UM-SCC-12 cells, the effects of E2 are the result of the convergence and crosstalk of the different membrane receptor pathways, ERα66 and ERα36. An alternate hypothesis could be that the tumorigenic effects of E2 are the result of E2 binding to ERα46 or ERα66, has been shown previously in osteoblasts148, or to an as-yet-uncharacterized novel membrane-bound estrogen receptor (Figure 3.7).

Classical sex hormone nuclear receptors have been found in the plasma membranes of hormone-responsive cancers where they elicit rapid signaling pathways, which enhance cancer cell survival and proliferation46. From the plasma membrane it is thought that the classical receptors exert their effect through transactivation of the EGF or IGF-1 tyrosine kinase receptors in order to stimulate kinase cascades47. It has been
suggested that the mechanism for trafficking of the classical sex hormones to the plasma membrane involves palmitoylation; the attachment of palmitic acid to an internal cysteine residue, which promotes the association of the receptor with caveolin-1, a transporter protein, to caveolae rafts in the plasma membrane. Once the receptor is incorporated in the caveolae it may interact with many signaling proteins incorporated in the caveolae rafts and activate various Gα and Gβ/γ proteins within seconds.

ERs have been the primary receptors studied in order to ascertain this theory of the nature of membrane steroid hormone receptors. Immunological studies found that the membrane-localized estrogen binding protein shared epitope homology to the nuclear receptors. Endothelial cells from combined ERα/ERβ deleted mice and breast cancer MCF-7 cells that lack nuclear ERα as well as small interfering RNA directed against the classical ER, eliminated estrogen binding at the cell surface and the rapid hormone signaling elicited by E2. Transfection of ER cDNA of classical ERα or ERβ in ER-null cells resulted in induction of both nuclear and membrane signaling pathways. Taken together there is evidence to support the role of classical nuclear receptors in membrane mediated rapid responses, which might clarify the effects seen in UM-SCC-12 cells of a synergistic effect of both ERα66 and ERα36 through membrane mediated signaling.

Another plausible explanation for the results found in UM-SCC-12 cells may originate from crosstalk between membrane ER signaling with growth factor receptors, which results in the activation of the common membrane delineated pathways such as ERK-MAPK, PI3k-Akt and Src. Thus once ERα36 has been activated with resulting activation of PKC, the subsequent activation of growth factor receptor pathways may be responsible for the ensuing enhancement of proliferation. Such signaling cascades have been
implicated to have a major role in the carcinogenesis and progression of breast cancer independently and in collaboration with classical genomic E₂ signaling. The interaction between membrane ER and transmembrane growth factor signaling pathways such as insulin-like growth factor I receptor (IGF-IR), epidermal growth factor (EGFR), Src and HER2 and the upregulation of these signaling pathways has been accepted as being one of the mechanisms underlying resistance to endocrine therapy in breast cancer. The activation of these signaling pathways has a variety of documented effects including inhibition of apoptosis, stimulation of cell proliferation, enhanced invasion and cell motility, and induction of angiogenesis.
Figure 3.7 17β-estradiol activates PKC in UM-SCC-11A cells; and PKC and PLD in UM-SCC-12 cells by binding to ERα36 in the caveolae.

In UM-SCC-12, 17β-estradiol activation of PLD results in increased DNA synthesis. 17β-estradiol also activates PLD through an unknown caveolae-bound estrogen receptor to decrease total p53 levels and apoptosis.
The findings of this study present compelling evidence that laryngeal cancer is sex hormone responsive, specifically to E₂. Rapid non-genomic membrane signaling has been found to have an important role in mediating the effect of E₂ in laryngeal cancer cells. These effects are hypothesized to be mediated by the activation of different membrane receptors and subsequent activation of growth factor receptors and tyrosine kinase receptors. The full impact of steroid hormones is the result of the convergence and crosstalk of the membrane and classical nuclear receptor pathways. Their integrative, at times synergistic or alternatively antagonistic, effects confer a variability and complexity to steroid hormone function.

This is emphasized in the two laryngeal cell lines studied, in which one exhibited a protective effect of E₂ while in the other, E₂ had a deleterious effect augmenting proliferation and conferring anti-apoptotic potential to the cancer cells. These findings stress the importance of establishing the molecular and clinical characterization of the specific tumor in order to tailor treatment accordingly, thus optimizing treatment while reducing adverse effects for each individual patient. Further investigation is warranted in order to elucidate the versatile role of E₂ in laryngeal cancer pathogenesis and progression, the array of membrane-mediated mechanisms by which these effects come in to play, and to shed light on their impact on the treatment and management of laryngeal cancer.
Chapter 4.

Loss of estrogen receptors is associated with increased tumor aggression in laryngeal squamous cell carcinoma

[Chapter 4 is in preparation as: Verma A, Schwartz N, Cohen DJ, Patel V, Boyan BD, Schwartz Z. Loss of estrogen receptors is associated with increased tumor aggression in laryngeal squamous cell carcinoma. Scientific Reports. In Preparation]

Introduction

The larynx is an overlooked secondary sex hormone organ. Similar to other sex hormone organs, it undergoes trophic changes in response to hormonal changes during puberty, and morphological changes during adulthood. Steroid hormones have been reported to play a significant role in voice changes during maturation, as their effects mediate the lengthening and thickening of the male vocal folds.\textsuperscript{153} In the female larynx, fluctuations in estrogen levels during the menstrual cycle are accompanied by variations in the pitch of the voice.\textsuperscript{154} Atrophy and dystrophy have been found to be pronounced in the larynges of postmenopausal women as well as in the vocal fold tissue of ovariectomized rats.\textsuperscript{155} These subjects are known to suffer from edema of the laryngeal lamina propria, inflammation in squamous and respiratory epithelia, pseudostratification, and cilia loss, but these symptoms were rectified with estrogen replacement therapy.\textsuperscript{156}

Despite the obvious responsiveness of the larynx to estrogen, the presence and differential expression of estrogen receptors (ER) is a matter of debate, particularly in laryngeal cancer. ER, mainly ER\textsubscript{\alpha}66, the traditional ER translated from the ESR1 gene, has been reported in laryngeal epithelia in both females\textsuperscript{157} and males,\textsuperscript{158} and changes in
ER are associated with inflammation and benign lesions.\textsuperscript{159} ER expression has also been verified in laryngeal cancer\textsuperscript{138,160} and at higher levels than in adjacent normal mucosa.\textsuperscript{139} Moreover, the non-traditional transmembrane ERs G-protein coupled estrogen receptor (GPER1, sometimes called GPR30) and ER\textalpha36, a splice variant of the traditional ER\textalpha66, have been found in human vocal folds\textsuperscript{71} and laryngeal cancer cells,\textsuperscript{66} respectively. Another ER, ER\textbeta, which is encoded by the ESR2 gene, has been inversely correlated with increased cancer aggression in laryngeal squamous cell carcinoma (LSCC).\textsuperscript{70} However, opposing studies have not identified ER\textsuperscript{161} or any other steroid hormone receptors\textsuperscript{162,163} in the larynx or laryngeal cancer cells.\textsuperscript{135} The clinical implication of ER expression in laryngeal cancer is equally confounding. In a previous study, we found a correlation between the number of ERs and regional lymph node metastasis.\textsuperscript{66} This differs from other studies that reported a reduced prognosis\textsuperscript{164} and lower rates of lymph node metastasis with increased ER expression.\textsuperscript{139}

17\textbeta-estradiol (E\textsubscript{2}) is produced from testosterone by the enzyme aromatase during normal physiological function. At the cellular level, E\textsubscript{2} elicits a wide array of nuclear and membrane signaling responses in different cells via different ERs. An assessment of estrogen’s full impact on laryngeal cancer must consider the vast number of nuclear, cytosolic, and membrane-associated ERs, the differential expression of ER in different cell types, and the convergence and crosstalk of the membrane and classical ER pathways in each cell. It is their integrative, at times synergistic or alternatively antagonistic, effects that confer the variability and complexity of E\textsubscript{2} function in both normal and cancer cells.\textsuperscript{68}
Estrogen exerts regionally specific effects on cell proliferation and mRNA expression of extracellular matrix (ECM)-associated genes in normal fibroblasts. Cervicothoracic fibroblasts from the vocal fold, trachea, and esophagus have been shown to respond differently to estrogen based on their specific estrogen profiles. Studies examining the presence of ERα in vocal cord fibroblasts show that it is localized predominantly in the nucleus and cytoplasm. Previous work by our group and others has shown that treatment with E2 suppresses extracellular matrix gene expression in laryngeal and vocal fold tissue, but through a membrane-associated estrogen receptor rather than a nuclear ER. The complexity of E2 function and its dependency on local ER expression are further emphasized in our previous study with laryngeal cancer cell lines, in which we found E2 exhibited either a protective effect inhibiting DNA synthesis, or a deleterious effect augmenting proliferation and conferring anti-apoptotic potential to the cancer cells in a manner specific to the ER profile in each cell line.

These findings stress the importance of further establishing the molecular and clinical characterization of laryngeal cancer in order to improve our understanding of the disease and its therapeutic options. The aim of our study was to further evaluate the different ER profiles that laryngeal cancer cells express and to correlate ER status with clinical prognoses. Furthermore, we implement our findings in vivo to evaluate how these cancers progress and behave with and without E2 in an animal model.
Materials and Methods

Cell Culture

UM-SCC-12 cells and UM-SCC-11A were purchased from the Cancer Research Laboratory, Department of Otolaryngology/Head and Neck Surgery at the University of Michigan. Cells were maintained as previously described. Estrogen receptor profiles were reported in a previous publication and were compared against previously established estrogen receptor profiles. The two cell lines were further characterized as described below.

Estradiol Production

Aromatase Activity.

Confluent cultures of UM-SCC-11A and UM-SCC-12 cells were harvested and assessed for aromatase activity (Aromatase (CYP19A) Activity Assay Kit (Fluorometric), Biovision, Milpitas, CA) according to manufacturer’s instructions. Monolayers were also assessed for total protein content (Pierce 660nm Protein Assay, ThermoFisher, Waltham, MA), and aromatase activity was normalized to total protein.

17β-estradiol parameter assay.

UM-SCC-11A and UM-SCC-12 cell monolayers were cultured to confluence in hormone-free phenol-red media supplemented with 10% charcoal-dextran stripped fetal bovine serum (CD-FBS). At confluence, cell supernatants were collected and assessed for total 17β-estradiol content (Estradiol parameter assay kit, RnD Systems, Minneapolis, MN). Monolayers were washed twice in 1X PBS, lysed in 0.05% Triton X 100, and assessed for total double-stranded DNA content as described below. Total supernatant 17β-estradiol was normalized to total double-stranded DNA content.
Total p53 Content

p53 is a tumor suppressor gene that can induce apoptosis. Total p53 levels were measured 24 hours post-treatment with a sandwich ELISA (Human Total p53 DuoSet® IC, R&D Systems, Minneapolis, MN). Cell monolayers were cultured to confluence in 24W plates, then treated with fresh full media containing either 0 or 10⁻⁷M E₂ for 9 minutes. These media were removed by aspiration and fresh media were added to the cultures for 24 hours. Cultures were then washed twice in 1X PBS and harvested in 200μL of p53 lysis buffer according to manufacturer’s instructions, then assayed for total protein content (ThermoFisher, Pierce 660 nm Protein Assay), and normalized as previously described⁶⁷.

DNA Quantification

 Cultures were treated with 0 or 10⁻⁷M E₂ for 9 minutes as above and harvested with p53 lysis buffer. Cultures were then assessed for total double-stranded DNA content using the QuantiFluor® dsDNA System according to manufacturer’s instructions (Promega, Madison, WI).

Production of UM-SCC-12 Cells Silenced for ESR1

Cells lines containing scramble control mRNA (TurboGFP Cat.# SHCOO4V, Millipore Sigma, Burlington, MA) and cells silenced for ESR1 (shESR1-UM-SCC-12) (Cat.# SHCLNV-NM_000125, Millipore Sigma) were created by transducing WT UM-SCC-12 cells with commercially available lentiviral shRNA plasmids and selecting the cells with 5μg/mL puromycin. Cells were maintained in full media containing 5μg/mL puromycin. Knockdown was assessed with western blots as described below.
Tumor Model

Subcutaneous xenograft laryngeal cancer mouse model.

UM-SCC-11A (n=16) and UM-SCC-12 (n=16) cell lines were mixed with 1X DPBS (Thermo Fisher Scientific, Waltham, MA) and phenol-red free Cultrex basement membrane extract (BME, Trevigen, Gaithersburg, MD) to a concentration of 7.5 mg/mL BME and 10 million cells/mL. The resulting cell suspension was kept on ice and 100μL (1 million cells) was injected subcutaneously into the left flank of a 6-week old male NOD.Cg-Prkdc<sup>scid</sup> Il2rg<sup>tm1Wjl</sup>SzJ (NSG) mouse, Cancer Mouse Models Core, Massey Cancer Center, Virginia Commonwealth University, Richmond, VA) to create a subcutaneous xenograft model of laryngeal cancer. At the time of cell injection, each animal was also subcutaneously implanted with a 0.96mg/60-day release E<sub>2</sub> pellet or a corresponding placebo pellet to create four experimental groups: UM-SCC-11A + Placebo, UM-SCC-11A + E<sub>2</sub>, UM-SCC-12 + Placebo, UM-SCC-12 + E<sub>2</sub> (n=8 for each condition). Tumors were allowed to grow for 8 weeks, and tumor measurements were taken with digital calipers starting at week 2 until the end of the study. After 8 weeks, mice were euthanized by CO<sub>2</sub> inhalation and cervical dislocation and tumors were extracted and preserved on wet ice (<6 hours) until μCT analysis. After μCT analysis, tumors were fixed in formalin and histologically analyzed as described below. All animal experiments were conducted in full compliance with the recommendations for the Care and Use of Laboratory Animals of the National Institutes of Health under a protocol approved by the Virginia Commonwealth University Institutional Animal Care and Use Committee (protocol number AD10000675).
In a second experiment, WT UM-SCC-12, and shESR1-UM-SCC-12 subcutaneous xenografts were created as described above (n=16 for each cell line). Animals were subcutaneously implanted with a 0.96mg/60-day release E₂ pellet or a corresponding placebo pellet as described above to create four experimental conditions: WT UM-SCC-12 + Placebo, WT UM-SCC-12 + E₂, shESR1-UM-SCC-12 + Placebo, shESR1-UM-SCC-12 + E₂ (n=8 for each condition). To preserve animal life, xenografts with scrambled control UM-SCC-12 cells were not created for this experiment. Tumors were monitored for 8 weeks until harvest. Animals were euthanized as described above and tumors were fixed in formalin.

μCT analysis.

After harvest, tumors were preserved for less than 6 hours on wet ice and scanned with a Bruker Skyscan 1173 μCT at 55kV and 70μA at a resolution of 560x560 pixels with an image pixel size of 40.26μm, an exposure time of 125ms and a rotation step of 0.8 degrees.¹⁶⁶ NRecon software version 1.6.10.4 (Kontich, Belgium) with a smoothing kernel of 0 and a beam hardening correction of 20% was used to reconstruct and analyze the tumors with a standard reconstruction protocol. Total tissue volume approximated total tumor volume.

Histology.

After harvest, xenograft tumors were fixed in 10% neutral formalin for 7 days, processed, and embedded in paraffin. Samples were then sectioned into 4μm thickness and stained with haemotoxylin and eosin as previously described.¹⁶⁸ Slides were imaged at 10X and 40X (Zeiss AxioVision Microscope, Carl Zeiss, Oberkuchen, Germany).
Clinical Sample Acquisition

Frozen tissue samples and prepared haematoxilin and eosin (H&E) slides were acquired from Meir Hospital, Kfar Saba, Israel under Institutional Review Board (IRB) protocol #0497-13-RMC or from Massey Cancer Center, Virginia Commonwealth University (VCU), Richmond, Virginia under an anonymization agreement in compliance with the Office for Human Research Protections’ “Guidance on Research Involving Coded Private Information or Biological Specimens”. Information on patient gender, age, tobacco and alcohol use, and tumor location (glottic/supraglottic), was provided by respective institutions. A value of '0' indicates no or infrequent use of tobacco or alcohol products as reported by the patient; a value of ‘1’ indicates self-reported regular, heavy use of tobacco or alcohol products.

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Table 4.1 Laryngeal squamous cell carcinoma samples and patient demographics
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**Table 4.2 Summary of clinical laryngeal squamous cell carcinoma samples.**

**Preparation of Lysates**

For samples obtained from Meir Hospital, Israel, tissue pieces were weighed, and >10mg of tissue was minced into pieces <1mm³ with a No.11 scalpel blade. Tissue pieces were homogenized with a Dounce tissue grinder in Nonidet p-40 (NP-40) containing 5mM NaF and 20μL of protease inhibitor cocktail (Sigma-Aldrich #P3480) per mL of NP-40. After homogenization, samples were sonicated on ice at 40 amperes for 5 seconds to obtain tissue lysates as previously described.¹⁶⁹,¹⁷⁰

For samples from the VCU Tissue and Data Acquisition and Analysis Core Laboratory (TDAAC), frozen tissue shavings were obtained and washed twice with 2 mL of 1X PBS containing 40μL of protease cocktail inhibitor. Samples were vortexed and centrifuged at 1000 rpm for 5 minutes between each wash. Tissue pellets were then resuspended in 200μL of radioimmunoprecipitation assay (RIPA) buffer and sonicated on ice at 40 amperes for 5 seconds to obtain tissue lysates.

To assess ERα66 and ERα36 knockdown in shESR1-UM-SCC-12, WT UM-SCC-12, and Scramble Control UM-SCC-12, monolayer cultures were grown in T-75 flasks.
(n=1 flask per cell line) until confluent, washed twice with 1X phosphate buffered saline (PBS), and harvested with 300μL RIPA as previously described to create cell lysates.171

Tissue and cell lysates were incubated on ice for 30 minutes and centrifuged at 13000g for 20 minutes at 4°C. Supernatants were saved and assayed for total protein content and used in western blots as described below.

**Western Blots**

50μL of tissue or cell lysate containing 8-35μg protein were loaded onto 4-20% Mini-PROTEAN®TGX™ precast polyacrylamide gels (Bio-Rad, Hercules, CA). Proteins were transferred to low-fluorescence PVDF membranes using a Trans-Blot® Turbo™Transfer System (Bio-Rad). Membranes were blocked for 1 hour at room temperature in odyssey blocking buffer (LI-COR) and then incubated with antibodies against GAPDH (mouse monoclonal, Millipore, Burlington, MA) and ERα (rabbit polyclonal, Chi Scientific, Maynard, MA) or ERβ (rabbit polyclonal, Abcam, Cambridge, United Kingdom) for 24 hours at 4°C. Membranes were then incubated with IRDye 700CW (goat anti mouse) and IRDye 800CW (goat anti-rabbit) conjugated secondary antibodies (LI-COR) for 45 minutes at room temperature and imaged using the LI-COR Odyssey® CLx Infrared Imaging System. Western blots were carried out at a central institution (VCU) to minimize variation and repeated once (a total of two experiments per assay) with 1 replicate per sample in each experiment. Signal intensity for proteins of interest (ERα66, ERα36, and ERβ) was normalized to GAPDH. Where no signal was detected, samples were considered to have ‘0’ signal intensity and marked with ‘N.D.’, or no signal detected, on graphs (Figure 4.1, Figure 4.2).
Normalized signal intensity for ERα66, ERα36, and ERβ in each tumor sample was grouped by stage (stage 1, stage 2 & 3, stage 4) and graphed (Figure 4.3A-4C); or normalized to corresponding signal intensities in adjacent epithelial tissue (Figure 4.3D). A normalized ratio of greater than 1 was recorded with a blue upward-pointing arrow ↑, and a ratio less than 1 was recorded with a red downward-pointing arrow ↓ (Table 4.3). To generate heat maps, log_{10} of normalized signal intensities of tumor (Figure 4.3E) or normal epithelial tissue (Figure 4.3F) was calculated and graphed using the Graphpad Prism 7 Heatmap generator. Samples were grouped by stage and arranged in order of increasing cancer aggression (top-bottom) (Figure 4.3E-F).

**Figure 4.1 Expression of ERα in individual LSCC samples.**

Quantification of western blots for [A] ERα66, [B] ERα36, and [C] ERβ for individual clinical tumor samples. Y-axes are given as log scales. Samples are color-coded according to clinical stage. Stage 1 samples are green, stage 2 yellow, stage 3 orange, and stage 4 red.
Figure 4.2 Expression of ERα in individual normal laryngeal tissue samples (LSCC surround).

Quantification of western blots for [A] ERα66, [B] ERα36, and [C] ERβ for normal epithelia adjacent to clinical tumor samples. Y-axes are given as log scales. Samples are color-coded according to clinical stage. Stage 1 samples are green, stage 2 yellow, stage 3 orange, and stage 4 red.
Figure 4.3 Expression of ERα isoforms in stage 1-4 clinical laryngeal cancer.

Quantification of western blots of [A] ERα66, [B] ERα36, and [C] ERβ. Samples were grouped by cancer stage (stage 1, stages 2 and 3, stage 4) and analyzed by one-way ANOVA with Tukey’s correction. Bars not sharing a letter are significantly different from each other with P < 0.05. Bars are shown as total protein expression, with solid bars representing protein expression in normal epithelia and patterned bars representing protein expression in tumor tissue. Y-axes are given as log scales. [D] Protein expression in LSCC samples was normalized to expression in surrounding epithelia, grouped by stage, and graphed. * indicates significance against stage 1 within protein groups. Panels E and F: Heat maps showing [E] tumor and [F] surrounding epithelia expression of ERα66, ERα36, and ERβ for each sample. Individual protein expression was measured by western blot and graphed as log10 of normalized signal intensity.
Modulation of 24R,25(OH)2D3 Tumorigenicity by ERα Isoforms in E2-Responsive Cancer • Spring 2019

Table 4.3 Estrogen receptor protein expression in clinical laryngeal squamous cell carcinoma samples.

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<td>invasive keratinizing SCC moderate to poorly diff</td>
<td>4</td>
<td>2b</td>
<td>x</td>
<td>4</td>
<td>↓ ↑</td>
</tr>
<tr>
<td>11</td>
<td>invasive keratinizing SCC moderate diff</td>
<td>4</td>
<td>2c</td>
<td>x</td>
<td>4</td>
<td>↓ ↑</td>
</tr>
<tr>
<td>12</td>
<td>invasive keratinizing SCC moderate diff</td>
<td>4</td>
<td>1</td>
<td>0</td>
<td>4</td>
<td>↓ ↓</td>
</tr>
<tr>
<td>13</td>
<td>invasive keratinizing SCC moderate to poorly diff</td>
<td>4</td>
<td>2</td>
<td>0</td>
<td>4</td>
<td>↓ ↓</td>
</tr>
<tr>
<td>14</td>
<td>invasive keratinizing SCC, NOS</td>
<td>4</td>
<td>0</td>
<td>0</td>
<td>4</td>
<td>↓ ↓</td>
</tr>
</tbody>
</table>

Ratio | Symbol
----|------
>1 | ↑
<1 | ↓

Ratio symbol: >1 for increase, <1 for decrease.
Accessing ESR1 and ESR2 Gene Transcript Expression Data

ESR1 and ESR2 expression data were publically available and were obtained from Total Cancer Genome Atlas (TCGA) cohort (Project Id: TCGA-HNSC) (n=525 patients) (https://portal.gdc.cancer.gov/projects/TCGA-HNSC) using University of California Santa Cruz (UCSC) Xena (http://xena.ucsc.edu/). In brief, ESR1 and ESR2 expression data were downloaded from Xena and only samples taken from primary solid head and neck squamous cell carcinomas were included in overall head and neck cancer analyses. Samples were further separated by neoplasm tissue type, and only primary LSCC samples were included in the second analysis. Overall survival (OS) was defined as time of death by any cause and capped at fifteen years. Kaplan-Meir curves of OS were generated by GraphPad Prism v5.0 (GraphPad Software Inc.). Patients were grouped according to median gene expression as described previously.

Statistical Analysis

µCT tumor volumes were compared with one-way ANOVA analysis with Tukey’s correction. Tumor volume measurements over time were analyzed with a repeated-measures ANOVA. Ex vivo western blots were grouped by stage and analyzed with one-way ANOVA analysis with Tukey’s correction to compare between stages and two-way ANOVA analysis with Bonferroni post-tests between stages to determine significance between precancerous and tumor tissue. Tumor protein expression for ERα66, ERα36, and ERβ was normalized to matched precancerous tissue and graphed as treatment/control. Two-way ANOVA analysis with Bonferroni post-tests between stages was done to determine significance between stages. Statistical analysis was performed by using GraphPad Prism v5.0. Comparison of high and low ESR survival curves was
done with a Log-rank Mantel-Cox χ² test. For all studies, p-values < 0.05 were considered significant.

Results

**UM-SCC-12 and UM-SCC-11A locally produce 17β-estradiol.**

Both ER positive (ER+) UM-SCC-12 cells, which express ERα66 and ERα36, and ER negative (ER-) UM-SCC-11A cells, which do not express ERα66 and only express ERα36, produce similar concentrations (~250 pg/mL supernatant) of 17β-estradiol (Figure 4.4A). However, ER- UM-SCC-11A produced ~1.5 times more 17β-estradiol per cell than their ER+ UM-SCC-12 counterparts (Figure 4.4B). Similarly, UM-SCC-11A cells had higher basal aromatase activity than UM-SCC-12 cells (Figure 4.4C), with approximately 4nU of enzyme per mg of protein compared to UM-SCC-12’s 1nU/mg protein.

**17β-estradiol increased tumor aggression in estrogen receptor positive laryngeal cancer, but not estrogen receptor negative cancer in vivo.**

ER positive UM-SCC-12 xenografts grown in placebo-treated mice initially increased in volume (slope ≠ 0, Table 4.4), but did not increase in size after 4 weeks of growth (slope ≈ 0, Table 4.5) and ultimately reached a volume of 250-500 mm³ (Figure 4.5A,C). In contrast, UM-SCC-12 xenografts grown in E₂-treated mice increased in size over time (slope ≠ 0, Table 4.4, Table 4.5) and attained a final volume of 500-1000 mm³, 2-fold larger than tumors grown in control-treated mice (Figure 4.5A,C). Most of this increase was observed after week 4. This effect was not observed in ER negative UM-SCC-11A
tumors, where both control and E\textsubscript{2} treated tumors increased in size at a similar rate and to a similar final volume (Figure 4.5B,C). Histology of UM-SCC-12 tumors revealed even cell shape, chromatin distribution, and uniform morphology in control tumors (Figure 4.5D), similar to stage 1 LSCC (Figure 4.6A,E). E\textsubscript{2} treated tumors stained with eosin and haematoxylin (H&E) had similar eosin staining but darker haematoxylin staining, more irregular nuclei, and uneven chromatin distribution as compared to control treated tumors (Figure 4.5E), similar to stage 2 LSCC (Figure 4.6B,F).

![Table image](image.png)

**Table 4.4 Xenograft models of LSCC tumor growth curve: linear fit, weeks 2-8.**
Table 4.5 Xenograft models of LSCC tumor growth curve: linear fit, weeks 4-8.

<table>
<thead>
<tr>
<th>Best-fit values</th>
<th>11A Control</th>
<th>11A 17b-Estradiol</th>
<th>12 Control</th>
<th>12 17b-Estradiol</th>
</tr>
</thead>
<tbody>
<tr>
<td>Slope</td>
<td>441.2 ± 69.19</td>
<td>684.9 ± 79.54</td>
<td>19.09 ± 22.63</td>
<td>209.0 ± 37.80</td>
</tr>
<tr>
<td>Y-intercept when X=0.0</td>
<td>-1728 ± 443.0</td>
<td>-2958 ± 503.3</td>
<td>343.3 ± 140.8</td>
<td>-638.4 ± 234.5</td>
</tr>
<tr>
<td>X-intercept when Y=0.0</td>
<td>3.917</td>
<td>4.318</td>
<td>-17.99</td>
<td>3.055</td>
</tr>
<tr>
<td>1/slope</td>
<td>0.002267</td>
<td>0.001460</td>
<td>0.05239</td>
<td>0.004785</td>
</tr>
<tr>
<td>95% Confidence Intervals</td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Slope</td>
<td>296.9 to 585.5</td>
<td>521.4 to 848.4</td>
<td>-26.93 to 65.11</td>
<td>132.7 to 285.3</td>
</tr>
<tr>
<td>Y-intercept when X=0.0</td>
<td>-2653 to -804.3</td>
<td>-3992 to -1923</td>
<td>56.95 to 629.7</td>
<td>-1112 to -164.9</td>
</tr>
<tr>
<td>X-intercept when Y=0.0</td>
<td>2.664 to 4.606</td>
<td>3.627 to 4.784</td>
<td>-infinity to -0.8896</td>
<td>1.221 to 3.969</td>
</tr>
<tr>
<td>Goodness of Fit</td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>R-square</td>
<td>0.6703</td>
<td>0.7404</td>
<td>0.02050</td>
<td>0.4213</td>
</tr>
<tr>
<td>Sy.x</td>
<td>417.2</td>
<td>574.1</td>
<td>194.5</td>
<td>350.3</td>
</tr>
<tr>
<td>Is slope significantly non-zero?</td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>F</td>
<td>40.66</td>
<td>74.15</td>
<td>0.7115</td>
<td>30.57</td>
</tr>
<tr>
<td>DFn, DFd</td>
<td>1.000, 20.00</td>
<td>1.000, 26.00</td>
<td>1.000, 34.00</td>
<td>1.000, 42.00</td>
</tr>
<tr>
<td>P value</td>
<td>&lt; 0.0001</td>
<td>&lt; 0.0001</td>
<td>0.4048</td>
<td>&lt; 0.0001</td>
</tr>
<tr>
<td>Deviation from zero?</td>
<td>Significant</td>
<td>Significant</td>
<td>Not Significant</td>
<td>Significant</td>
</tr>
<tr>
<td>Data</td>
<td></td>
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<td></td>
<td></td>
</tr>
<tr>
<td>Number of X values</td>
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<td>5</td>
<td>5</td>
<td>5</td>
</tr>
<tr>
<td>Maximum number of Y replicates</td>
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<td>7</td>
<td>8</td>
<td>9</td>
</tr>
<tr>
<td>Total number of values</td>
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<td>28</td>
<td>36</td>
<td>44</td>
</tr>
<tr>
<td>Number of missing values</td>
<td>28</td>
<td>22</td>
<td>14</td>
<td>6</td>
</tr>
</tbody>
</table>

Figure 4.4 E2 is locally produced in vitro by UM-SCC-12 and UM-SCC-11A cultures.

Production of 17β-estradiol by ER+ UM-SCC-12 and ER- UM-SCC-11A [A] overall and [B] per cell. [C] Basal aromatase activity in laryngeal cancer cell lines. P-values less than 0.05 were considered significant. *indicates significance against UM-SCC-12.
Figure 4.5 E2 increases tumor burden in UM-SCC-12, but not UM-SCC-11A xenografts.

[A,B] Effect of 17β-estradiol on tumor growth of ER+ and ER- xenografts in vivo. Tumor growth of [A] UM-SCC-12 (ER+) and [B] UM-SCC-11A over time with and without estradiol supplementation. * indicates significance against week-matched control tumor volume. [C] Final tumor volume was measured with μCT Bars not sharing a letter have significantly different volumes with P < 0.05. [D-F] Representative histology of [D,E] ER+ and [F,G] ER- tumors [E,G] with and [D,F] without 17β-estradiol stained with hematoxylin and eosin and imaged at 40X. Scale bars = 20μm.
Estrogen receptor negative laryngeal cancer is more aggressive than estrogen receptor positive cancer.

UM-SCC-11A subcutaneous xenografts implanted into mice grew ~250 times their original size to a final volume of 1000-2000 mm³ (Figure 4.5B). UM-SCC-11A tumors attained a final volume 2-4 times greater than that observed in UM-SCC-12 xenografts (Figure 4.5C). UM-SCC-11A also grew at a continuous exponential rate throughout the study (R²=0.77-0.88, Table 4.6) with a faster doubling time than comparable UM-SCC-12 tumors (Table 4.6). Histology on UM-SCC-12 xenografts revealed moderately differentiated tumors with defined edges and uniform cell shapes (Figure 4.5D-E). Conversely, histology of UM-SCC-11A xenografts revealed anaplastic tumors with high variability in nucleus and cell size within the tumor. These tumors also showed a significant amount of invasion into the surrounding tissue (Figure 4.5F-G). The limited invasion and defined tumor edges observed in UM-SCC-12 cells are reminiscent of the defined tumor edges and even cell morphology observed in Stage 1 and 2 LSCC (Figure 4.6A,B,E,F). The increase in invasion in UM-SCC-11A tumors as compared to UM-SCC-12 tumors is similar to the increase in tumor invasion observed in samples of stages 3 and 4 (Figure 4.6C,D). The similar eosin staining, darker haematoxylin staining, irregular cell shape, and irregular nuclei structure observed in UM-SCC-11A tumors are similar to the cell morphology observed in stage 3 and 4 clinical LSCC samples (Figure 4.6G,H).
Figure 4.6 Representative Histology of Clinical Samples of Laryngeal Cancer.


<table>
<thead>
<tr>
<th>Exponential growth equation</th>
<th>11A Control</th>
<th>11A 17b-Estradiol</th>
<th>12 Control</th>
<th>12 17b-Estradiol</th>
</tr>
</thead>
<tbody>
<tr>
<td>Best-fit values</td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Y0</td>
<td>44.95</td>
<td>15.50</td>
<td>163.4</td>
<td>54.62</td>
</tr>
<tr>
<td>k</td>
<td>0.4723</td>
<td>0.6563</td>
<td>0.1533</td>
<td>0.3782</td>
</tr>
<tr>
<td>Tau</td>
<td>2.117</td>
<td>1.524</td>
<td>6.524</td>
<td>2.644</td>
</tr>
<tr>
<td>Doubling Time</td>
<td>1.468</td>
<td>1.056</td>
<td>4.522</td>
<td>1.833</td>
</tr>
<tr>
<td>Std. Error</td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Y0</td>
<td>24.49</td>
<td>7.819</td>
<td>38.50</td>
<td>21.42</td>
</tr>
<tr>
<td>k</td>
<td>0.07290</td>
<td>0.06586</td>
<td>0.03665</td>
<td>0.05400</td>
</tr>
<tr>
<td>95% Confidence Intervals</td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Y0</td>
<td>-5.601 to 95.49</td>
<td>-0.3556 to 31.35</td>
<td>86.01 to 240.8</td>
<td>11.77 to 97.48</td>
</tr>
<tr>
<td>k</td>
<td>0.3218 to 0.6227</td>
<td>0.5228 to 0.7898</td>
<td>0.07960 to 0.2270</td>
<td>0.2701 to 0.4862</td>
</tr>
<tr>
<td>Tau</td>
<td>1.606 to 3.106</td>
<td>1.266 to 1.913</td>
<td>4.406 to 12.56</td>
<td>2.057 to 3.702</td>
</tr>
<tr>
<td>Doubling Time</td>
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<td>0.9776 to 1.326</td>
<td>3.054 to 8.705</td>
<td>1.426 to 2.566</td>
</tr>
<tr>
<td>Goodness of Fit</td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Degrees of Freedom</td>
<td>24</td>
<td>37</td>
<td>50</td>
<td>59</td>
</tr>
<tr>
<td>R square</td>
<td>0.7679</td>
<td>0.8778</td>
<td>0.3106</td>
<td>0.5898</td>
</tr>
<tr>
<td>Absolute Sum of Squares</td>
<td>3.168e+006</td>
<td>5.376e+006</td>
<td>1.738e+006</td>
<td>5.025e+006</td>
</tr>
<tr>
<td>Syx</td>
<td>363.3</td>
<td>381.2</td>
<td>186.5</td>
<td>291.8</td>
</tr>
<tr>
<td>Number of points</td>
<td>26</td>
<td>39</td>
<td>52</td>
<td>61</td>
</tr>
</tbody>
</table>

Table 4.6 Xenograft models of LSCC tumor growth curve: exponential fit, weeks 2-8.
Estrogen receptor expression in clinical cases of laryngeal squamous cell carcinoma is highly variable and regionally localized.

Quantification of western blots showed that ERα66 (Figure 4.1A, Figure 4.2A), ERα36 (Figure 4.1B, Figure 4.2B), and ERβ (Figure 4.1C, Figure 4.2C) signal intensity varied exponentially from samples to sample. While all samples taken from stage 4 patients had similar total levels of ERα66 (Figure 4.1A, Figure 4.2A) regardless of locality, ERα36 and ERβ expression was highly variable and appeared to be regionally localized. Stage 4 samples collected at Meir Hospital, Israel had greater ERα36 staining (Figure 4.1B, Figure 4.2B) and ERβ staining (Figure 4.1C, Figure 4.2C) than samples collected at Massey Cancer Center, U.S.A. All samples collected in Israel expressed high levels of ERβ, but none of the samples collected in the U.S.A. expressed ERβ (Figure 4.1C, Figure 4.2C). This regional variability was also apparent when considering the ratio of ER expression in tumor vs. precancerous tissue. All stage 3 and 4 samples collected in the U.S.A. had higher ratios of ERα36 in tumor tissue vs precancerous tissue (Table 4.3). However, stage 4 samples collected in Israel expressed less ERα36 in tumor tissue as compared to precancerous tissue (Table 4.3). If neither tumor nor matched normal tissue expressed protein for ERα66, ERα36, or ERβ as measured by western blot, a value of ‘N/A’ was recorded for that sample (Table 4.3).

Decreased absolute and normalized ERα66 expression is associated with advanced cancer stage in clinical samples of laryngeal squamous cell carcinoma.

A general trend of towards lower ERα66 expression was observed in more aggressive tumors (Stage 2-4) as compared to indolent tumors (Stage 1) (Figure 4.3A,E).
The total amount of ERα66 in these samples was greatest in stage 1 cancer (Figure 4.3A), and the proportion of ERα66 expressed by the tumor tissue as compared to matched precancerous tissue was also greater in stage 1 samples as compared to stage 4 samples (Figure 4.3D-F, Table 4.3). Samples from patients with stage 2-3 cancer had less ERα66 overall as compared to stage 1 samples (Figure 4.3A) but, like stage 1 samples, tumor tissues expressed more ERα66 than matched precancerous tissue (Figure 4.3D, Table 4.3). This trend did not hold true for Stage 4 cancer samples, where the ratio of ERα66 expression to precancerous tissue was flipped and precancerous tissue expressed more ERα66 than matched tumor tissue samples (Figure 4.3A,D-F; Table 4.3).

Patterns of relative ER expression among cancer stages described for ERα66 above also held true for ERα36 and ERβ (Figure 4.3B,C). Tumor tissue taken from patients with stage 4 cancer generally expressed less ERα36, and ERβ than matched precancerous samples (Figure 4.3B-F, Table 4.3). The ratio of ER expression in tumor vs. precancerous tissue was significantly lower in stage 4 samples as compared to stage 1 samples for all ERs: ERα66, ERα36, and ERβ (Figure 4.3D, Table 4.3).

Unlike ERα66, absolute expression of ERα36 and ERβ was similar in stage 1 and stage 4 cancers (Figure 4.3B,C). The most aggressive cancers (stage 4) generally had low ERα66 expression and high ERα36 expression (Figure 4.1A-B). Absolute ERα36 expression in stage 2 and 3 cancer samples was lower than stage 1, but ERα36 expression generally increased with stage in tumor samples taken from stage 2-4 patients (Figure 4.1B). Normal tissue surrounding stage 4 samples expressed higher ERα36 than matched normal tissue from lower stage cancers. Neither tumor nor precancerous samples taken from stage 2 and 3 samples patients expressed ERβ. However, it is
important to note that all stage 2 and 3 samples were collected in the U.S.A., while stage 4 samples were collected in both the U.S.A. and Israel.

Higher expression of ESR1 but not ESR2 is associated with increased survival in patients with primary laryngeal squamous cell carcinoma.

Analysis of RNAseq data taken from the TCGA study of 525 patients with primary head and neck cancer showed that patients with higher than median tumor ESR1 expression (ESR1 > 5.56) had significantly better survival rates than patients with lower tumor ESR1 expression (Figure 4.7A). This trend was also observed when patients were stratified by tumor ESR2 expression, with patients with higher than median ESR2 expression (ESR2 > 3.69) (Figure 4.7B) having significantly higher survival rates than patients with lower ESR2 expression after 15 years.

RNAseq data taken from a TCGA study of 128 patients with primary LSCC showed that patients with higher than median tumor ESR1 expression (ESR1 > 5.56) had significantly higher survival rates as compared to patients with lower tumor ESR1 expression (Figure 4.7C) with p-value < 0.0001 and a hazard ratio of 0.02845. However, no correlation between ESR2 expression and survival was observed when patients were stratified into groups of higher and lower than median ESR2 expression (Figure 4.7D) (p-value = 0.83).

Silencing ESR1 in estrogen receptor positive laryngeal squamous cell carcinoma eliminates the 17β-estradiol response and increases aggression.

Silencing the ESR1 gene, which encodes all known ERα isoforms, in UM-SCC-12 cells reduces ERα66 expression by 76% as compared to a 33.5% knockdown in scramble
control cells (Figure 4.8A,B). Similarly ESR1 silencing knocked down ERα36 expression by 75.2% as compared to 44.4% knockdown in scramble control cells (Figure 4.8A,C).

Silencing ESR1 in UM-SCC-12 did not change cell number in vitro as measured by total double-stranded DNA content (Figure 4.9A). However, silencing ESR1 altered total p53 content in these cells (Figure 4.9B). p53 protein production is associated with apoptosis. Total p53 content was significantly lower for shESR1-UM-SCC-12 cells as compared to WT and scramble control UM-SCC-12 cultures. Furthermore, silencing ESR1 in UM-SCC-12 eliminated the response of these cells to E2. Both vehicle and E2-treated shESR1-UM-SCC-12 cultures had approximately 1/3 the p53 content of WT vehicle-treated UM-SCC-12, whereas both vehicle and E2-treated shESR1-UM-SCC-12 cultures levels of p53 similar to those observed in WT UM-SCC-12 cells treated with E2 (Figure 4.9B).

UM-SCC-12 subcutaneous xenografts silenced for the ESR1 gene grew approximately 4 times larger than WT UM-SCC-12 xenograft tumors and twice as large as WT UM-SCC-12 xenografts treated with E2. shESR1-UM-SCC-12 xenografts grew to similar sizes regardless of E2 supplementation. Both silenced and WT xenografts grew in an approximately linear fashion, suggesting a faster doubling time in vivo for shESR1-UM-SCC-12 cells as compared to WT UM-SCC-12.
Figure 4.7 Intra-tumoral ESR1 expression correlates with survival.

Kaplan-Meier survival curves for patients with any [A,B] primary head and neck squamous cell carcinoma or [C,D] primary LSCC stratified by higher or lower than median [A,C] ESR1 or [B,D] ESR2 expression. Survival curves were analyzed with Log-rank Mantel-Cox χ² tests. P-values less than 0.05 were considered significant.
Figure 4.8 Knockdown of ERα in UM-SCC-12 cells.

A representative [A] western blot showing ERα expression in WT, scramble-control, and ESR1-silenced UM-SCC-12 cell lines. GAPDH was used as a protein loading control. [B] ERα66 and [C] ERα36 protein expression were normalized to GAPDH and quantified by normalized signal intensity. N=1.
Figure 4.9 Silencing ESR1 in ERα+ LSCC increases tumor aggression.

Effect of silencing ESR1 in ERα66+ UM-SCC-12 cells *in vitro* on [A] cell number and the response to estrogen as measured by [B] total p53. Bars that do not share a letter are considered significant with p-values less than 0.05. *In vivo*, WT and ESR1-silenced UM-SCC-12 cell-line xenografts were implanted in a subcutaneous xenograft mouse model and [C] tumor burden was measured over time. Black * indicates significance against week-matched WT-vehicle tumor volume. Red squares ■ indicate significance against week-matched WT-E2 tumor volumes. P-values less than 0.05 were considered significant.
Discussion

Accumulated evidence has substantiated that laryngeal cancer, a common head and neck cancer in the United States, is a hormone responsive cancer, comparable to other more renowned secondary sex hormone cancers. Despite the reports of E₂ detrimental effects in laryngeal cancer⁸⁸,⁹⁰ and that anti-estrogen treatment has a beneficial effect⁶⁴ originating almost three decades ago,⁶² there has been little advance in translating this recognition of the importance of E₂ to practical clinical implications. This might be explained by the cofounding and heterogeneous ER profile detected in these cancer cells. The cumulative effects of this heterogeneity translate to disparate responses to E₂ and must be clarified before implementation to clinical practice.

Local production of E₂ has been described in many steroid hormone responsive cancers, including breast,¹⁷⁵ endometrium,¹⁷⁶ cervical,¹⁷⁷ and testicular cancer.¹⁷⁸ Here, we observed a saturation in local E₂ production that occurred around 250 pg/mL, roughly 20 times previously reported levels of estradiol in serum from healthy adult males,¹⁷⁹ but not dissimilar from E₂ levels reported in plasma from pre-menopausal breast cancer patients.¹⁸⁰ Similar levels of E₂ production have also been observed in breast cancer-associated fibroblasts.⁸² The tumorigenic properties of estrogen have been well-described in breast cancer,⁵⁵,⁷⁶ and our previous work has shown that E₂ is also tumorigenic in ER+, but not in ER-, laryngeal cancer.⁶⁶,⁶⁷ The increase in aromatase activity and corresponding increase in E₂ production per cell in ER- LSCC vs. ER+ LSCC was surprising; however similar disparities in E₂ production have been observed in ER+ and ER- breast cancer.¹⁸¹ Increased serum E₂ is associated with compensatory mechanisms that arise in conjunction with defective estrogen signaling in normal breast
tissue, and elevated serum E\textsubscript{2} has also been reported in estrogen insensitive triple-negative breast cancer (TNBC) patients as compared to those with ER\textsuperscript{+} tumors. It is possible that the increase in aromatase activity and subsequent increase in E\textsubscript{2} production per cell was due to a saturation in E\textsubscript{2} concentration reminiscent of a classic negative signaling feedback loop. However, it is also possible that the elevated E\textsubscript{2} production and aromatase activity observed in ER\textsuperscript{-} LSCC is a result of similar compensatory signaling associated with these cells’ insensitivity to estradiol.

Consistent with previous reports, evaluation of the laryngeal epithelia adjacent to laryngeal cancer reveals expression of both classical nuclear ERs and membrane ERs, specifically the ER\textalpha isoform ER\textalpha36. ER\textalpha36 is a well-established ER\textalpha splice variant which can mediate the effects of E\textsubscript{2} independent of nuclear ER\textalpha. This effect has been demonstrated in ER\textalpha66 null cells and in studies using antibodies against the nuclear and membrane isoforms of ER\textalpha. The receptor initiates divergent pathways from the plasma membrane that re-converge downstream to affect cancer cell survivability. In our study a striking difference is evident between the epithelia adjacent to advanced aggressive tumors and to more indolent tumors. While ER\textbeta expression is highly variable, both ER\textalpha variants, the classical nuclear ER\textalpha66 and ER\textalpha36, are expressed in high levels in the epithelia adjacent to tumors displaying aggressive behavior, while absent in the indolent samples. In this, the ER\textalpha expression patterns in the epithelia adjacent to aggressively malignant samples (stage 4) is reminiscent of the ER\textalpha66 and ER\textalpha36 expression profiles generally seen in more indolent tumor samples.

It is well known that laryngeal squamous cell carcinoma arises from precancerous lesions, mainly dysplastic lesions. However, the exact molecular mechanisms of
malignant transformation of laryngeal mucosa are not clear. Moreover, the ability to identify patients who are most likely to progress into invasive laryngeal cancer or possess warning aggressive markers early in the course of their disease is paramount to early diagnosis and treatment to confer better prognoses. The presence of markers that correlate with pre-malignancy or early-stage cancer could have cardinal bearing on clinical decisions to observe or aggressively treat lesions.

Various classification systems have been crafted in an attempt to describe the histologic features of these laryngeal epithelial precursor lesions. Unfortunately, a universally accepted histopathological classification system and consensus on diagnostic criteria for LSCC are lacking. Different diagnostic techniques, specifically imaging modalities of laryngeal epithelial lesions, have been developed, but they do not offer a single system to make a differential diagnosis. Consequently, new markers are required to reliably identify those high-risk precancerous lesions. Effort has been directed to identify molecular biomarkers that will enable the stratification of risk for malignant transformation and aggressive tumor behavior in patients with precancerous lesions. Some proposed biomarkers include the presence of chromosome instability markers, cell cycle proteins, β-catenin, and microRNA-21.

To our knowledge, ER characterization and profiling has not been studied in the context of precancerous lesions. Our findings imply that laryngeal epithelia expressing high levels of ERα66 and ERα36 are at risk for aggressive malignant transformation. Thus patients expressing this phenotype should be considered for more aggressive treatment and followed closely. Our sample number is too small to draw conclusions; however, in
this evolving field, ER should be considered as another biomarker that can shed light on the malignant potential of the tumor and direct treatment.

Evaluation of advanced and aggressive tumors, with clinical stage 4, revealed recovered high rates of ERα36, while remarkably losing or reducing their expression of ERα66. Indolent tumors, clinical stage 1, unexpectedly were found to upregulate the expression of both ERα66 and ERα36 compared to the adjacent epithelia. The majority of stage 2 and 3 tumors have reduced ERα66 expression as compared to more indolent samples, but retain their overexpression of ERα66 as compared to adjacent tissue. Conversely, there is a general trend of increasing ERα36 expression in more aggressive samples, suggesting that it is only in the final most aggressive stage 4 that tumors begin to loose ERα66 and regain ERα36 expression. In an attempt to interpret the differentiate ER expression between more indolent and aggressive tumors we turn to the prototype by which intricate ER cellular functions have been extensively studied, breast cancer.

The presence of the classical nuclear ERα66 has long been the most important molecular marker used in the diagnosis and as a predictor of prognosis in breast cancer. The presence of the receptor has a substantial role in dictating treatment, making anti-estrogen treatment a viable option for those patients with ERα overexpressing tumors. However, eventually most breast tumors become resistant to anti-estrogen treatment either through de-novo or acquired resistance. One prevailing theory for the resistance to anti-estrogen treatment is the loss of the expression of ERα66, either as the presenting phenotype in ER negative tumors or in ER positive tumors during treatment, imparting loss of constraints and deleterious effects on tumor behavior. ER negative breast cancer, specifically the subclass of triple negative cancers, are renowned for their aggressive
nature and grave malignant potential\textsuperscript{191–193}. Thus, in the case of the most renowned hormone responsive cancer, it is the loss of that responsiveness that causes the most aggressive behavior.

Interestingly, ER positive breast cancer acquired loss of the expression of ER precipitates the same aggressive phenotype. That same loss of ER expression has been described to incur trans-differentiation from epithelial to mesenchymal phenotype, which is responsible for increased aggressiveness and metastatic propensity\textsuperscript{194}. More than a third of patients with recurrence and metastases from primary ER positive breast cancer were found to have lost their expression of ER upon recurrence\textsuperscript{195,196} which incurred a worse prognosis\textsuperscript{197}. Coupled with the fact that almost half of patients with aggressive breast cancer characterized with high rates of recurrence and distant metastasis are classified as bearing primary triple negative cancers\textsuperscript{198}, it seems evident that the lack of ER expression confers poor prognosis.

That loss of ER confers a poorer prognosis was further evidenced in our animal model. While the subcutaneous xenograft model we used does not traditionally metastasize\textsuperscript{199–201} the increased aggression associated with low ER and E\textsubscript{2} sensitivity in breast cancer was observed, with ER- LSCC reaching final tumor volumes more than twice as large as their ER+ LSCC counterparts regardless of estrogen supplementation. This was further borne out in a second LSCC xenograft model, which showed that natively ER+ LSCC silenced for the ESR1 gene behaves like ER- LSCC \textit{in vivo}. shESR1-UM-SCC-12 xenografts grew to larger tumor sizes than either WT or E\textsubscript{2}-treated ER+ LSCC xenografts regardless of estrogen supplementation.
In vitro studies confirm this, with silencing ESR1 in ER+ LSCC reducing total p53 levels to those on par with E2-treated ER+ LSCC. This suggests that ERα loss mimics the effects of E2 treatment on ER+ LSCC, implying that the presence of ERα, independent of its role as an estrogen receptor, affects some anti-tumorigenic signaling, which is superseded in the presence of supra-physiological doses of E2. E2-ERα signaling has been well established as pro-tumorigenic in breast cancer, but the role of ERα independent of its actions as an E2 conduit has not been widely studied. However, other studies in breast and other hormonally response cancers have shown that loss of ER contributes to tumor aggression.

Although ER expression profiles were not confirmed in the harvested xenograft, expression of ERα and ERβ isoforms pre-implantation suggests that both UM-SCC-12 and UM-SCC-11A have similar levels of ERα36 expression, but only UM-SCC-12 express ERα66. This is in line with observations from clinical and TCGA data, which suggest that increased ERα is associated with lower aggression in LSCC. It should also be noted that silencing ESR1 in UM-SCC-12 reduces both ERα66 and ERα36 expression in these cells, making it difficult to determine if the increase in aggression in shESR1-UM-SCC-12 is due to a reduction in ERα66, ERα36, or some combination of the two. The similar levels of ERα36 in UM-SCC-11A and UM-SCC-12 suggest that the increased aggression observed in shESR1-UM-SCC-12 is due to a reduction in ERα66 expression, but further studies are needed to determine which isoform(s) of ERα are involved in ERα-silencing induced increases in tumor aggression.

Survival data from head and neck cancer and LSCC patients further suggest that lack of ERα is associated with poorer prognoses, again in a manner similar to breast
cancer. It is well known that loss of ER correlates with reduced survival in breast cancer patients, and similar trends were observed in our meta-analysis of recent head and neck cancer cohorts. Although all head and neck cancer patients show a slight correlation between increased ERα or ERβ expression and improved survival odds; this trend is not mirrored in laryngeal cancer, where ERα is the predominant determinate of ER-associated survival. It should be noted that the majority of the samples in this data set came from patients with stage 4 cancer (n = 81 of 140), suggesting that the prognostic value of ESR1 expression as a marker of LSCC aggression may exist independent of any association with late-stage cancer. It is also interesting to note the discrepancy between the whole head and neck cancer dataset and laryngeal cancer survival specifically. RNAseq data necessarily lacks information about post-transcriptional modification of transcribed proteins and can offer limited insights about splice variants and alternative isoforms of the traditional ERα and ERβ, many of which have been identified.

A lack of correlation between ESR1 and ESR2 expression may indicate a mechanistic difference in the ERα and ERβ membrane and cytosolic signaling pathways that could be dependent on alternative isoforms of both ERs.

The mechanism underlying the loss of ER expression in primary ER positive tumors has not yet been clarified. Genomic and posttranscriptional silencing mechanisms offer an explanation for the loss of ER expression. One proposed mechanism is the inactivation of ER gene transcription due to methylation of cytosine-rich areas termed CpG islands. This mechanism has also been described in another hormone responsive tumor, endometrial cancer. Similar silencing of ER gene transcription is the basis of the mechanism underlying the action of micro-RNA. The micro-RNA are a class of regulatory
molecules that have been shown to control gene expression such as ERα in breast cancer,\textsuperscript{207,208} and have been implicated in ovarian,\textsuperscript{209} endometrial\textsuperscript{210} and laryngeal cancer.\textsuperscript{211} An alternative genomic mechanism has been proposed, that entails the loss of heterogeneity in ER genes such as allelic loss in microsatellites located in regulatory regions of ER genes,\textsuperscript{212} which was verified as an independent prognostic factor for relapse free survival.\textsuperscript{213}

These mechanisms of loss of ER expression resemble the findings we have observed in laryngeal cancer. The indolent laryngeal cancer cells express both ERα66 and ERα36, and their ER profile resembles that of the epithelia adjacent the aggressive tumors, being part of a spectrum from precancerous lesions to low grade tumors. Stage 2 and 3 samples begin to lose ERα66 and gain ERα36 expression, and, the aggressive tumors are distinctly characterized by the loss of ERα66 and high expressing ERα36.

There is growing interest in the clinical implication of the membrane ERα36, particularly in regard to its role in mediating E\textsubscript{2} responsiveness in ER negative tumors and anti-estrogen resistance in ER positive tumors. About 40% of ER positive and ER negative breast cancer cells express ERα36 in the plasma membrane.\textsuperscript{214} High levels of ERα36 expression correlate with an unfavorable prognosis, independent of ER status and have been suggested as an important novel marker for breast cancer clinical characterization.\textsuperscript{88} In our previous studies and in this recent study we have consistently identified high rates of ERα36 expression in laryngeal cancer cells.\textsuperscript{66,67} Furthermore, in a previous study, we demonstrated that activation of ERα36 resulted in an upregulation of angiogenic and metastatic factors and observed an association between the amount of ERα36, VEGF, and lymph node metastasis in laryngeal cancer patients, indicating that
ERα36 has a role in metastasis. In laryngeal cancer it seems that ERα36 has a pivotal role in tumorigenesis and tumor progression, with both indolent laryngeal cancer samples and epithelia adjacent to aggressive samples expressing high levels of ERα36.

The epithelia adjacent to all the tumors collected from Meir Hospital, Israel in our study expressed high levels of ERβ, and the level of expression of ERβ was maintained in the indolent tumor. In contrast the aggressive tumors from this sample set demonstrated a marked decline or full suppression of the expression of ERβ. These results indicate that ERβ confers protective effect in laryngeal cancer cells, an effect that has also been reported in previous studies of head and neck carcinoma. ERβ has been postulated to be a dominant-negative regulator of ERα modulating transcriptional responses to estrogens. The ratio of ERα vs. ERβ within a cell may determine the cell sensitivity to E2 and its biological responses. This protective effect of ERβ has been observed in breast cancer cells in which the expression of ERβ was found to inhibit proliferation and cell invasion. Further, ERβ expression has been clinically correlated with low grade tumors, low S phase fraction, negative axillary node status and most importantly higher likelihood to respond to hormonal therapy. These findings coincide with our observation that advanced laryngeal tumors with metastasis to regional lymph nodes, fully suppress the expression of ERβ. The relative conservation of ERβ was correlated with a more indolent tumor behavior. Up to 90% of triple negative breast cancer have been found to express ERβ, the activation of which resulted in suppression of cell proliferation, presumably due to suppression of cyclin kinase 1 and 7 and blockage of cell cycle progression. ERβ target genes have been postulated to regulate apoptosis and cell survival, cell development and movement, growth and proliferation as well as genes
involved in β-catenin and cell cycle checkpoint pathways.\textsuperscript{217} Knockdown of the ERβ expression in triple negative breast cancer cells increased the invasiveness of the cells about three fold,\textsuperscript{219} while ERβ agonists had the opposite effect,\textsuperscript{220} raising the possibility of their role in the treatment of these cancers.

A number of studies have identified geographic and racial disparities in ERα and ERβ expression in other hormonally responsive cancers, particularly breast cancer and prostate cancer. Geographic incidences of ER negative breast cancer are highly variable both within the U.S. and internationally,\textsuperscript{221–225} and this disparity has been associated with higher incidences of ERα negative, ERβ positive, and triple-negative breast cancer in women of African descent.\textsuperscript{226–231} The opposite trend has been observed in prostate cancer, with one study of 300 prostate cancer samples from African American (AA) and Caucasian American (CA) men identifying increased ERβ expression in tumor and precancerous samples from AA patients as compared to CA men.\textsuperscript{232} However, both sets of patients had increased intra-tumor ERβ staining as compared to matched precancerous samples.

The proportion of ERα positive breast cancer incidence relative to the total diagnoses is similar for patients in the United States and Northern Israel;\textsuperscript{224,225} however, our data suggest that ERβ positive cancer incidence may be more regionally localized. Other studies have reported high variability of ERβ expression in laryngeal cancer samples, similar to the trends we observed in our samples.\textsuperscript{233} It is important to note that, although western blots were conducted in a centralized lab, our samples were originally fixed and processed at two different institutions, which could have caused ‘batch processing effects’ that could also account for the regional variation we observed in our
samples. Additional demographic data on race and ethnicity were not available for our samples; however, the wide variability of ERα and ERβ expression in our data could be indicative of variability in race and ethnicity among our sample set. While our sample size is too small to draw conclusions, further investigation into geographic variability of ERα and ERβ expression in laryngeal cancer is needed to further understand the potential of ERs as diagnostic and prognostic markers of laryngeal cancer.
Chapter 5.

24R,25-Dihydroxyvitamin D₃ regulates breast cancer cells \textit{in vitro} and \textit{in vivo}

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Introduction

Interest in anti-tumorigenic effects of vitamin D₃ began with reports of an inverse association between sunlight exposure and age-adjusted breast cancer mortality \cite{234}. Subsequent studies have demonstrated a connection between serum 25-hydroxyvitamin D₃ (25(OH)D₃) and breast cancer risk and prognosis, with low serum levels correlating with increased incidence of breast cancer and high serum levels correlating with reduced recurrence and increased survival in breast cancer patients \cite{112,235,236}. Despite this, studies evaluating the effectiveness of vitamin D supplementation on cancer progression and survival have been inconclusive \cite{236–240}. Interestingly, one study examining post-remission vitamin D supplementation showed that supplement use was associated with reduced recurrence rates among patients with estrogen receptor (ER) positive breast cancer, but not patients with ER negative breast cancer \cite{8}. Similarly, high serum 25(OH)D₃ levels correlate with improved survival as measured by reduced hazard ratios in post-menopausal patients (statistically associated with higher rates of ER positive cancer \cite{241}), but no correlation was identified in pre-menopausal patients (statistically associated with ER negative or triple-negative breast cancer \cite{242} \cite{243}).
Many physicians prescribe vitamin D3 to breast cancer patients, and even more breast cancer patients are believed to supplement their dietary vitamin D intake with over-the-counter supplements. Vitamin D3 is hydroxylated in the liver to form 25(OH)D3, which binds to albumin and vitamin D-binding protein in the blood and is subsequently transported to the kidney where it is metabolized to 1,25-dihydroxyvitamin D3 (1,25(OH)2D3) and 24,25-dihydroxyvitamin D3 (24,25(OH)2D3). Although the kidney produces the bulk of circulating 1,25(OH)2D3 and 24,25(OH)2D3, 25(OH)D3 metabolism also occurs locally in some cell types.

Preclinical studies indicate that 1,25(OH)2D3 and its precursor 25(OH)D3 may have potential as a preventive therapy for breast cancer. Oral vitamin D3 increased autophagy and cell turnover in normal murine mammary gland and both oral vitamin D3 and intravenous 1α,25(OH)2D3 reduced tumor growth in estrogen receptor (ER) positive breast cancer xenograft models. In vitro, 1α,25(OH)2D3 reduced cancer cell growth, angiogenesis, and metastasis, and induced apoptosis in ER positive breast cancer cells but not in normal breast epithelial cells. This may be a function of 1α,25(OH)2D3’s genomic or non-genomic actions. 1α,25(OH)2D3-liganded vitamin D receptor (VDR) suppresses transcription of the ESR1 gene; and both 1α,25(OH)2D3 and its non-VDR binding analogs 1,24,25(OH)3D3 and 1,25,26(OH)3D3 inhibit growth of breast cancer cells in vitro.

While these studies are promising, there is some concern about the risk of hypercalcemia that accompanies treatment with high doses of 1α,25(OH)2D3. Both 25(OH)D3 and 1α,25(OH)2D3 have narrow therapeutic windows when given intravenously, with serum levels of ~1x10^-6M 25(OH)D3 and ~9x10^-10M 1,25(OH)2D3 being...
associated with clinical hypercalcemia. The potentially toxic effect of 1α,25(OH)_{2}D_{3} with regard to calcium handling and its narrow therapeutic window prompted the search for synthetic vitamin D analogs that share its anti-cancer properties while bearing a less calcemic profile.

We propose an alternative endogenous metabolite of vitamin D, 24R,25(OH)_{2}D_{3}. This naturally occurring steric epimer of 24,25(OH)_{2}D_{3} plays an active role in developing skeletal tissue. Furthermore, 24,25(OH)_{2}D_{3} is less calcemic than 1,25(OH)_{2}D_{3}, with doses up to 500 times greater than 1,25(OH)_{2}D_{3} toxicity thresholds having no effect on serum calcium in rats.

The identity of the 24R,25(OH)_{2}D_{3} receptor remains elusive. Larsson et al. reported specific receptors for this vitamin D metabolite in lysosomal membranes in intestinal cells and Seo et al. and Kato et al. have shown evidence for a membrane-bound receptor for 24,25(OH)_{2}D_{3} in chick fracture healing callus. Martineau et al. recently identified a novel 24R,25(OH)_{2}D_{3} receptor, FAM57B2, which may regulate the 24R,25(OH)_{2}D_{3} response in chondrocytes. Analysis of the crystal structure of the classical VDR suggests that conformational changes in the receptor may accommodate binding of a variety of 1,25(OH)_{2}D_{3} analogues, raising the possibility that 24,25(OH)_{2}D_{3} might also occupy the ligand binding pocket on the receptor, at least transiently. Studies using rat growth plate resting zone chondrocytes have characterized specific membrane-associated signal transduction pathways that mediate both rapid non-genomic, and genomic responses, supporting the presence of receptor(s) for 24R,25(OH)_{2}D_{3} but whether one or more of the receptors identified above are involved is not known.
Steroid hormones act through both traditional and non-traditional receptors, which operate in parallel pathways and collaborate to exert the hormone’s full effect\textsuperscript{68}. Such a model exists in the case of 1\(\alpha\),25(OH)\(_2\)D\(_3\); VDR and membrane-associated Pdia3 co-localize with the caveolae scaffolding protein, caveolin-1, in osteoblasts. Ligand binding changes the interactions of the two receptors and rapidly activates phospholipase A\(_2\) (PLA\(_2\)) and Src\textsuperscript{254,255}. 24R,25(OH)\(_2\)D\(_3\) acts via membrane-associated phospholipid metabolism, resulting in activation of phospholipase D (PLD)-mediated signaling and protein kinase C, amplifying the complexity of the actions of this secosteroid\textsuperscript{15,105}.

24R,25(OH)\(_2\)D\(_3\) is produced via hydroxylation of 25(OH)D\(_3\) on C-24 by CYP24A1\textsuperscript{256}. Overexpression of CYP24A1 increases 24,25(OH)\(_2\)D\(_3\), which may have growth modulating effects independently and in combination with 1,25(OH)\(_2\)D\(_3\) on various cancers, including ovarian cancer\textsuperscript{257}, colon cancer\textsuperscript{98}, lung cancer\textsuperscript{258} and breast cancer\textsuperscript{128,259}. One such study showed that 24R,25(OH)\(_2\)D\(_3\) treatment prolonged survival of mice with Lewis lung carcinoma \textit{in vivo}\textsuperscript{260,261}. In rats, 24R,25(OH)\(_2\)D\(_3\) reduced the development of cancerous and precancerous lesions in the glandular stomach in a dose-dependent manner by altering calcium pharmacodynamics during the post-initiation phase of glandular stomach carcinogenesis\textsuperscript{262}. In the colon, a similar chemopreventive effect was observed via anti-proliferative actions. Pretreatment and, more markedly, continuous administration of 24R,25(OH)\(_2\)D\(_3\) reduced the total numbers of aberrant crypt foci and putative pre-neoplastic lesions to inhibit development of colon cancer foci\textsuperscript{263}. Alterations in c-fos, c-myc, and c-jun oncogene expression were proposed as potential mechanisms for incurring these anti-proliferative actions, suggesting that rapid actions of the vitamin D metabolite were involved.
In the search for compounds that mimic the growth regulating effects of vitamin D₃ independent from its calcium mobilizing actions, 24R,25(OH)₂D₃ has emerged as an exceptional candidate as it incorporates the anti-proliferative and anti-tumorigenic effects of 1α,25(OH)₂D₃ without disrupting calcium homeostasis by elevating calcium blood levels. To further improve the understanding of the role of 24R,25(OH)₂D₃ in the tumorigenesis and progression of breast cancer we established an in vivo model of tumor growth and metastasis in mice using MCF7 breast cancer xenografts. In addition, we examined the underlying mechanism of 24R,25(OH)₂D₃’s effect in vitro using MCF7. We hypothesized that the effect exerted by 24R,25(OH)₂D₃ on breast cancer cells involves protecting against proliferation, promoting apoptosis and suppressing migration and metastatic gene expression, resulting in reduced invasion and metastasis. To validate the hypothesis that the secosteroid can modulate breast cancer cell survival, we also assessed the response of two breast cancer cell lines.

Materials & Methods

Cell Lines

MCF7, T-47D, and HCC38 breast cancer cells were authenticated by ATCC (Manassas, VA) using STR profiling and were used within 6 months of purchase as described previously. Cells were maintained in media and passaged in accordance with ATCC guidelines. Red fluorescent protein (RFP) expressing-MCF7 cells (RFP-MCF7) were created by transfecting cells with an RFP-encoding plasmid (pMirTarget, #PS100062, Origene, Rockville, MD) using TurboFectin 8.0 (Origene #TF81001) according to manufacturers’ protocol.
In Vivo Studies

Female NSG mice skeletally mature (6-8 weeks old) were obtained from the Virginia Commonwealth University (VCU) Cancer Mouse Models Core Laboratory at Massey Cancer Center, (Richmond, VA). All animal experiments were approved by VCU’s Institutional Animal Care and Use Committee.

MCF7 Xenograft Model.

To create MCF7 tumors, female 6-8 weeks old mice (N = 8) were injected intraperitoneally with 2.5mg/kg of 17β-estradiol (#E8875; Sigma-Aldrich, St. Louis, MO) or 2% ethanol:1X distilled phosphate buffered saline (DPBS) vehicle one week prior to tumor initiation. These injections continued twice a week for the remainder of the study (11 weeks in total). One week after the first 17β-estradiol or vehicle injection, tumors were initiated by injecting 1 million MCF7 cells in 50μL of 1:1 DPBS:BME (Cultrex Pathclear basement membrane extract Type 3, 14.5mg/mL; #3632; R&D Systems, Minneapolis, MN) solution into the lactiferous duct of the right fourth mammary gland of the mouse\textsuperscript{264–266}. Beginning at 2 weeks post-cell injection, tumors were measured along two perpendicular axes (length and width) with digital calipers, and tumor volume was calculated using the following formula: \( Volume = \frac{Length \times Width^2}{2} \). Tumor volume was plotted against time and comparisons between vehicle and control mice were made using a repeated-measured analysis of variance (ANOVA). P-values <0.05 were considered significant. Tumor measurements were taken once a week for 10 weeks until harvest.
Mice were euthanized at 10 weeks via CO₂ inhalation. Tumors were harvested and preserved in 10% neutral buffered formalin. Fixed tissues were processed, embedded in paraffin, and cut into 3-5μm sections. Sections were stained with H&E to examine cell morphology.

In a second study, 5-week old female NSG mice were ovariectomized. Two weeks post-operatively, mice were injected with 17β-estradiol (N=6) or vehicle (N=6) twice a week for 11 weeks as described above. One week after the first 17β-estradiol or vehicle injection, mice were implanted with MCF7 cells in the right fourth mammary gland as described above. Tumor size was measured once a week with digital calipers and tumor volume was calculated using the established system described above. Tumors grown in ovariectomized mice were harvested after 10 weeks, analyzed with μCT, and preserved in formalin. Histologic sections were stained with H&E and analyzed. Tumor volume and percent growth of tumor were graphed vs. time. Animals in both studies were monitored for signs of estrogen toxicity, such as urinary bladder distension and skin rash.

**μCT Tumor Volume Measurements.**

Tumors were harvested and immediately scanned with a Bruker Skyscan 1173 μCT at 55kV and 70μA, with a resolution of 560x560 pixels with an image pixel size of 40.26μm, an exposure time of 125ms and a rotation step of 0.8°. A standard reconstruction was conducted on the tumors using the NRecon software version 1.6.10.4 (Kontich, Belgium) with a smoothing kernel of 0 and a beam hardening correction of 20%. μCT reconstructions of the tumors were analyzed using the CTAn software version 1.16.1.0 (Kontich, Belgium). Total tumor volume was measured as the total tissue volume.
Calculated tumor volumes as measured by digital calipers and total tumor volumes from μCT scans were compared using a 2-way ANOVA with Bonferroni post-tests with α=0.05.

Effect of 24R,25(OH)2D3.

Two weeks after subcutaneous implantation of zero-order release 17β-estradiol pellets (0.96mg/45 days) (Innovative Research of America, Sarasota, FL) at ‘week 0’, female NSG mice (N=24) were injected with 1 million RFP-MCF7 cells in the right lactiferous duct of the fourth mammary fat pad. MCF7 tumors were allowed to grow for 5 weeks. At that time, mice were divided into 3 groups (n=8) and each group was injected intraperitoneally 3 times a week with 0, 25ng, or 100ng of 24R,25(OH)2D3 (#BML-DM300-0050; Enzo Life Sciences, Farmington, NY) in a 2% ethanol: DPBS vehicle. Mammary fat pad tumors were measured with calipers throughout the study. Tumor volume and tumor accretion rate (tumor volume over time normalized to original tumor volume) were graphed vs. time. Tumor growth rate from weeks 6-10 was analyzed with a linear regression model for each group (0, 25, and 100ng of 24R,25(OH)2D3). Animal survival was tracked throughout the study and percent surviving mice vs. length of survival was recorded. Animal survival was plotted on a hazard curve and analyzed with the chi-squared logrank test for trend at α=0.05. Throughout the study, animals were monitored for signs of hypercalcemia, such as changes in appetite or lethargy.

On the day of harvest, animals were euthanized with CO2 and metastases was identified by the presence of RFP (443/581nm) using a Zeiss SteroDiscovery.V12 fluorescence dissecting microscope with an AxioCam MRm digital camera. Tumors were harvested and analyzed with μCT. Axial and inguinal lymph nodes, lungs, and livers were grossly examined for RFP expression. Fluorescing organs were harvested and fresh
flash-frozen on block dry ice in optimal cutting temperature compound (OCT) (#25608; VWR, Radnor, PA) for histological analysis. After μCT analysis, tumors were fresh flash-frozen in OCT as above. OCT-embedded samples were cut into 3-5μm sections on a cryostat, post-fixed with fresh 4% paraformaldehyde for 10 minutes, and stained with H&E to confirm metastasis. Total metastatic incidence and metastatic incidence by organ were recorded.

**In vitro Studies**

**Response to Vitamin D3 Metabolites.**

To assess specificity of the response of MCF7 cells to 24R,25(OH)_{2}D_{3} confluent cultures were treated with media containing 1α,25(OH)_{2}D_{3} (10^{-10}-10^{-8}M, Enzo #BML-DM200-0050); 24R,25(OH)_{2}D_{3} (10^{-9}-10^{-7}M); 24S,25(OH)_{2}D_{3} (10^{-9}-10^{-7}M, Sigma Aldrich #29447); 25(OH)D_{3} (10^{-9}-10^{-7}M, Sigma Aldrich #17938); or vehicle. All vitamin D metabolites were reconstituted to 1mM stock solutions in absolute ethanol. Stock solutions were diluted in warm complete media to final treatment concentrations. Media were removed by aspiration at 15 min or 24 hours and cells were incubated with fresh complete media until harvest as indicated below.

**Signaling Pathway Inhibition.**

In these experiments, cells were pre-treated for 30 minutes with inhibitors or vehicle in warm complete media. At that time, fresh media containing 24R,25(OH)_{2}D_{3} or vehicle were added on top of inhibitor treatments, and cells were incubated for a further 15 minutes before the conditioned media were aspirated. Cultures were then incubated in fresh complete media until harvest. To determine if the effect of 24R,25(OH)_{2}D_{3} was
mediated by palmitoylation, cultures were pretreated with 1-10μM 2-bromopalmitate (Sigma-Aldrich #21604)\textsuperscript{269}. PLD was inhibited by pretreatment with 0.1 to 1 μM wortmannin (VWR #80055508, Radnor, PA)\textsuperscript{15}. To assess whether intact caveolae were required, cultures were pretreated with 1 or 10mM methyl β-cyclodextrin (Sigma-Aldrich #C4555) in serum-free media to deplete cell membranes of cholesterol\textsuperscript{169,270,271}. In order to assess the role of ERα66, cultures were pretreated with 10\textsuperscript{-7}M ICI-182780, an inhibitor to cytosolic and nuclear ERs (Sigma-Aldrich #I4409)\textsuperscript{272,273}. The contribution of ERα36 was determined by pretreating MCF7 cells with 1μg of a neutralizing antibody to the membrane-associated estrogen receptor (Chi Scientific #8-80113, purchased from VWR)\textsuperscript{55}.

**Outcome Measures**

_DNA synthesis_. Cells were cultured to 70% confluence, then serum-starved for 48 hours in phenol-red free MEM supplemented with 1% charcoal-dextran filtered FBS and 50U/50U penicillin-streptomycin. At that time, cells were treated with 24R,25(OH)\textsubscript{2}D\textsubscript{3}, 24S,25(OH)\textsubscript{2}D\textsubscript{3}, 25(OH)D\textsubscript{3}, or 1α25(OH)\textsubscript{2}D\textsubscript{3} for 15 minutes and incubated with fresh complete media for 24 hours. In a separate experiment, cells were treated with these vitamin D3 metabolites for 24 hours. At the 20 hour mark, cells in both experiments were pulsed with 10μL of a 1:100 dilution of 5-ethynyl-2'-deoxyuridine (EdU) and incubated for 4 hours. Cells were harvested and assayed for EdU incorporation according to manufacturer’s instructions (ThermoFisher Scientific #C10214).

_Gene expression_. Cells were cultured to confluence, then treated with 24R,25(OH)\textsubscript{2}D\textsubscript{3} or 24S,25(OH)\textsubscript{2}D\textsubscript{3} or vehicle in complete media for 15 minutes as described above. Cultures were then aspirated and incubated with complete media for 12
hours. RNA was extracted using TRIzol (Invitrogen, Carlsbad, CA) and quantified (Take3 Micro-Volume Plate, Biotek, Winooski, VT) before being used to synthesize cDNA libraries (High Capacity cDNA Reverse Transcription kit, Promega, Madison, WI). Apoptosis markers were B cell lymphoma protein 2-associated X protein (BAX) to B cell lymphoma protein 2 (BCL2) mRNA expression. Epithelial to mesenchymal transition markers were C-X-C- motif chemokine 12 (CXCL12), chemokine receptor type 4 (CXCR4), snail family transcriptional repressor 1 (SNAI1), erythroblastic oncogene B (ERBB2), and matrix metalloproteinase 1 (MMP1). Markers of metastasis were osteoprotegerin (OPG), and nuclear factor kappa-B ligand (RANKL)

mRNAs were quantified with real-time quantitative PCR using Power SybrGreen® Master Mix (Applied Biosystems) and gene-specific primers described in Table 5.1. Commercially available primers (Qiagen, Hilden, Germany) were used to quantify BCL2 (Hs_BCL2-1-SG QuantiTect Primer #249900), ESR1 (Hs_ESR1-1-SG QuantiTect Primer #044492), RANKL (Hs_TNFSF11-1-SG QuantiTect Primer #PPH01048F), and SNAI1 (Hs_SNAI1-1-SG QuantiTect Primer #010010) mRNA. All other primers were self-designed using the National Center for Biotechnology Information PrimerBLAST and ordered from ThermoFisher Scientific (Waltham, MA). Exact sequences for all ThermoFisher primers are given in Table 5.1. RNA quantities were determined by the standard curve method. mRNA levels were normalized to levels of glyceralde-3-phosphate dehydrogenase (GAPDH) mRNA. mRNA for BAX, BCL2, CXCR4, and CXCL12 are presented as ratios of BAX to BCL2 and CXCR4 to CXCL12.
Table 5.1 Human primers used in real-time PCR analysis.

<table>
<thead>
<tr>
<th>Gene</th>
<th>Primer Sequence</th>
</tr>
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<tbody>
<tr>
<td>BAX</td>
<td>F GACGAACTGGACAGTAACATGG</td>
</tr>
<tr>
<td></td>
<td>R AAAGTGAAGAAAGGCCGACAACC</td>
</tr>
<tr>
<td>CXCL12</td>
<td>F GCCTCTGAAGCCTATGTATG</td>
</tr>
<tr>
<td></td>
<td>R GACGAACTGGACAGTAACATGG</td>
</tr>
<tr>
<td>CXCR4</td>
<td>F TAGCAAAGTGACGCCGAGGG</td>
</tr>
<tr>
<td></td>
<td>R TGGTTTCTCCAGATGCGGTGG</td>
</tr>
<tr>
<td>ERBB2</td>
<td>F CCGCTGAAGTCCACACAGTT</td>
</tr>
<tr>
<td></td>
<td>R AAAGGTTCTACCCGCATGG</td>
</tr>
<tr>
<td>GAPDH</td>
<td>F CTATAATAATGAGCCCAGCC</td>
</tr>
<tr>
<td></td>
<td>R TGGGCAGCCAAAAAGAGATG</td>
</tr>
<tr>
<td>MMP1</td>
<td>F AGAAAGAAGACAAAGGCAAGTT</td>
</tr>
<tr>
<td></td>
<td>R TCCCCAGTCACTTTCCAGCC</td>
</tr>
<tr>
<td>OPG</td>
<td>F GTGTGCCAATGCAAGGAAGG</td>
</tr>
<tr>
<td></td>
<td>R CACTCCAAATCCAGGAGGC</td>
</tr>
</tbody>
</table>

Total p53 content. Total p53 was measured 24 hours post-cell treatment with vitamin D metabolites as described above using a sandwich enzyme-linked immunosorbent assay (ELISA) (R&D Systems, Human Total p53 DuoSet® IC). Cells were plated in 24 well plates, treated with vitamin D3 metabolite for 15 minutes, and incubated for 24 hours in complete media. Conditioned media were collected and cell layers were lysed by sonication (40A for 5 seconds) in 200μL lysis buffer according to manufacturer’s protocol. Lysates were centrifuged at 10,000g for 15 minutes; the supernatant was collected and assayed for total protein content (ThermoFisher, Pierce 660 nm Protein Assay) and total p53 content following the manufacturer’s instructions. Data are presented as a ratio of nanograms of total p53 to total protein content.
**DNA fragmentation.** DNA fragmentation was assessed by colorimetric terminal deoxynucleotidyl transferase (TdT)-mediated dUTP nick end labeling (Trevigen, TiterTAC™ *in situ* microplate TUNEL assay). Cells were incubated with vehicle, 24R,25(OH)₂D₃, or 24S,25(OH)₂D₃ for 15 minutes followed by incubation in fresh media for 24 hours. After fixing the cultures with 3.7% sucrose-buffered formaldehyde, samples were processed for TUNEL staining following manufacturer’s instructions. Final absorbance was detected at 450 nm using a microplate spectrophotometer.

**Enzyme-linked immunosorbent assays for secreted proteins.** Confluent cultures of MCF7 cells were plated in 6 well plates, treated with vehicle or 24R,25(OH)₂D₃ for 15 minutes as described above, aspirated, and then incubated in complete media. After 24 hours, conditioned media were collected in 5mL polypropylene tubes, frozen at -80°C and lyophilized (Labconco, Kansas City, Missouri, Freezone Freeze Dryer #7740020). Lyophilized samples were reconstituted in 200 μL of filtered 1XPBS containing 1% BSA and assayed for OPG and RANKL using sandwich ELISAs (R&D Systems, Human TRANCE/RANKL/TNFSF11 DuoSet ELISA #DY626, Human Osteoprotegerin/TNFRSF11B DuoSet ELISA #DY805). Cell monolayers were lysed in 1mL 0.5% Triton™ X-100. Cell lysates were assayed for total DNA content (Promega, QuantiFluor dsDNA, #E6150). OPG and RANKL content are expressed as a ratio of OPG/RANKL.

**Matrix metalloproteinase 1.** Collagenase (MMP1) content in the cultures was measured using an MMP1 assay kit (SensoLyte® Plus 520 MMP-1 Assay Kit *Fluorimetric and Enhanced Selectivity*, Anaspec #AS-72012). Cells were plated in 24 well plates and treated with vehicle or 24R,25(OH)₂D₃ as described above. At harvest
conditioned media were collected; cell monolayers were lysed in 500μL 0.05% Triton-X-100 and total DNA content measured. Total MMP1 was measured in the media and normalized to total DNA content. Data are presented as a ratio of ng MMP1/μg DNA.

Wound closure assay. A wound closure assay was used to demonstrate metastasis. Confluent cultures of MCF7 cells were treated for 15 min with media containing vehicle or 24R,25(OH)2D3. Another set of cultures was treated with 10−7M 17β-estradiol as a positive control. Following treatment, a scratch was drawn from one end of the well to the other using a sterile 1 mL pipette tip. The loosened cells were aspirated along with the treatment media, and fresh media added to the cultures. A phase-contrast microscope was used to take pictures of each well at 0, 3, 6, 9, 12, and 24 hours after the scratch was made. Wound area was calculated using Adobe Photoshop™ 2 software, normalized to the original wound closure area, and presented as percent wound closure. Data presented are from one of two repeated experiments.

Statistical Analysis

Data shown are from single representative experiments of two or more repeats and are presented as mean ± standard error of six independent cultures per treatment group. Data were analyzed by ANOVA with Tukey’s post-test to determine significance between groups. Experiments with two independent variables (DNA synthesis, total DNA content, and total p53 time courses) were analyzed with two-way ANOVA with Bonferroni’s modification of Student’s t-test post-tests to determine the variance between row and column factors. The wound closure assay was analyzed with a repeated measures ANOVA test with comparisons between time points. P-values <0.05 were considered significant.
Results

17β-Estradiol supplementation stimulated MCF7 breast cancer tumor growth in mice.

MCF7 (estrogen receptor positive, ER+, progesterone receptor positive, PR+) breast cancer cells were injected into the duct of the fourth mammary gland. Mice receiving 17β-estradiol injections grew tumors 3-5 times their original size. In contrast, tumors grown in animals given a vehicle control injection showed minimal growth over 10 weeks (Figure 5.1). Tumors in mice given 17β-estradiol increased from approximately 40mm³ to an average of 200mm³, approximately 500% of their original volume, with a linear growth increase over the course of 8 weeks (week 2 to week 10). However, tumors grown in mice given a vehicle injection reached an average size of 55mm³, an increase of approximately 10% of their original volume (Figure 5.1A). In ovariectomized animals, the difference between vehicle versus 17β-estradiol injected mice was even more pronounced, with tumors grown in 17β-estradiol injected mice reaching an average size of 100mm³ versus only 25mm³ in vehicle-injected ovariectomized mice (Figure 5.1B,C). Tumor morphology was similar in both 17β-estradiol and vehicle-injected mice, and there was no observed difference in tumor morphology in samples taken from normal vs. ovariectomized mice [representative image, (Figure 5.1D)]. At harvest, final tumor volume in 17β-estradiol animals was approximately 4 times greater than tumor volume in vehicle-treated animals in both the normal and ovariectomized mouse models (Figure 5.1E). None of the animals in either experiment showed signs of estrogen toxicity.
Figure 5.1 17β-estradiol stimulates MCF7 tumor growth in a mammary fat pad xenograft model in NSG mice.

[A] Non-ovariectomized mice injected with 17β-estradiol showed a significant increase in tumor growth as compared to the vehicle after 3 weeks of injection until the end of the study at week 10. [B] Similar trends were observed in a separate study with ovariectomized animals, with animals treated with 17β-estradiol showing a significant increase in tumor growth as compared to vehicle at weeks 4 and 5 and weeks 7-10 of the study. [C] At harvest, tumors grown in ovariectomized mice supplemented with 17β-estradiol reached an average volume of 120mm³, nearly 6 times greater than tumors grown in vehicle-treated mice. [D] Histology of the fourth mammary fat pad revealed neoplastic tissue characteristic of MCF7 tumors.
(continued from page 124) [E] Final tumor volume was largest in control mice supplemented with 17β-estradiol. Ovariectomized mice supplemented with 17β-estradiol had tumor volumes that were smaller than those from normal mice, but still larger than tumors grown in ovariectomized mice given a vehicle control. Groups that share a letter are not statistically significant at α=0.05 with two-way ANOVA.

Treatment with 24R,25(OH)2D3 reduced tumor aggression and increased animal survival.

Red fluorescent protein (RFP)-overexpressing MCF7 mammary fat pad tumors were grown in female NSG mice for 10 weeks. Estrogen supplementation was given for 2 weeks prior to tumor initiation, then continued for the first 4 weeks of tumor growth. Dose-dependent 24R,25(OH)2D3 was initiated at the beginning of week 5, and continued for the rest of the 10 week study. Mice receiving either 25ng or 100ng per injection of 24R,25(OH)2D3 survived longer than mice injected with vehicle alone. 6/8 and 7/8 animals in the 25ng and 100ng groups, respectively, survived until harvest, while only 3/8 animals survived in the control group (Figure 5.2A). Animals in the control group also started dying earlier (at day 38) than animals given 24R,25(OH)2D3 (day 56) (Figure 5.2B). Both 25ng and 100ng treated tumors exhibited a lower percent increase in size compared to vehicle-treated mice. Tumors in both 24R,25(OH)2D3 treated groups achieved approximately 500% of their original volume, while vehicle-treated tumors grew to approximately 1000% of their original volume (Figure 5.2C). Tumors in mice given 25ng 24R,25(OH)2D3 reached volumes of approximately 400mm³ at harvest, roughly 80% the size of the tumors in
vehicle-treated mice, which reached a volume of 500mm$^3$ on average. Tumors in mice given 100ng showed an even steeper growth depression, reaching final volumes of 200mm$^3$ on average, roughly 40% of vehicle-treated tumors (Figure 5.2D,E).

The difference in tumor volume was due to growth that occurred in the last 4 weeks of the study, after 24R,25(OH)$_2$D$_3$ injections were begun. All tumors were approximately the same size at week 6, at the onset of treatment. From week 6-10, vehicle-treated tumors had a growth rate of 79.54 ± 9.40 mm$^3$/week ($R^2$ =0.96); 25 ng-treated tumors had a growth rate of 57.90 ± 8.70 mm$^3$/week ($R^2$ =0.94); and 100ng-treated tumors had a growth rate of 40.61 ± 6.60 mm$^3$/week ($R^2$ =0.93). The slopes were significantly different ($P$=0.0213). Calculated tumor volumes from caliper measurements were not significantly different from tumor volumes generated by μCT analysis (Figure 5.2E). Haematoxylin and eosin (H&E) staining revealed similar tumor morphology in tumors from each group [representative image, (Figure 5.2F)].
Figure 5.2 24R,25(OH)2D3 reduces MCF7 tumor burden and increases animal survival.

[A] After 8 weeks, 3 animals from the vehicle group, 6 animals from the low-dose 24R,25(OH)2D3 group, and 7 animals from the 24R,25(OH)2D3 group survived out of an original n of 8. [B] Animals in the vehicle group had a statistically significant reduced survival rate as compared to animals in the low or high dose groups by the chi-squared logrank test for trend (P=0.0433). [C] Tumor accretion rate was calculated by normalizing tumor volumes at each week to the original tumor volume calculated at week 2. (continued from page 122) [D] Animals given 24R,25(OH)2D3 showed a decreased rate of growth after vitamin D3 injections were begun at week 6 of the study. Statistics were done using a repeated-measures ANOVA test to compare between the vehicle, low, and high-dose groups within weeks.
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(continued from page 127) Purple squares □ indicate significance against the high-dose group, and blue open circles ○ indicate significance against the low-dose group. [E] After harvest, tumor volume was quantified using µCT analysis. Calculated caliper volumes and µCT-derived volumes were graphed together, and statistics were done using ANOVA with Tukey’s correction. Bars that share a letter are not significant at α=0.05. Both caliper measurements and µCT analyses showed statistically significant decreases in final tumor volume between the high-dose 24R,25(OH)₂D₃ group and the vehicle control; and µCT analysis showed a statistically significant difference between both the vehicle and the low-dose 24R,25(OH)₂D₃ group as compared to the high-dose 24R,25(OH)₂D₃ group. [F] Histology of the fourth mammary fat pad showed cells with a dense epithelial-like phenotype consistent with MCF7 morphology and previous MCF7 xenograft studies, bar 50 µm.

Metastatic burden was detected in animals by assessing individual organs for RFP expression using a fluorescent dissection microscope at harvest. Animals treated with 24R,25(OH)₂D₃ had a lower metastatic burden than vehicle-injected controls, with fewer organs that were positive for RFP-tagged MCF7 cells. Animals treated with 25ng 24R,25(OH)₂D₃ had, on average, 1-2 incidences of metastasis/animal, while 100ng treated animals averaged 0-1 metastatic incidence per animal. Control group animals averaged 3-4 incidences of metastasis per animal (Figure 5.3A). Animals treated with 24R,25(OH)₂D₃ also had fewer incidences of local and distant metastasis than control group animals, with animals injected with 25ng 24R,25(OH)₂D₃ having fewer incidences of multiple lymph node, lung, and liver metastases. This effect was even greater in
animals treated with higher doses of 24R,25(OH)$_2$D$_3$ (100ng). These animals had fewer lymph node and lung metastases and no incidences of multiple lymph node or liver metastases (Figure 5.3B). Metastatic sites were characterized by round clusters of RFP-positive cells with defined edges, characteristic of MCF7 metastases$^{279}$ (Figure 5.3C-E). Morphology of metastatic sites was similar to the morphology of the primary tumor (Figure 5.3F-H). None of the animals showed signs of hypercalcemia throughout the study or upon dissection at harvest.

24R,25(OH)$_2$D$_3$ induced both proliferation and apoptosis in ER$^+$ cells in vitro.

When MCF7 cells were treated for 15 minutes with 24R,25(OH)$_2$D$_3$, its enantiomer 24S,25(OH)$_2$D$_3$, or its precursor 25(OH)D$_3$, proliferation was increased after 24 hours. The effect of 24R,25(OH)$_2$D$_3$ and 24S,25(OH)$_2$D$_3$ was dose-dependent with the greatest increase at $10^{-8}$M 24R,25(OH)$_2$D$_3$ (Figure 5.4A) and $10^{-7}$M 24S,25(OH)$_2$D$_3$ (Figure 5.4B); all doses ($10^{-9}$-$10^{-7}$M) of 25(OH)D$_3$ increased proliferation (Figure 5.4C). In contrast, 1α,25(OH)$_2$D$_3$, did not increase proliferation at 24 hours (Figure 5.4D). None of these metabolites increased proliferation in MCF7 monolayers that were treated with vitamin D metabolites for 24 hours (Figure 5.5), and only 24R,25(OH)$_2$D$_3$ increased apoptosis in these cells after 24 hours (Figure 5.6).
Figure 5.3 24R,25(OH)2D3 reduces metastatic burden in MCF7 xenograft mouse models.

[A] High-dose 24R,25(OH)2D3 reduced the total metastatic burden of red fluorescent protein-expressing MCF7 tumors and [B] reduced the number of metastatic incidences to the lymph nodes, lungs, and liver. Statistics were not done on metastatic incidence per organ data due to the ordinal nature of the data collected and the small sample size of the surviving animals in the vehicle group (n=3).
In fig [A], groups that share a letter are not significant at α=0.05. Metastatic organs – [C,E] lymph node, [D,G] lungs, and [E,H] liver – were identified at harvest with a fluorescent microdissection scope set [C,E] an RFP/tdTomato filter at 10X magnification and [D] 40X magnification. Metastases were confirmed with [F-H] H&E staining, and images are displayed at [F] 10X or [G,H] 40X magnification.
Figure 5.4 24,25(OH)2D3 induces both proliferation and apoptosis in MCF7 cells with varying specificities.

[A] 24R,25(OH)2D3 induced proliferation in MCF7 cells 24 hours after a 15-minute treatment. [B] Its enantiomer 24S,25(OH)2D3 also increased proliferation in this system [C] and so did their precursor 25(OH)D3. [D] The vitamin D isoform 1α,25(OH)2D3 did not increase proliferation in these cells. [E-G] In contrast, 24R,25(OH)2D3 induced apoptosis in MCF7 cells 24 hours after a 15-minute treatment, [H-J] but its enantiomer 24S,25(OH)2D3 did not. [E,H] Apoptosis was measured by BAX/BCL2 expression, [F,I] total p53 levels, [G,J] and TUNEL staining. Groups that do not share a letter are statistically significant at α=0.05.
Figure 5.5 Treatment with vitamin D3 metabolites for 24 hours does not alter MCF7 cell proliferation.

[A] 24R,25(OH)\textsubscript{2}D\textsubscript{3}, [B] 24S,25(OH)\textsubscript{2}D\textsubscript{3}, [C] 1\alpha,25(OH)\textsubscript{2}D\textsubscript{3}, and [D] 25(OH)\textsubscript{3}D did not induce DNA synthesis in MCF7 cells after 24 hours of treatment.
Figure 5.6 24R,25(OH)₂D₃ specifically induces apoptosis in MCF7 cells after 24 hours of treatment.


24R,25(OH)₂D₃ caused a dose-dependent increase in BAX to BCL2 expression ratios (BAX/BCL2), which was significant at 10⁻⁷M (~4-fold) (Figure 5.4E). 24R,25(OH)₂D₃ had a bi-phasic effect on total p53 levels and TUNEL staining, with the 10⁻⁸M dose producing the maximum increase in p53 levels (~1.5 fold) (Figure 5.4F) and TUNEL (~2 fold) (Figure 5.4G). The enantiomer 24S,25(OH)₂D₃ did not affect apoptosis in these cells, as measured by BAX/BCL2 expression (Figure 5.4H), total p53 levels (Figure 5.4I), and TUNEL staining (Figure 5.4J). In another ER+ cell line, T-47D cells, 10⁻⁷M 24R,25(OH)₂D₃ increased total p53 content (Figure 5.7A), TUNEL staining (Figure 5.8A), and proliferation (Figure 5.8B) after 15 minutes of treatment. In contrast, in a third breast cancer cell line, HCC38, which are ER-, all doses of 24R,25(OH)₂D₃ decreased total p53 content (Figure 5.7).
Figure 5.7 24R,25(OH)₂D₃ differentially changes apoptosis in other breast cancer cell lines.

Figure 5.8 24R,25(OH)_{2}D_{3} increases apoptosis and proliferation in T47D, an ER+ breast cancer cell line.

Treatment of cultures for 15 minutes with 24R,25(OH)_{2}D_{3} increased [A] total TUNEL staining and [B] proliferation in T-47D cell monolayers.

24R,25(OH)_{2}D_{3} decreased epithelial-to-mesenchymal transition, metastasis, and migration in MCF7 cells in vitro.

MCF7 monolayers were treated with dose-dependent 24R,25(OH)_{2}D_{3} for 15 minutes, then harvested after 12 hours to assess gene expression. Treatment with 10^{-8}M and 10^{-7}M 24R,25(OH)_{2}D_{3} significantly reduced expression of the mesenchymal-associated transcription factor SNAI1 compared to cells treated with vehicle (Figure 5.9A). ERBB2 and MMP1, which are associated with tumor aggression and matrix turnover respectively, were also reduced. 24R,25(OH)_{2}D_{3} (10^{-7}M) caused a 50% reduction in ERBB2 expression (Figure 5.9B), and both 10^{-8}M and 10^{-7}M 24R,25(OH)_{2}D_{3} reduced expression of MMP1 expression by more than 75% (Figure 5.9C).
24R,25(OH)$_2$D$_3$ had a biphasic effect on collagenase with the most significant reduction observed in cultures treated with 10$^{-8}$M (Figure 5.9D). 24R,25(OH)$_2$D$_3$ decreased cell migration in the scratch test assay; 10$^{-7}$M 24R,25(OH)$_2$D$_3$ increased wound size to 300% of its original area while both the vehicle control and the positive control (10$^{-7}$M 17β-estradiol) reduced wound size to 50% of original area (Figure 5.9E). 24R,25(OH)$_2$D$_3$ also decreased metastatic markers. 24R,25(OH)$_2$D$_3$ decreased CXCR4 to CXCL12 ratios (CXCR4/CXCL12) in a dose-dependent manner, with 10$^{-7}$M 24R,25(OH)$_2$D$_3$ decreasing CXCR4/CXCL12 expression in MCF7 cells 8-fold as compared to cells treated with vehicle (Figure 5.9F). 24R,25(OH)$_2$D$_3$ also increased OPG to receptor activator of RANKL ratios (OPG/RANKL) - which is contraindicative of osteoclast activity - in both gene expression and protein. 24R,25(OH)$_2$D$_3$ increased OPG/RANKL gene expression 5-fold at 10$^{-7}$M (Figure 5.9G), and both 10$^{-8}$M and 10$^{-7}$M 24R,25(OH)$_2$D$_3$ increased OPG/RANKL protein by 100% (Figure 5.9H).
Figure 5.9 24R,25(OH)2D3 reduces epithelial-to-mesenchymal transition and migration in MCF7 cultures.

[A] 24R,25(OH)2D3 decreases the gene expression of snail [B], her2 [C], collagenase and [D] total collagenase protein. [E] 24R,25(OH)2D3 also inhibited migration in MCF7 monolayers and significantly increased wound size in a scratch test assay as compared to vehicle and E2-treated cultures, and [F] decreased the gene expression ratio of migratory markers CXCR4/CXCL12. [G] 24R,25(OH)2D3 also inhibited bone resorption and distant metastasis marker OPG/RANKL as measured by gene expression and [H] protein. Groups that do not share a letter are statistically significant at α=0.05.
24R,25(OH)$_2$D$_3$ increased proliferation and apoptosis through a membrane-mediated, PLD-dependent pathway.

MCF7 cell monolayers were treated for 30 minutes with increasing concentrations of inhibitors of membrane-associated steroid hormone signaling mechanisms, then treated with high-dose 24R,25(OH)$_2$D$_3$ for 15 minutes. After 24 hours, proliferation and apoptosis markers were assessed. Inhibition of palmitoylation with 2-bromopalmitate blocked the stimulatory effect of 24R,25(OH)$_2$D$_3$ on DNA synthesis (Figure 5.10A), TUNEL staining (Figure 5.10I), and p53 production (Figure 5.10E), indicating that the 24R,25(OH)$_2$D$_3$ effect on both proliferation and apoptosis required docking of protein receptors to the membrane. Treatment of the cultures with wortmannin blocked the 24R,25(OH)$_2$D$_3$ dependent increase in DNA synthesis (Figure 5.10B) and p53 production (Figure 5.10F). The PLD inhibitor increased TUNEL staining in control cultures to levels comparable to 24R,25(OH)$_2$D$_3$ treated cultures (Figure 5.10J). Depletion of membrane cholesterol by methyl β-cyclodextrin blocked 24R,25(OH)$_2$D$_3$ stimulated increases in DNA synthesis (Figure 5.10C). At the higher concentration, it increased p53 levels in control cultures to levels seen in 24R,25(OH)$_2$D$_3$-treated cultures (Figure 5.10G); all methyl-β cyclodextrin treated cultures had TUNEL staining comparable to the 24R,25(OH)$_2$D$_3$-treated cells (Figure 5.10K). Antibodies blocking ERα36 prevented 24R,25(OH)$_2$D$_3$-stimulated proliferation but the treatment of the culture with 10$^{-7}$M ICI-182780, a cytosolic and nuclear ERα inhibitor, had no effect (Figure 5.10D). Both anti-ERα36 and ICI-182780 blocked the stimulatory effect of 24R,25(OH)$_2$D$_3$ on p53 (Figure 5.10H) and TUNEL (Figure 5.10L).
Modulation of 24R,25(OH)\textsubscript{2}D\textsubscript{3} Tumorigenicity by ER\textsubscript{α} Isoforms in E\textsubscript{2}-Responsive Cancer • Spring 2019

Inhibitors against palmitoylation ([A], [E], [I], 2-bromopalmitate), phospholipase D ([B], [F], [J], wortmannin), caveolae ([C], [G], [K], methyl β-cyclodextrin), membrane estrogen receptor α 36 ([D], [H], [L], anti-ER\textsubscript{α36} antibody) and cytosolic and nuclear estrogen receptor α ([D], [H], [L]) (ICI-182780) were used to gain insight into the mechanism by which 24R,25(OH)\textsubscript{2}D\textsubscript{3} acts on apoptosis and proliferation in MCF7 cells. Proliferation was measured by [A-D] DNA synthesis assay and apoptosis was measured by [E-H] total p53 ELISA and [I-L] TUNEL staining. All doses of 2-bromopalmitate, which inhibits palmitoylation, inhibited a 24R,25(OH)\textsubscript{2}D\textsubscript{3} induced increase in [A] proliferation, [E] p53 content, and [I] TUNEL staining. Wortmannin, a non-specific phospholipase-D inhibitor,
(continued from page 135) [B] inhibited a 24R,25(OH)₂D₃ induced increase in proliferation, [F] p53 content, and [J] TUNEL staining in a similar manner. Methyl β-cyclodextrin, which depletes the cell membrane of cholesterol to inhibit caveolae function, [C] inhibited a 24R,25(OH)₂D₃ induced increase in proliferation at 1 and 10mM, [G] p53 content at 10mM, and [K] TUNEL staining at 0.1-10mM. The anti-ERα36 antibody to block membrane-ERα isoforms also [D] inhibited a 24R,25(OH)₂D₃ increase in proliferation, [H] p53 content, and [L] TUNEL staining. [D, H, and L] To confirm membrane ER involvement, ICI-182780 was included as a control in anti-ERα36 experiments. However, [D] while ICI-182780 did not inhibit 24R,25(OH)₂D₃-induced proliferation, it did inhibit apoptosis as measured by both [H] TUNEL staining and [L] total p53 content. All graphs were analyzed by two-way ANOVA at α=0.05 with Bonferroni’s correction.

Discussion

Vitamin D has been shown to have a significant anti-tumorigenic effect against breast cancer in epidemiological, pre-clinical, cellular and molecular studies. The expanding body of evidence using new synthetic analogs of vitamin D has demonstrated the possibility of developing compounds that can regulate cellular processes independently from their calcemic effects, enabling the expansion of the therapeutic window of vitamin D-related compounds.

We have identified an alternative active endogenous metabolite of vitamin D, 24R,25(OH)₂D₃. 24R,25(OH)₂D₃ has a marked inhibitory action on bone resorption, reducing calcium release and thus not causing hypercalcemia. In the present study, we have established its anti-tumor and pro-apoptotic effects in MCF7 breast cancer cells.
Our results clearly demonstrate an anti-cancer effect indicated by reduced tumor growth, improved prognosis, and longer survival rates. Treatment with 24R,25(OH)2D3 significantly reduced tumor volumes in mice implanted with MCF7 breast cancer xenografts. 17β-estradiol supplementation was required to initiate MCF7 tumor growth in these mice, but after 4 weeks of 17β-estradiol supplementation, tumors continued growing without additional supplementation. Supplementation with 24R,25(OH)2D3 caused a dose-dependent decrease in the rate of tumor growth, with improvement in survival and final tumor volume. Furthermore, increasing concentrations of 24R,25(OH)2D3 correlated with a significant reduction in total metastatic burden, loco-regional and distant metastatic incidence. These effects resemble the correlation described in the literature between increasing serum vitamin D3 and improved survival in breast cancer\textsuperscript{251,286}, and support the hypothesis that 24R,25(OH)2D3 does, in fact, possess anti-tumorigenic properties.

These findings are further supported by the observation that 24R,25(OH)2D3 inhibited expression of metastatic markers, including expression of Snail, Her2, and collagenase, as well as the ratio of CXCR4 to CXCL12. Migration was inhibited in a dose-dependent manner when testing wound closure using the scratch test, and the ratio of OPG to RANKL, both in terms of expression and protein production, was stimulated by high-dose 24R,25(OH)2D3. Total collagenase was not affected to the same extent as MMP1 expression, probably due to the experimental design, where mRNA was assessed at 12 hours and enzyme content was assessed at 24 hours.

In contrast with our in vivo observations, and the evidence shown above that 24R,25(OH)2D3 had an anti-tumor effect, 24R,25(OH)2D3 enhanced proliferation of MCF7
cells \textit{in vitro}. The effect of 24R,25(OH)$_2$D$_3$ on proliferation of MCF7 cells was not stereospecific as a similar response was seen with 24S,25(OH)$_2$D$_3$. Although the precursor 25(OH)D$_3$ had the same stimulatory effect on proliferation as 24R,25(OH)$_2$D$_3$, 1α,25(OH)$_2$D$_3$ had no effect, indicating that the site of the second hydroxyl on either C-24 or C-1 was the important differentiator, but whether the hydroxyl on C-24 was in a R or S configuration did not matter. It is possible that the addition of the C-1 hydroxyl caused a stereospecific inhibition in the binding of the molecular to an as-yet-unidentified receptor.

The study did not take into account, however, that vitamin D3 metabolism occurs in a cyclic manner \textit{in vivo}, both locally in cells that possess appropriate hydroxylases, and in the kidney \cite{13,287}. MCF7 cells normally express 1-hydroxylase, but do not typically express 24-hydroxylase unless stimulated by 1,25(OH)$_2$D$_3$ \cite{288}. It is possible that either local levels or systemic levels of 1α,25(OH)$_2$D$_3$ may have been sufficient to reduce or prevent the proliferative effect of 24R,25(OH)$_2$D$_3$, particularly in response to indirect upregulation of Cyp27B1 and production of 1,25(OH)$_2$D$_3$ due to the 24R,25(OH)$_2$D$_3$ treatment. None of these vitamin D$_3$ metabolites had any effect on cell proliferation after 24 hours of treatment \textit{in vitro}, which could also indicate another undiscovered pathway that counteracts the initial pro-proliferative pathway stimulated by the 15 minute treatment.

Our in vitro studies were performed using MCF7 cells in order to have a direct correlation with our in vivo observations. In order to determine whether our results using this well-studied ER+ breast cancer cell line were also found using other breast cancer cell lines, we examined the responses of ER+ T-47D cells and ER- HCC38 cells to 24R,25(OH)$_2$D$_3$. At least with respect to p53 abundance, the effect of 24R,25(OH)$_2$D$_3$ was not consistent between MCF7, T-47D, and HCC38 cells, suggesting that the response to
this secosteroid may be cell-type dependent. 24R,25(OH)₂D₃ increased p53 content in both MCF7 and T-47D cell lines, which express similar levels of ERα66 and its isoforms. HCC38, on the other hand, expresses high levels of ERα36 and no ERα66. It is possible that each cell line’s ERα profile has a modulating effect on the tumorigenicity of 24R,25(OH)₂D₃. This could provide an explanation for conflicting reports on the tumorigenicity of 24R,25(OH)₂D₃ and 24-hydroxylase. In ERα+ Lewis lung carcinoma cells, 24R,25(OH)₂D₃ was reported to have an anti-tumorigenic effect, but in ERα-ERβ-expressing HT29 colorectal cancer cells, CYP24A1 overexpression enhanced tumor volume, an effect that was further compounded by a diet high in soy-based phytoestrogens. Further studies are ongoing to investigate the universality of 24R,25(OH)₂D₃’s therapeutic effect against breast cancer in other cell types. However, it is clear from our data that 24R,25(OH)₂D₃ has a cell survival modulating effect on breast cancer.

Our results using MCF7 cells indicate that the stimulatory effects of 24R,25(OH)₂D₃ and 24S,25(OH)₂D₃ involved membrane-associated signaling pathways. Pretreatment with 2-bromopalmitate, which blocks palmitoylation of membrane receptors, and inhibition of PLD activity as well as destruction of caveolae structural integrity with β-cyclodextrin abolished their effects. The stimulatory effect of both 24,25(OH)₂D₃ enantiomers on DNA synthesis was only evident in cultures treated for 15 minutes, suggesting a rapid signaling mechanism was involved, although a specific membrane receptor for either the R or S form of the secosteroid might not be required. The effect on DNA synthesis was not evident in cultures treated for 24 hours, raising the possibility that the augmentation of
proliferation was not specific to either enantiomer and might reflect a crosstalk with other membrane or nuclear signaling pathways, as seen extensively in steroid hormones $^{34,68}$.

Treatment with 24R,25(OH)$_2$D$_3$ induced apoptosis in MCF7 cell monolayers as evidenced by elevated levels of Bax/Bcl2 gene expression, total p53 production, and DNA fragmentation (as measured by TUNEL staining). This effect was stereo-specific, as the enantiomer 24S,25(OH)$_2$D$_3$ did not alter Bax/Bcl2 expression, p53 content, or TUNEL staining after treatment for either 15 minutes or 24 hours. The mechanism by which 24R,25(OH)$_2$D$_3$ induced apoptosis is not yet known, although our findings indicate that a membrane-mediated pathway was involved. The 24R,25(OH)$_2$D$_3$-dependent increase in p53 production and TUNEL staining required palmitoylation, PLD activation, and intact caveolae. It should be noted that although the effect of 24R,25(OH)$_2$D$_3$ on apoptosis was stereo-specific and the effect on proliferation was not specific to the 'R' isoform, inhibitor studies suggest that both effects use similar pathways.

Treatment with 24R,25(OH)$_2$D$_3$ not only retarded breast cancer tumor growth, but also lead to the suppression of the primary tumor from metastasizing to other secondary sites. This may be due to an inhibitory effect on the epithelial-to-mesenchymal transition associated with metastasis as previously observed in MCF7 cells $^{194}$. 24R,25(OH)$_2$D$_3$ reduced the expression of several epithelial-mesenchymal transition and tumor aggression-associated factors such as Snail1, which leads to a down-regulation of cell-cell interaction proteins such as cadherins to assist the cell’s transition from a static epithelial cell to a migratory mesenchymal cell $^{289,290}$. 24R,25(OH)$_2$D$_3$ also suppressed migration-associated markers, such as the ratio of gene expression of metastatic factor CXC chemokine receptor type 4 (CXCR4) to its ligand CXC ligand 12 $^{291–294}$, and the
migration of cells in vitro. Matrix metalloproteinases (MMPs), a family of zinc-dependent endopeptidases, which are involved in the degradation of extracellular matrix, have also been associated with cancer invasion, metastasis and angiogenesis. 24R,25(OH)₂D₃ caused a reduction in MMP1 expression. Together with its effect in reducing chemotaxis, as evidenced by reducing the rate of wound closure in a two-dimensional culture, our results indicate that 24R,25(OH)₂D₃ mediates a reduction in breast cancer cell invasion.

Overexpression of Her2, which is regarded as a growth factor receptor oncogene, plays an essential role in the development and progression of certain aggressive types of breast cancer. Expression of Her2 has become an important biomarker and target of therapy for approximately 30% of breast cancer patients, as its downregulation is correlated with growth inhibition of breast cancer cells. Studies with antibodies against Her2 demonstrated improved survival and response rates to chemotherapy. 24R,25(OH)₂D₃ downregulated expression of Her2, supporting its potential as a negative regulator of tumor growth, by inhibiting the ability of breast cancer cells to detach from the primary tumor and migrate to distant sites of the body. This anti-metastatic effect was further observed in vivo, in the reduction of loco-regional and distant metastasis observed in animals treated with 24R,25(OH)₂D₃. Thus, the reduction of migratory factors observed with 24R,25(OH)₂D₃ treatment suggests that it confers a protective effect opposing the growth of breast cancer cells.

The pro-apoptotic effect of 24R,25(OH)₂D₃ was stereo-specific; 24S,25(OH)₂D₃ failed to elicit a response from the cells. This effect was rapid, with 15 minutes of 24R,25(OH)₂D₃ treatment inducing the pro-apoptotic response in cultures harvested 12
to 24 hours later. The effect of 24R,25(OH)$_2$D$_3$ on proliferation was similarly rapid, although it was not stereo-specific. This prompted us to determine the role of the membrane in the 24R,25(OH)$_2$D$_3$ pro-apoptotic response, as membrane-associated actions of 24R,25(OH)$_2$D$_3$ have been shown to affect apoptosis and DNA synthesis in other cell types $^{17,18,93,105,301}$. Pretreatment with methyl β-cyclodextrin, which depletes the membranes of cholesterol and thereby disrupts the caveolae $^{169}$, abolished the pro-apoptotic effect, indicating the importance of the specialized membrane compartment for the 24R,25(OH)$_2$D$_3$-dependent signaling pathway in MCF7 breast cancer cells.

24R,25(OH)$_2$D$_3$ is a steroid hormone, and like 1α,25(OH)$_2$D$_3$ and ERα, acts by a set of membrane-associated mechanisms to exert its effect. Steroid hormone receptors are trafficked to the plasma membrane to serve as transmembrane receptors via a uniform mechanism involving palmitoylation $^{35}$, the attachment of palmitic acid to an internal cysteine residue, which promotes the association of the receptor with caveolin-1, a transporter protein, to caveolae rafts in the plasma membrane $^{49}$. Palmitoylation plays a role in signaling through the membrane-associated ERα $^{78,302}$. The canonical vitamin D receptor (VDR), which binds 1α,25(OH)$_2$D$_3$, can act through rapid membrane associated signaling pathways $^{39,254}$. Our results show that the apoptotic effects of 24R,25(OH)$_2$D$_3$ require palmitoylation, as they were blocked by 2-bromopalmitate, which is an inhibitor of palmitoylacetetyl-transferase and prevents palmitoylation of steroid hormone receptors $^{269}$.

Previous studies in our lab using chondrocytes show that 24R,25(OH)$_2$D$_3$ increases proliferation (DNA synthesis) and differentiation while inhibiting apoptosis $^{303}$. These actions involved PLD-mediated increases in PKC, as well as through inhibition of the phospholipase A$_2$ (PLA$_2$) pathway $^{15,16,255}$. It has been suggested that the point of
regulation in the mechanism of 24R,25(OH)$_2$D$_3$ stimulated PKC is PLA$_2$, which controls the availability of substrate for Cox-1 $^{17}$. Further, the activation of PLD resulted in increased production of lysophosphatidic acid (LPA) and consequently LPA-mediated proliferation, maturation, inhibition of phosphate-induced apoptosis, and reduction of p53 $^{18,92,93}$. Both genomic and non-genomic signaling pathways have been implicated in the actions of 24R,25(OH)$_2$D$_3$ and it has been suggested that it exerts its effects via more than one signaling pathway. These pathways may be interrelated via the modulation of PLA$_2$ $^{17}$. In our study, both the pro-apoptotic and proliferative effects incurred by 24R,25(OH)$_2$D$_3$ in MCF7 breast cancer cells were abolished by pretreatment with wortmannin, a non-specific inhibitor of PLD $^{15,142}$. Thus, a PLD mediated signaling pathway is implicated in both 24R,25(OH)$_2$D$_3$’s pro-apoptotic and proliferative effects in breast cancer cells.

The full impact of 24R,25(OH)$_2$D$_3$ is the result of the convergence and crosstalk of the different signaling pathways, which at times are antagonistic, and its interaction with other steroid hormones such as other vitamin D metabolites $^{18}$ or sex hormones. However, it is important to note that wortmannin is also a weak inhibitor of PKC and PI3K, which have both been shown to be involved in rapid signaling by steroid hormones and resultant regulation of proliferation and apoptosis $^{31,68}$. Further investigation is necessary to determine whether this effect is cell type specific or if the PLD, PLA$_2$ mechanism shown in chondrocytes universally governs 24R,25(OH)$_2$D$_3$’s apoptotic effects. 24R,25(OH)$_2$D$_3$ function is variable and complex and necessitates further research to clarify the signaling pathways involved and to clarify the effects of 24R,25(OH)$_2$D$_3$ on breast cancer cells.
Finally, we assessed the relationship between the effect of 24R,25(OH)$_2$D$_3$ and ER$\alpha$, the classical nuclear receptor for the sex steroid 17$\beta$-estradiol (E$_2$). A known correlation between 1$\alpha$,25(OH)$_2$D$_3$ and E$_2$ exists. 1$\alpha$,25(OH)$_2$D$_3$ downregulates E$_2$ receptor abundance by binding to VDR and activating negative response elements in the ESR1 promoter $^{120}$; suppresses E$_2$ actions in MCF7 breast cancer cells $^{120,304}$; and restores antiestrogen responsiveness $^{129}$. Our findings stress a striking resemblance in the signaling pathways of the two steroid hormones despite having antagonist actions in breast cancer cells. E$_2$ signaling is pivotal in the initiation and progression of breast cancer; it functions as a potent mitogen inducing the activation of c-Myc and an array of cyclin genes promoting cell cycle progression from G1 to S phase $^{305}$. The novel membrane receptor ER$\alpha$36 is gaining interest as a key player in breast cancer, especially in conferring aggressive features and resistance to hormonal treatment.

ER$\alpha$36 mediates the anti-apoptotic effect of E$_2$ by activating PLD at the membrane leading to activation of LPA signaling and PI3K, which in turn leads to attenuation of the caspase cascade that promotes apoptosis $^1$. E$_2$-ER$\alpha$36 signaling has also been implicated in the activation of other rapid pathways such as ERK-MAPK, PI3K-Akt, and PKC-ERK, and may have downstream effects on the employment of IGF-IR, EGFR, HER2 $^{306}$, resulting in a decrease in apoptosis and an increase in proliferation and metastatic potential in a variety of cancer cells, including breast cancer cells $^{68,109}$. These pathways have been reported to stimulate tumorigenicity in both ER positive and negative breast cancer tumors $^{27,55}$. These rapid signaling mechanisms mediated by ER$\alpha$36 are insensitive to anti-estrogens and have even been shown to employ agonist activities in response to anti-estrogens, such as tamoxifen and ICI-182,780, which are both known
inhibitors of the classical ERα66\textsuperscript{129}. Both antibodies that block the ERα36 mediated effect of E\textsubscript{2} and ICI 182,780 (which blocks cytosolic and nuclear ERs) abolished the pro-apoptotic effect of 24R,25(OH)\textsubscript{2}D\textsubscript{3}. However, only the antibody to ERα36 blocked the proliferative effect. This implies that 24R,25(OH)\textsubscript{2}D\textsubscript{3} and both the canonical and membrane-associated signaling pathways of E\textsubscript{2} share a common downstream pathway, the exact mechanism of which remains to be revealed (Figure 5.11).

Upon activating a signaling cascade at the membrane, 24R,25(OH)\textsubscript{2}D\textsubscript{3} stimulates the secondary messenger PLD. PLD stimulates the LPAR1/3 receptor and downstream signaling stimulates PI3K, upregulates Bax, and downregulates Bcl2 expression to increase p53 production and induce apoptosis in MCF7 cells. A similar mechanism is responsible for the proliferative actions of 24R,25(OH)\textsubscript{2}D\textsubscript{3}, which is regulated by a membrane-mediated receptor in the caveolae niche via a PLD-dependent mechanism. 24R,25(OH)\textsubscript{2}D\textsubscript{3} also decreases invasion and metastatic markers through an as yet unidentified mechanism.
Figure 5.11 24R,25(OH)2D3 induces apoptosis and proliferation through a membrane-mediated caveolae-dependent mechanism involving one or more as yet undescribed palmitoylated nuclear receptors.
Chapter 6.

24R,25-dihydroxyvitamin D₃ regulates breast cancer progression in an estrogen receptor α 66-dependent manner.

[Introduction]

Vitamin D₃ has been of interest as an anti-tumorigenic agent for decades. Interest in vitamin D₃ first peaked when epidemiologists noticed that the incidence of certain cancers, including breast cancer, inversely correlated with daily sun exposure ⁹⁻¹⁰. Vitamin D₃ is an essential pro-hormone that is synthesized in response to UVB radiation from sunlight. Retrospective clinical studies have shown that reduced exposure to UVB correlates with low levels of serum vitamin D₃, and low serum vitamin D₃ correlates with an increased risk for breast cancer ¹¹. Similarly, epidemiological studies have shown vitamin D₃ supplementation is associated with a reduced risk of breast cancer recurrence ². However, recent large-scale double-blind clinical trials have concluded that there is no connection between vitamin D₃ supplementation and breast cancer incidence in the general population ⁶,⁷.

There are two possible reasons for this discrepancy. One reason may be that most clinical studies that examine the efficacy of vitamin D₃ treat breast cancer as a homogenous disease with uniform presentation. In actuality, breast cancer is a heterogeneous disease that can present with several distinct molecular subtypes.
Detailed clinical and retrospective analyses have shown that low serum vitamin D$_3$ is associated with an increased risk of developing estrogen receptor positive (ER+) cancer, but not estrogen receptor negative (ER-) cancer\(^{309}\). Furthermore, vitamin D supplementation is associated with a reduced risk of developing new or recurrent tumors in patients with estrogen receptor positive (ER+) cancer, but not patients with estrogen receptor negative (ER-) cancer\(^{8,310}\).

Another reason may be that most studies that examine the anti-cancer properties of vitamin D$_3$ limit their investigations by correlating cancer incidence with oral vitamin D$_3$ intake or serum 25-hydroxyvitamin D$_3$ (25(OH)D$_3$)\(^{112,240,311}\). Serum 25(OH)$_2$D$_3$ is an incomplete picture of the active vitamin D metabolome. Once ingested, vitamin D$_3$ is metabolized in the liver into 25(OH)D$_3$\(^{246}\). From there it circulates in the blood as serum 25(OH)D$_3$ until it reaches the kidney, where it is hydroxylated into a number of active vitamin D$_3$ metabolites, including the well-described metabolite 1$\alpha$,25-dihydroxyvitamin D$_3$ (1$\alpha$,25(OH)$_2$D$_3$) and the understudied 24R,25-dihydroxyvitamin D$_3$ (24R,25(OH)$_2$D$_3$)\(^{13}\). 1$\alpha$,25(OH)$_2$D$_3$ is partially responsible for maintaining calcium homeostasis and bone mineralization and has been shown to have some anti-cancer properties. However, it has a narrow therapeutic window and can induce hypercalcemia. 24R,25(OH)$_2$D$_3$ is a less studied, less calcemic vitamin D$_3$ metabolite. Previous studies in our lab and others have shown that 24R,25(OH)$_2$D$_3$ is responsible for maintaining cartilage differentiation and may regulate the cell cycle in chondrocytes\(^{23,93,312}\).

24,25(OH)$_2$D$_3$ has been shown to have anti-tumorigenic properties in certain cancers\(^{260}\), including lung and stomach carcinomas\(^{260,262}\). However, an increase in 24-hydroxylase, the enzyme that increases 24R,25(OH)$_2$D$_3$ \textit{in vivo}, has been associated with
increased tumorigenicity in models of colorectal carcinoma. Recently, our lab has shown that 24R,25(OH)_{2}D_{3} is active in breast cancer cell lines and that 100 ng of 24R,25(OH)_{2}D_{3} given three times a week is non-calcemic and anti-tumorigenic in a model of ER+ breast cancer. However, while 24R,25(OH)_{2}D_{3} was anti-tumorigenic in ER+ breast cancer cell lines, it was pro-tumorigenic in HCC38, an ER- breast cancer cell line. In vitro studies revealed that the actions of 24R,25(OH)_{2}D_{3} are modulated by a rapid, membrane-associated, mechanism in breast cancer.

Breast cancer cells that are ‘ER+’ express the classical estrogen receptor α (ERα) isoform ERα66, while ‘ER-’ cells do not express ERα66. Both cell types may express other ERα isoforms, including the membrane isoform ERα36. HCC38 does not express ERα66, however, it expresses ERα36. ERα36 mediates rapid signaling by 17β-estradiol (E\textsubscript{2}) in cancer cells. We hypothesized that the pro-apoptotic effects of 24R,25(OH)_{2}D_{3} might be regulated in part by the canonical receptor, ERα66, rather than the rapid signaling receptor, ERα36, thus implying a role for the canonical receptor ERα66 in the rapid signaling of 24R,25(OH)_{2}D_{3}.

**Materials & Methods**

**Cell Lines**

MCF7 and HCC38 breast cancer cell lines and C2C12, a muscle cell line, were purchased from ATCC (Manassas, VA). Experiments were conducted within 6 months of cell purchase. Cells were maintained in media and passaged following ATCC guidelines as previously described.
Caveolin-1 knockdown.

Stable knockdown of Cav-1 was achieved by transducing wild-type HCC38 cells with CAV-1 shRNA lentiviral transduction particles packaged in Mission® shRNA lentiviral particles as previously described. Four shRNAs targeted to the CAV1 gene (Millipore-Sigma, Burlington, MA, cat#SHCLNV-NM_0001753, clone TRCN0000000310, clone TRCN0000000508, clone TRCN00000000508 and clone TRCN0000008002) were cloned into a cassette containing a puromycin-resistance gene and packaged as lentiviral particles. Empty control cells were created with MISSION® pLKO.1 puro Empty Vector Control Transduction Particles (Sigma MISSION® cat#SHC001V). HCC38 cells were transduced with shRNA-containing lentiviral particles according to manufacturer’s protocol and a pure population of cells was enriched by using 50μg/mL as a selection agent. CAV1-knocked down-HCC38 cells were tested for effective Caveolin-1 knockdown, and the two clones with the best knockdown, clone TRCN0000000508 and clone TRCN0000008002, were chosen for subsequent experiments and labeled “clone 1” and “clone 2” respectively. Caveolin-1 knockdown was confirmed with western blot as previously described. Antibodies for all western blots are given in Table 6.1. All transduced-HCC38 cells were maintained in full media supplemented with 50μg/mL puromycin.

ERα isoform overexpression in cell lines

ERα isoforms were overexpressed in HCC38 and C2C12 cells with a commercially available ESR1, GFP-tagged open-reading-frame (ORF) plasmid (Origene, cat# RG213277) as previously described. Empty vector controls were created with open
reading frame (ORF) control particles (Origene, cat# PS100093) according to Origene protocols.

HCC38 monolayer cultures were transiently transduced for 48 hours with the ESR1 or control particles according to manufacturer’s instructions. In brief, a solution of 5μg of plasmid per 1mL of opti-mem serum-free media per 6μL turbofectin 8.0 (Origene, cat# TF81001) was added drop-wise to T25 (1mL plasmid solution/well), 24W (100μL/well) or 48W (50μL/well) monolayer cultures. Transfection was confirmed 48 hours after particle addition by visualizing GFP expression in wells with a Zeiss SteroDiscovery.V12 fluorescence microscope with an AxioCam MRm digital camera. In a representative set of cultures, 48 hours after transient transfection a T25 culture of cells was harvested with radioimmunoprecipitation assay (RIPA) buffer and ERα66 overexpression was confirmed by western blot as described previously.

C2C12, a mesenchymal cell line derived from myoblasts, were stably transfected with ESR1 or empty control cells using the ORF particles described above. 48 hours after transfection, cells were selected with full media supplemented with 1mg/mL G418 as previously described. ESR1 overexpression was confirmed by western blot as described previously.

ERα66 silencing in MCF7 cell lines

ERα66 was stably knocked down in MCF7 cells with a commercially available short hairpin ESR1 lentiviral vector from Sigma MISSION® (cat# TRCN0000003301). Scrambled control cells were created with MISSION® TurboGFP® Control Transduction Particles (Sigma MISSION® cat#SHC016V). Both vectors included a coding region for a puromycin resistance gene. Cells were transduced according to manufacturer’s protocol.
and a pure cell population was enriched using 50μg/mL puromycin selection media. Cells were maintained in 50μg/mL puromycin full media and ERα66 silencing was confirmed via western blot as previously described\textsuperscript{169}.

**Table 6.1 Antibodies used in western blots.**

<table>
<thead>
<tr>
<th>Protein</th>
<th>Source</th>
<th>Catalog #</th>
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<tr>
<td>Caveolin-1</td>
<td>Abcam</td>
<td>Ab2910</td>
</tr>
<tr>
<td>ERα66</td>
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<td>8-80119</td>
</tr>
<tr>
<td>ERα36</td>
<td>Chi Scientific</td>
<td>8-80119</td>
</tr>
<tr>
<td>GAPDH</td>
<td>Millipore-Sigma</td>
<td>MAB374</td>
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</table>
In Vivo Studies

All in vivo experiments were done on female 6-8 week old Nod-Scid-gamma IL2γ (NSG) mice obtained from the Virginia Commonwealth University (VCU) Cancer Mouse Models Core Laboratory at Massey Cancer Center (Richmond, VA). All animal experiments were approved by VCU’s Institutional Animal Care and Use Committee. Animals were monitored for signs of hypercalcemia and other health and behavioral changes throughout the course of the study.

Mammary fat pad xenograft model.

Two mammary fat pad xenograft models were tested for this study. The first study was done with un-operated mice. A second study was conducted at a later date with ovariectomized mice. 5-6 week old female NOD.Cg-PrkdcscidIl2rgtm1Wjl/SzJ (NSG) mice were ovariectomized as previously described 167. Two weeks after ovariectomy, tumors were initiated as previously described313. In brief, intact or ovariectomized NSG mice were injected twice weekly intraperitoneally with 2.5mg/kg of 17β-estradiol (n=6) (Millipore-Sigma cat#E8875) or vehicle (n=6) (2% ethanol:1X distilled phosphate buffered saline [DPBS]) starting at 1 week prior to tumor initiation and continuing until the end of the study 10 weeks later. To initiate tumors, 1 million HCC38 cells in 50μL of 1:1 DPBS:BME (Cultrex Pathclear basement membrane extract Type 3, 14.5 mg/mL; #3632; RnD Systems, Minneapolis, MN) were injected into the lactiferous duct of the right fourth mammary gland of 5-6 week old NSG mice as previously described 313. Tumors were measured with digital calipers starting at 2 weeks post cell injection and continued to be measured once a week until harvest at 10 weeks.
Effect of 24R,25(OH)₂D₃ on xenograft tumor growth.

One million HCC38 cells in 50μL of 1:1 DPBS: BME were injected into the lactiferous duct of the right fourth mammary gland of un-operated female NSG mice to initiate tumors as described in the Error! Reference source not found.. The next day, mice were injected with 0, 25 ng, or 100 ng of 24R,25(OH)₂D₃ dissolved in 2% ethanol:1X DPBS. Injections continued 3 times a week for the remainder of the study. Tumors were measured with digital calipers, and tumor volume was calculated as previously described. After 8 weeks, mice were euthanized and tumors were harvested as described above. Immediately after harvesting, tumors were scanned for μCT analysis and then fixed in 10% neutral-buffered formalin. μCT tumor measurements were normalized to vehicle-treated tumors and compared to μCT measurements taken from MCF7 tumors from a previous study and similarly analyzed to assess changes in tumor volume relative to their respective controls. Animal survival was assessed with a Mantel-Cox logrank survival curve analysis, and P-values <0.05 were considered significant as previously described.

In vitro Studies

Response to vitamin D₃ metabolites.

24R,25(OH)₂D₃, and 1α,25(OH)₂D₃ were purchased from Enzo Life Sciences (Farmingdale, NY); 24S,25(OH)₂D₃ was purchased from Millipore-Sigma (cat# 29447). Stock solutions were prepared in absolute ethanol and diluted in culture medium as previously described.

Confluent cultures were treated with dose-dependent 24R,25(OH)₂D₃ (10⁻¹⁰-10⁻⁷M) as previously described. To assess specificity of the response to 24R,25(OH)₂D₃,
confluent cultures were also treated with 1α,25(OH)₂D₃ (10⁻¹⁰-10⁻⁸M), 24S,25(OH)₂D₃ (10⁻⁹-10⁻⁷M), or 25(OH)D₃ (10⁻⁹-10⁻⁷M), or a vehicle (10⁻⁴M ethanol). All vitamin D metabolites were reconstituted in ethanol, as previously described.¹⁵ Stock solutions were diluted in warm complete media to final treatment concentrations. Media were removed by aspiration at 15 min or 24 hours, and cells were incubated with fresh complete media until harvest as indicated below.

Signaling Pathway Inhibition.

In these experiments, cells were pre-treated for 30 minutes with inhibitors or vehicle in warm complete media as previously described.³¹³ In brief, cultures were pretreated with 1-10μM 2-bromopalmitate (Millipore-Sigma #21604)²⁶⁹ to inhibit palmitoylation, 0.1-1 μM wortmannin (VWR, Radnor, PA, cat# 80055508)¹⁵ to inhibit PLD, or 1-10mM methyl β-cyclodextrin (Millipore-Sigma #C4555) in serum-free media to deplete cell membranes of cholesterol and inhibit intact caveolae.¹⁶⁹,²⁷⁰,²⁷¹ Cultures were pretreated with 10⁻⁷M ICI-182780, an inhibitor to cytosolic and nuclear ERs (Millipore-Sigma #I4409)²⁷²,²⁷³, or 1μg of a neutralizing antibody to the membrane-associated estrogen receptor (Alpha Diagnostic, San Antonio, Texas, cat#ERA361-A), to assess the role of ERα66 or ERα36 respectively on the actions of 24R,25(OH)₂D₃.⁵⁵

Outcome Measures

DNA synthesis

Cells were cultured to 70% confluence, then serum-starved for 48 hours as previously described.³¹³ At that time, cells were treated with 24R,25(OH)₂D₃, 24S,25(OH)₂D₃, or 1α25(OH)₂D₃ for 15 minutes and incubated with fresh complete media for 24 hours, harvested, and assayed for EdU incorporation according to manufacturer’s
instructions (ThermoFisher Scientific, Waltham, Massachusetts, #C10214) as previously described 313.

**Gene expression**

Cells were cultured to confluence, then treated with 24R,25(OH)₂D₃ or vehicle in complete media for 15 minutes as described above. Cultures were then aspirated, incubated with complete media for 12 hours. RNA was extracted using TRIzol (Invitrogen, Carlsbad, CA) and quantified (Take3 Micro-Volume Plate, Biotek, Winooski, VT) before being used to synthesize cDNA libraries (High Capacity cDNA Reverse Transcription kit, Promega, Madison, WI) and assessed for gene expression using qRT-PCR as previously described 313. Gene-specific primers were designed as previously described and are given in Table 6.2 313. HCC38, a triple-negative breast cancer cell line that does not express ERα66 1 but does express ERα36, and MCF7, a luminal breast cancer cell line that expresses all ERα isoforms 1, were both assessed for hydroxylase enzyme mRNA expression.
Table 6.2 Gene expression primers used in quantitative real-time PCR.

<table>
<thead>
<tr>
<th>Species</th>
<th>Gene</th>
<th>Primer Sequence</th>
</tr>
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<td>F GACGAACTGGACAGTAACATGG</td>
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<td></td>
<td></td>
<td>R AAAGTAGAAAAGGGCGACAACC</td>
</tr>
<tr>
<td>Human</td>
<td>BCL2</td>
<td>Hs_BCL2-1-SG QuantiTect Primer #249900</td>
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<tr>
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<td>BCL2</td>
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**p53 content**

Total p53 was measured 24 hours post-cell treatment in human cells with 24R,25(OH)2D3 as described above using a sandwich enzyme-linked immunosorbent assay (ELISA) (R&D Systems, cat# DYC1043) and phosphorylated p53 was measured using a sandwich ELISA specific to the serine-15 residue (R&D Systems, cat# DYC1839). Total p53 content was measured in C2C12 cells with a p53 pan ELISA (Millipore-Sigma cat# 11828789001). For all cell types, cells were plated in 24 well plates, treated with
24R,25(OH)₂D₃ for 15 minutes, and incubated for 24 hours in complete media. Conditioned media were collected, and cell layers were lysed by sonication (40A for 5 seconds) in 200μL lysis buffer according to manufacturer's protocol. Lysates were centrifuged at 10,000g for 15 minutes; the supernatant was collected and assayed for total protein content (ThermoFisher, Pierce 660 nm Protein Assay) and total p53 content following the manufacturer’s instructions. Data are presented as a ratio of nanograms of total p53 to total protein content.

**DNA fragmentation**

DNA fragmentation was assessed by colorimetric terminal deoxynucleotidyl transferase (TdT)-mediated dUTP nick end labeling (Trevigen, TiterTAC™ in situ microplate TUNEL assay). Cells were incubated with vehicle or 24R,25(OH)₂D₃ for 15 minutes, followed by incubation in fresh media for 24 hours. Samples were harvested and processed for TUNEL staining following the manufacturer’s instructions ³¹³.

**Enzyme-linked immunosorbent assays for secreted proteins**

Confluent cultures cells were plated in 6 well plates, treated with vehicle or 24R,25(OH)₂D₃ for 15 minutes as described above, aspirated, and then incubated in complete media. After 24 hours, conditioned media were collected in 5mL polypropylene tubes, frozen at -80°C and lyophilized (Labconco, Kansas City, Missouri, Freezone Freeze Dryer #7740020). Lyophilized samples were reconstituted in 200 μL of filtered 1XPBS containing 1% BSA and assayed for OPG and RANKL using sandwich ELISAs (R&D Systems, Human TRANCE/RANK L/TNFSF11 DuoSet ELISA #DY626, Human Osteoprotegerin/TNFRSF11B DuoSet ELISA #DY805) as previously described ³¹³. Cell monolayers were lysed in 1mL 0.5% Triton™ X-100. Cell lysates were assayed for total...
DNA content (Promega, QuantiFluor dsDNA, #E6150). OPG and RANKL content are expressed as a ratio of OPG/RANKL.

**Collagenase**

Collagenase content in the cultures was measured using an MMP1 assay kit (SensoLyte® Plus 520 MMP-1 Assay Kit *Fluorimetric and Enhanced Selectivity*, Anaspec #AS-72012). Cells were plated in 24 well plates and treated with vehicle or 24R,25(OH)2D3 as described above. At harvest conditioned media were collected; cell monolayers were lysed in 500μL 0.05% Triton-X-100 and total DNA content measured. Total MMP1 was measured in the media and normalized to total DNA content. Data are presented as a ratio of ng MMP1/μg DNA.

**Osteoclast Activity**

Osteoclast (OC) activity was measured using a commercially available europium-conjugated human type I collagen-coated OsteoLyse™ Assay Kit, as previously described. Human OC precursors (OCPs; Lonza Biosciences, Walkersville, MD, cat# 2T-110) were cultured on a human type I collagen-coated, 96-well OsteoLyse™ Assay Kit (Lonza Biosciences). OCPs were differentiated into mature OCs using osteoclast precursor growth medium (OCGM, Lonza Biosciences, cat# PT-8001) supplemented with 33ng/mL macrophage colony stimulating factor (M-CSF) and 66ng/mL RANKL as previously described for 7 days. OCPs were also cultured in OCGM supplemented with M-CSF without RANKL for use as a negative control.

HCC38 cells were plated in 6 well plates. At confluence, cultures were treated with vehicle or dose-dependent 24R,25(OH)2D3 for 15 minutes as described above. 24 hours after treatment, conditioned media were collected. OC cultures were aspirated and
treated with conditioned media. OCs were cultured for another 7 days in conditioned media. After 7 days of culturing with HCC38-conditioned media (14 days total), OC activity was measured using the OsteoLyse™ Assay Kit, as previously described

**Wound closure assay**

A wound closure assay was used to demonstrate metastasis. Confluent cultures of HCC38 cells were treated for 15 min with media containing vehicle or 24R,25(OH)2D3. Another set of cultures was treated with 10^-7M 17β-estradiol as a positive control. Following treatment, a scratch was drawn from one end of the well to the other using a sterile 1 mL pipette tip. The loosened cells were aspirated along with the treatment media, and fresh media added to the cultures. A phase-contrast microscope was used to take pictures of each well at 0, 3, 6, 9, 12, and 24 hours after the scratch was made. Wound area was calculated using Adobe Photoshop™ 2 software, normalized to the original wound closure area, and presented as percent wound closure. Data presented are from one of two repeated experiments.

**Enzyme Activity Assays**

Cells were plated in 24-well plates and treated with dose-dependent 24R,25(OH)2D3 as described above for 9 minutes. Immediately after treatment, cells were aspirated, washed twice with 1X DPBS, and harvested with 200μL/well of phospholipase D (PLD) or protein kinase C (PKC) assay buffer according to manufacturer’s instructions. PLD activity was measured using the Amplex Red PLD assay (Fisher Scientific, Hampton, NH, cat# A12219) as previously described. PKC activity was measured using the PKC Activity Kit (Abcam, Cambridge, UK, cat# ab139437) as previously described.
Statistical Analysis

Data shown are from single representative experiments of two or more repeats and are presented as the mean ± standard error of six independent cultures per treatment group. Data were analyzed by ANOVA with Tukey’s post-test to determine significance between groups. Animal studies were analyzed with a repeated-measures two-way ANOVA with $\alpha = 0.05$. Tumor growth curves were analyzed with linear regression analysis to determine whether tumor growth was linear and non-zero. μCT data were analyzed with a one-way ANOVA within xenograft model types (i.e., MCF7 and HCC38 experiments were analyzed separately). Animal survival was assessed with a Kaplan-Meier log-rank analysis.

*In vitro* experiments with two independent variables (DNA synthesis, total DNA content, and total p53 time courses) were analyzed with two-way ANOVA with Bonferroni’s modification of Student’s t-test post-tests to determine the variance between row and column factors. The wound closure assay was analyzed with a repeated measures ANOVA test with comparisons between time points. P-values <0.05 were considered significant. Treatment over control data were calculated as the mean fold change of treated groups as compared to vehicle-treated cultures (dashed lines) from three or more experiments and analyzed with Wilcoxon signed-rank non-parametric paired t-tests as compared to control. All other experiments were analyzed with a one-way ANOVA with Tukey’s correction. All statistics were conducted in Graphpad Prism 6™. For all experiments, P-values < 0.05 were considered significant.
Results

*Breast cancer cell lines differentially express 24-hydroxylase enzyme.*

HCC38 expressed both 24-hydroxylase (CYP24A1) and 1α-hydroxylase (CYP27B1) (Figure 6.1A), while MCF7 did not express CYP24A1. Both HCC38 and MCF7 expressed similar levels of CYP27B1 (Figure 6.1B).

*17β-Estradiol supplementation is not required to support HCC38 mammary fat pad xenografts in NSG mice.*

HCC38 xenografts were created in the 4th mammary fat pad of un-operated or ovariectomized NSG mice given systemic 17β-estradiol (E2) or a vehicle. E2 did not significantly alter HCC38 tumor burden until week 8 (ovariectomized mice) or week 9 (un-operated mice) (Figure 6.1C-6.1D). At weeks 9, 11, and 12, E2 statistically increased HCC38 tumor burden in un-operated mice. In ovariectomized mice, E2 statistically decreased tumor burden at week 8, but did not significantly alter tumor burden at any other time point in the study. Tumors grown in mice with intact ovaries (un-operated mice) were slightly larger at 3 weeks in un-operated mice (~50-75 mm³) as compared to ovariectomized mice (~30 mm³). This slight increase in tumor burden in un-operated mice continued throughout both studies, with tumors in un-operated mice being slightly larger (~200 mm³) than tumors in ovariectomized mice (~150 mm³) at the end of the study at 10 weeks (*Error! Reference source not found.* C-6.1D).

Tumors grew at a linear rate in vehicle-treated ($R^2=0.7857$) and E2-treated ($R^2=0.7638$) un-operated mice with slopes of $2.660 \pm 0.1824$ mm³/day and $3.206 \pm 0.2341$ mm³/day, respectively. The slopes of the vehicle-treated and E2-treated tumors were significantly different from each other with E2-treated tumors growing faster than vehicle-
treated tumors in un-operated mice. In ovariectomized mice, tumors also grew at a linear rate with vehicle-treated ($R^2=0.8257$) and E$_2$-treated ($R^2=0.7916$) tumors having slopes of $2.579 \pm 0.1767$ mm$^3$/day and $2.034 \pm 0.1539$ mm$^3$/day respectively. The slopes of the vehicle-treated and E$_2$-treated tumors were also significantly different from each other, but E$_2$ treated tumors grew more slowly than vehicle-treated tumors in ovariectomized mice. A linear regression analysis found that both vehicle and E$_2$-treated tumors had significantly non-zero slopes in both un-operated and ovariectomized mice.

*Treatment with 24R,25(OH)$_2$D$_3$ increased tumor burden in an HCC38 xenograft model of breast cancer.*

Mice receiving either 25 ng or 100 ng per injection of 24R,25(OH)$_2$D$_3$ had a greater HCC38 tumor burden than mice given a vehicle control injection (Figure 6.1E). This increase in tumor volume in 24R,25(OH)$_2$D$_3$ treated mice began at week 6 and continued until harvest at week 8. In previous HCC38 mammary fat pad xenograft studies, vehicle-treated HCC38 tumors reached a final volume of 150-200 mm$^3$ (Figure 6.1C, 6.1D), consistent with vehicle-treated HCC38 tumors in this study. Tumors in mice given 100ng of 24R,25(OH)$_2$D$_3$ per injection were significantly larger than tumors in mice given 25 ng at weeks 7 & 8. At harvest at week 8, tumors in mice given 100 ng were ~33% larger (final volume ~400 mm$^3$) than tumors in mice given 25ng of 24R,25(OH)$_2$D$_3$ per injection, which reached a final volume of ~300 mm$^3$. As in previous HCC38 mammary fat pad xenograft studies, HCC38 tumors were linear with non-zero slopes of $1.635 \pm 0.4265$ mm$^3$/day (vehicle), $5.48 \pm 0.4642$ mm$^3$/day (25 ng 24R,25(OH)$_2$D$_3$) and $7.575 \pm 1.241$ mm$^3$/day (100 ng 24R,25(OH)$_2$D$_3$). All three slopes were significantly non-zero and different from each other. Treatment with high-dose 24R,25(OH)$_2$D$_3$ increased tumor growth rate and
tumor burden in HCC38 xenografts by approximately 50%, as compared to a 50% decrease in tumor burden by 24R,25(OH)₂D₃ in MCF7 xenografts as measured by μCT (Error! Reference source not found. Figure 6.1F). Signs of hypercalcemia, such as anorexia, vomiting, lethargy, fatigue, and polyuria, were not detected in animals treated with a vehicle or with either dose of 24R,25(OH)₂D₃. In the high-dose 24R,25(OH)₂D₃ HCC38 xenograft group, only 6 out of 8 animals survived to the end of the study as compared to 8 out of 8 animals in the vehicle and low-dose 24R,25(OH)₂D₃ groups. However, this reduction in animal survival was not statistically significant (Figure 6.1G).
Figure 6.1 Breast cancer cell lines differentially expression hydroxylase enzymes

HCC38 and MCF7 breast cancer cells were cultured to confluence and harvested after 12 hours for gene expression. Cells were assessed for [A] CYP24A1 or [B] CYP27B1 expression and normalized to GAPDH as a housekeeping gene. [C] Unoperated female NSG mice implanted with HCC38 xenograft mammary fat pad tumors were intraperitoneally injected with 2.5mg/kg 17β-estradiol or a vehicle.
twice a week for 12 weeks. Tumor burden was measured twice a week throughout the course of the study and plotted against time. [D] A similar study was done with ovariectomized NSG mice. HCC38 xenografts were treated with the same dose (2.5mg/kg) or 17β-estradiol) or a vehicle and tumors were measured twice a week until harvest at 10 weeks. *indicates statistical significance against week-matched vehicle tumors at α=0.05 with two-way ANOVA. [E] NSG mice with HCC38 mammary fat pad xenograft tumors were intraperitoneally injected with 25 or 100 ng of 24R,25(OH)₂D₃ 3 times a week for 8 weeks. Tumor burden was measured with digital calipers and plotted against time. Animals given 24R,25(OH)₂D₃ had increased tumor burden as measured by digital calipers. *indicates significance as compared to vehicle-treated animals within the same time point. o indicates significance as compared to animals treated with 25 ng of 24R,25(OH)₂D₃ within the same time point. [F] Animals given 24R,25(OH)₂D₃ had increased tumor burden as measured by μCT as compared to tumor burden in vehicle-treated mice. Capital letters were used to indicate significance within HCC38 xenograft tumor groups, while lower-case letters were used to indicate significance within MCF7 tumor groups. Groups that do not share a letter are statistically significant. [G] After 8 weeks, 8 animals from the vehicle group, 8 animals from the low-dose (25 ng per injection) 24R,25(OH)₂D₃ group, and 6 animals from the high-dose (100 ng per injection) 24R,25(OH)₂D₃ group survived out of an original n of 8. Animals in the high-dose group did not have a statistically significant reduced survival rate as compared to animals in low dose or vehicle groups.
24R,25(OH)₂D₃ specifically induced proliferation in HCC38 cells in vitro.

When HCC38 cells were treated for 15 minutes with 24R,25(OH)₂D₃ proliferation increased after 24 hours (Figure 6.2A) as measured by DNA synthesis. However, treatment with 24S,25(OH)₂D₃, an enantiomer of 24R,25(OH)₂D₃, or 1α,25(OH)₂D₃ for 15 minutes did not change proliferation after 24 hours (Figure 6.2B, C). The effect of 24R,25(OH)₂D₃ was dose-dependent with the greatest increase at 10⁻⁷M 24R,25(OH)₂D₃. The effect of 24R,25(OH)₂D₃ was also observed in MCF7 cells, with both 10⁻⁸ and 10⁻⁷ 24R,25(OH)₂D₃ increasing DNA synthesis (Figure 6.2D). In both HCC38 and MCF7 cells, 10⁻⁷M 24R,25(OH)₂D₃ increased DNA synthesis ~two-fold above vehicle-treated cultures.
Figure 6.2: 24R,25(OH)2D3 specifically induces proliferation in HCC38 cells.

HCC38 monolayer cultures were serum-starved for 48 hours, treated with 24R,25(OH)2D3, 24S,25(OH)2D3, or 1α,25(OH)2D3 for 15 minutes and assayed 24 hours later for EdU incorporation. [A] 24R,25(OH)2D3, but not [B] 24S,25(OH)2D3 nor [C] 1α,25(OH)2D3, induced proliferation in HCC38 cell monolayers 24 hours after a 15-minute treatment. [D] Treatment over control was calculated as the average fold-change of treatment as compared to vehicle-treated cultures in three or more experiments and analyzed with a paired t-test against normalized vehicle controls *indicates significance at P<0.05 as compared to vehicle-treated cultures. High-dose 24R,25(OH)2D3 induced proliferation in HCC38 and MCF7 cell monolayers 2-fold over control.
24R,25(OH)₂D₃ reduces apoptotic markers in HCC38 cell monolayers, contrary to its effect in MCF7 cultures.

24R,25(OH)₂D₃ caused a dose-dependent decrease in B cell lymphoma protein 2-associated X protein (BAX) to B cell lymphoma protein 2 (BCL2) expression ratios (BAX/BCL2), which was significant at 10⁻⁸M and 10⁻⁷M doses (an ~8-fold decrease) (Figure 6.3A). 24R,25(OH)₂D₃ had a similar reducing effect on total p53 levels (Figure 6.3B) and TUNEL staining (Figure 6.3C), with the 10⁻⁷M dose producing the maximum decrease in p53 levels (~2 fold decrease) and TUNEL (~.25 fold decrease). 24R,25(OH)₂D₃-induced fold-change decreases in apoptosis in HCC38 cultures are similar in proportion to 24R,25(OH)₂D₃-induced fold change increases in apoptosis observed in MCF7 cultures (Figure 6.3D-F). However, in HCC38 cells, 24R,25(OH)₂D₃ has the greatest effect at 10⁻⁷M on BAX/BCL2, total p53, and TUNEL staining; unlike MCF7, where maximal effect sizes are observed at 10⁻⁸M 24R,25(OH)₂D₃.
Figure 6.3: 24R,25(OH)2D3 prevents apoptosis in HCC38 cells, but induces apoptosis in MCF7 cells.

HCC38 monolayer cultures were treated with 24R,25(OH)2D3 for 15 minutes, and harvested 12 hours later for gene expression and 24 hours later for protein and TUNEL staining. Cultures were assessed for apoptosis as measured by [A, D] BAX/BCL2 gene expression, [B, E] total p53 protein, and [C, F] TUNEL staining. [A-C] 24R,25(OH)2D3 reduced apoptosis in HCC38 cells. Groups that share a letter are not significant at p<0.05. [D-F] HCC38 or MCF7 monolayer cultures were treated with 24R,25(OH)2D3 for 15 minutes and assessed for apoptosis 12-24 hours later. Treatment over control was calculated as the average fold-change of treatment as compared to vehicle-treated cultures in three or more experiments and analyzed with a paired t-test against normalized vehicle controls *indicates significance at P<0.05 as compared to vehicle-treated cultures.
24R,25(OH)\textsubscript{2}D\textsubscript{3} increased epithelial-to-mesenchymal transition, metastatic markers, and migration in HCC38 cells in vitro.

Treatment with 10\textsuperscript{-8}M and 10\textsuperscript{-7}M 24R,25(OH)\textsubscript{2}D\textsubscript{3} significantly increased expression of the mesenchymal-associated transcription factor snail family transcriptional repressor 1 (SNAI1) compared to cells treated with vehicle (Figure 6.4A). Expression of receptor tyrosine-protein kinase erbB-2 (ERBB2) and matrix metalloproteinase 1 (MMP1), which are associated with tumor aggression and matrix turnover respectively, was also increased by similar doses of 24R,25(OH)\textsubscript{2}D\textsubscript{3}, with the maximal effect at 10\textsuperscript{-7}M (Figure 6.4B, 6.4C). 24R,25(OH)\textsubscript{2}D\textsubscript{3} had a biphasic effect on collagenase enzyme protein expression, with statistically significant induction of collagenase protein observed in cultures treated with 10\textsuperscript{-8}M 24R,25(OH)\textsubscript{2}D\textsubscript{3} (Figure 6.4D), and a non-significant increase in cultures treated with 10\textsuperscript{-7}M 24R,25(OH)\textsubscript{2}D\textsubscript{3}. 24R,25(OH)\textsubscript{2}D\textsubscript{3} (10\textsuperscript{-7}M) stimulated a 4-fold increase in SNAI1 (Figure 6.4E), a 6-fold increase in ERBB2 expression (Figure 6.4F), a 6-fold increase in MMP1 expression (Figure 6.4G), and a 10-fold increase in collagenase protein expression (Figure 6.4H) as compared to vehicle-treated cultures (Figure 6.4E-H). This is in direct contrast to the effect of 24R,25(OH)\textsubscript{2}D\textsubscript{3} on MCF7 cultures, where 10\textsuperscript{-7}M 24R,25(OH)\textsubscript{2}D\textsubscript{3} reduced SNAI1, and all doses of 24R,25(OH)\textsubscript{2}D\textsubscript{3} reduced ERBB2 and MMP1 transcript expression and reduced collagenase protein expression (Figure 6.4E-H).

24R,25(OH)\textsubscript{2}D\textsubscript{3} also increased metastatic markers in HCC38 cultures. 10\textsuperscript{-7}M 24R,25(OH)\textsubscript{2}D\textsubscript{3} increased chemokine receptor type 4 (CXCR4) to C-X-C-motif chemokine 12 (CXCL12) ratios (CXCR4/CXCL12) (Figure 6.4I). 10\textsuperscript{-8}M 24R,25(OH)\textsubscript{2}D\textsubscript{3} decreased the ratio of osteoprotegerin (OPG) to receptor activator of nuclear factor
kappa-B ligand (RANKL) transcripts (OPG/RANKL) (Figure 6.4J), and both 10^{-8}M and 10^{-7}M 24R,25(OH)_{2}D_{3} decreased OPG/RANKL protein expression (Figure 6.4K). Conditioned media from 24R,25(OH)_{2}D_{3}-treated HCC38 cells increased osteoclast activity more than conditioned media from vehicle-treated HCC38 cells (Figure 6.4L). 24R,25(OH)_{2}D_{3} increased osteoclast activity by 100% as compared to vehicle-treated cells. 10^{-7}M 24R,25 induced a 10-fold increase CXCR4/CXCL12 ratios (Figure 6.4M), and a 2-fold decrease in OPG/RANKL ratios (Figure 6.4N, 6.4O) in HCC38 cells. Both 10^{-8}M and 10^{-7}M 24R,25(OH)_{2}D_{3} decreased CXCR4/CXCL12 ratios and 10^{-9}M and 10^{-7}M increased OPG/RANKL transcript and protein expression ratios, respectively in MCF7 cells.
**Figure 6.4:** 24R,25(OH)2D3 increases epithelial-to-mesenchymal transition and metastatic markers in HCC38 cultures.
(Figure 4, cont. from the previous page): HCC38 and MCF7 monolayer cultures were treated with 24R,25(OH)$_2$D$_3$ for 15 minutes and harvested 12 hours later for gene expression. Effect of dose-dependent 24R,25(OH)$_2$D$_3$ on gene expression of [A, E] snail, [B, F] her2, [C, G] MMP1, [D, H] collagenase protein, [I, M] CXCR4/CXCL12 ratios, [J, N] OPG/ RANKL ratios, and [K, O] OPG/RANKL protein in [A-D, I-L] HCC38 cells, but reduces them in MCF7 cells [E-H, M-O]. [L] 24R,25(OH)$_2$D$_3$ increases osteoclast activity in osteoclasts treated with HCC38 conditioned media from cultures treated with 24R,25(OH)$_2$D$_3$. Groups that share a letter are not significant at p<0.05. Treatment over control was calculated as the average fold-change of treatment as compared to vehicle-treated cultures in three or more experiments and analyzed with a paired t-test against normalized vehicle controls *indicates significance at P<0.05 as compared to vehicle-treated cultures.
24R,25(OH)\textsubscript{2}D\textsubscript{3} increased cell migration in a scratch test assay. 12 hours after a wound was made on a confluent culture of HCC38 cells, wound size in cultures given a vehicle treatment had not significantly altered, while \(10^{-10}\)M 24R,25(OH)\textsubscript{2}D\textsubscript{3} had slightly decreased wound size, and \(10^{-7}\)M 24R,25(OH)\textsubscript{2}D\textsubscript{3} had reduced wound size to \(\sim25\%\) of its original area (Figure 6.5). 24 hours after the scratch test was initiated, all cultures had closed their wounds.

![Figure 6.5: Effect of 24R,25(OH)2D3 on cell migration in HCC38 cell monolayers.](image)

HCC38 monolayer cultures were treated with 24R,25(OH)\textsubscript{2}D\textsubscript{3} for 15 minutes and incubated with fresh media. A scratch was made across the culture with a 1mL pipette tip, and monolayer cultures were observed over 24 hours for wound closure.
24R,25(OH)₂D₃ differentially regulates PLD and PKC activity in HCC38 and MCF7 cells.

24R,25(OH)₂D₃ dose-dependently increases PLD activity in HCC38 cells, with the maximal effect occurring at 10⁻⁷M 24R,25(OH)₂D₃ (~3-fold increase as compared to vehicle-treated control cultures) (Figure 6.6A). However, 24R,25(OH)₂D₃ dose-dependently decreases PLD activity in MCF7 cells, with 10⁻⁷M reducing PLD activity to 25% that of vehicle-treated control cultures (Figure 6.6A). Treatment with 24R,25(OH)₂D₃ did not change PKC activity in HCC38 cells (Figure 6.6B); however, it did slightly increase PKC activity in MCF7 cells (Figure 6.6B). Baseline PKC activity in MCF7 cells was ~2-fold higher in MCF7 cells (~60 μU/μg protein/min) than in HCC38 cells (~30 μU/μg protein/min) (Figure 6.6B).
Modulation of 24R,25(OH)_2D_3 Tumorigenicity by ERα Isoforms in E_2-Responsive Cancer • Spring 2019

Rapid Enzyme Activity

Figure 6.6 24R,25(OH)_2D_3 reduces total p53 levels through a caveolae-associated, PLD-dependent mechanism.

HCC38 or MCF7 monolayer cultures were treated with 24R,25(OH)_2D_3 for 15 minutes, and assessed for [A] PLD activity decreases or [B] PKC activity. In a
separate set of experiments, HCC38 monolayer cultures were treated with inhibitors for 30 minutes before treatment with 24R,25(OH)$_2$D$_3$ for 15 minutes and assessed for total p53 content. 24R,25(OH)$_2$D3 reduced p53 levels in HCC38 cell monolayers, but this effect was prevented by treatment with [C] wortmannin to inhibit PLD activity, [D] 2-bromopalmitate to inhibit palmitoylation of nuclear receptors trafficked to the membrane, [E] methyl beta-cyclodextrin to deplete the membrane of cholesterol, and [F] short-hairpin RNA to silence caveolin-1.

24R,25(OH)$_2$D$_3$ inhibited apoptosis through a caveolae-associated, PLD-dependent pathway.

PLD inhibition with low-dose wortmannin prevented a 24R,25(OH)$_2$D$_3$-induced reduction in p53 (Figure 6.6 Error! Reference source not found.C). High-dose wortmannin reduced p53 levels in HCC38 cultures, but treatment with 24R,25(OH)$_2$D$_3$ reversed this effect and increased p53 levels back to baseline p53 levels in vehicle-treated control HCC38 cultures (Figure 6.6 Error! Reference source not found.C). 2-bromopalmitate (2BP) was used to prevent trafficking of palmitoylated nuclear receptors to the membrane. Treatment with all doses of 2BP prevented a 24R,25(OH)$_2$D$_3$-induced reduction in total p53 levels (Figure 6.6 Error! Reference source not found.D). Methyl β-cyclodextrin (MBC) was used to deplete the cell membrane of cholesterol and destroy caveolae and lipid rafts. Treatment with 1mM and 10mM MBC prevented a 24R,25(OH)$_2$D$_3$-induced reduction in total p53 levels (Error! Reference source not found.Figure 6.6E).
To evaluate the role of the caveolae on 24R,25(OH)_2D_3 signaling in HCC38 cells, HCC38 cells were stably knocked down for caveolin-1 with shRNA (Figure 6.7). Caveolin-1 knocked down HCC38 cells (shCAV1-HCC38) had <10% of wild-type caveolin-1 protein, while empty control-transduced cells had >100% of wild-type caveolin-1 protein expression (Figure 6.7A). Caveolin-1 levels were similar for both clone #1 and clone #2 shCAV1-HCC38 cells (Figure 6.7A, 6.7B). 24R,25(OH)_2D_3 reduced total p53 protein expression in wild-type and scramble control HCC38 cells, but did not affect shCAV1-HCC38 cells. Baseline levels of p53 in shCAV1-HCC38 cultures were lower than wild-type, and empty-control transduced HCC38 cultures (Error! Reference source not found. Figure 6.6F).

Caveolin-1 was stably knocked down in HCC38 using a recombinant plasmid with a puromycin antibiotic selection marker. Four short-hairpin clones targeting the CAV1 gene were screened for caveolin-1 silencing, and the two best clones (8002, “clone 1”, and 508, “clone 2”) were sub-cultured and expanded for further experiments. A separate set
of HCC38 cultures was also transfected with an empty cassette vector as a control. [A] Caveolin-1 silencing by shRNA particles and [B] Representative western blot (n=1).
Overexpressing and silencing ERα changes the 24R,25(OH)2D3 response.

ERα was transiently overexpressed in HCC38 cells (Figure 6.8A-D) and stably overexpressed in C2C12 (Figure 6.9A-C). ERα66 expression in ESR1-transfected HCC38 cells (ovERα-HCC38) was 20-times greater than wild-type HCC38 cells and 16-times greater than empty-control HCC38 cells (Figure 6.8C). ERα36 was 2.3-times greater in ESR1-transfected vs. wild-type HCC38 and 3-times greater than ORF-control HCC38 cells (Figure 6.8D). In C2C12 cells, ESR1-transfected C2C12 cells (ovERα-C2C12) expressed 11-times more ERα66 protein expression than wild-type C2C12 cells (Figure 6.9A), and 2.8-times more ERα36 than wild-type C2C12 (Figure 6.9B, 6.9C).

24R,25(OH)2D3 induced apoptosis in cells that do not express ERα66 but induced apoptosis in cells that express ERα66. In wild-type and ORF-control HCC38 cells, which express very low levels of ERα66 (Figure 6.8B, 6.8C), all doses of 24R,25(OH)2D3 prevented apoptosis as measured by BAX/BCL2 (Figure 6.11A) and total p53 levels (Figure 6.11B). However, treatment with 10^-8M 24R,25(OH)2D3 induced apoptosis in ovERα-HCC38 cells (Figure 6.11A, 6.11B) as measured by both BAX/BCL2 and total p53. 10^-7M 24R,25(OH)2D3 upregulated BAX/BCL2 ratios in ovERα-HCC38 (Figure 6.11A). 24R,25(OH)2D3 had a biphasic effect on total p53 levels in ovERα-HCC38. 10^-8 24R,25(OH)2D3 increased total p53 protein in ovERα-HCC38, but 10^-7M 24R,25(OH)2D3 did not affect p53 (Figure 6.11). Wild-type and ORF-control HCC38 cells had similar baseline ratios of BAX/BCL2 mRNA (Figure 6.11A) and similar baseline levels of total p53 protein (Figure 6.11B), but ovERα-HCC38 cells had lower ratios of BAX/BCL2 mRNA and lower baseline p53 than wild-type HCC38 (Figure 6.11A, 6.11B).
Figure 6.8 Overexpression of ESR1 in HCC38 changes ERα isoform expression.

[A] ERα and an empty control were transiently transfected in HCC38 using a recombinant plasmid with a GFP reporter gene. 48 hours after transfection, cells were assessed for ERα isoform expression by [B] western blot (representative image, n=1) and [C] ERα66 and [D] ERα36 protein expression were quantified (OV = overexpressed; KD = knockdown as compared to WT).
ERα was stably overexpressed in C2C12 using a recombinant plasmid with a G418 antibiotic selection marker. ovERα-C2C12 cells were selected for G418 resistance and assessed for protein expression via western blot. [A] ERα66 and [B] ERα36 protein expression were quantified (n=1) to confirm overexpression of ERα isoforms as compared to WT. [C] Representative western blot (n=1).

In C2C12 muscle cells, which express low levels of ERα66 (Figure 6.9A), 24R,25(OH)2D3 had an anti-apoptotic effect as measured by BAX/BCL2 mRNA ratios, but upregulated BAX/BCL2 mRNA ratios in ovERα-C2C12 cells (Figure 6.11C). Wild-type C2C12 cultures had higher baseline BAX/BCL2 mRNA ratios than ovERα-C2C12 cultures (Figure 6.11C). 24R,25(OH)2D3 did not affect total p53 levels in wild-type C2C12 cells but
increased total p53 in ovERα-C2C12 (Figure 6.11D). Baseline total p53 was higher in ovERα-C2C12 cells than in wild-type C2C12 (Figure 6.11D). In both C2C12 and HCC38 cells, transfecting with ESR1 to overexpress the ERα isoforms changed the cell response to 24R,25(OH)2D3. 24R,25(OH)2D3 stimulated apoptosis in all HCC38 and C2C12 cultures that overexpressed ERα66 (Figure 6.11A-D).

ERα66 was stably knocked down in MCF7 cells (Figure 6.10A, 6.10C). Knocked down MCF7 (shESR1-MCF7) cells had approximately one-third the ERα66 expression of wild-type MCF7 cells, while scrambled controls had >80% of wild-type MCF7 cells expression Figure 6.10A. ERα36 levels were similar for wild-type, scramble control, and shESR1-MCF7 cells, with the scrambled control and the shESR1-MCF7 cells expressing ~90% as much ERα36 as wild-type cells (Figure 6.10B, 6.10C).
Figure 6.10 ERα66 was stably knocked down in MCF7 using a recombinant plasmid with a puromycin antibiotic resistant selection marker.

shESR1 and scramble-control cells were selected for puromycin resistance and assessed for protein expression via western blot. [A] ERα66 and [B] ERα36 protein expression were quantified (n=1) to confirm silencing of ERα66 as compared to WT. [C] Representative western blot (n=1).
24R,25(OH)₂D₃ upregulated BAX/BCL2 mRNA ratios and increased total p53 levels in wild-type and scrambled MCF7 cells at 10⁻⁸M (Figure 6.11E, 6.11F). 24R,25(OH)₂D₃ had no effect on BAX/BCL2 mRNA ratios or total p53 levels in shESR1-MCF7 cells (Figure 6.11E, 6.11F). Baseline BAX/BCL2 mRNA ratios and total p53 were the same in wild-type, scrambled, and shESR1 MCF7 cells. In wild-type, MCF7 cells, 10⁻⁷M 24R,25(OH)₂D₃ also increased phosphorylated p53 but did not increase phosphorylated p53 in cultures that were knocked down for ERα66 (Figure 6.12). While 24R,25(OH)₂D₃ increased the average amount of phosphorylated p53 in scramble-control MCF7 cells as compared to vehicle-treated scramble-control MCF7 cells, this change was not statistically significant. Similarly, 10⁻⁷M 24R,25(OH)₂D₃ reduced average phosphorylated p53 in shESR1-MCF7 cells as compared to vehicle-treated shESR1-MCF7 cells; however, this difference was not statistically significant. Baseline phosphorylated p53 was not statistically different between wild-type, scrambled, and shESR1-MCF7 cells (Figure 6.12).
Figure 6.11: Expression and silencing of ERα isoforms change the 24R,25(OH)2D3 response.
(Figure 7, cont. from the previous page):

Overexpressing all ERα isoforms reversed the 24R,25(OH)₂D₃ anti-apoptotic effect in [A, B] HCC38 cells and [C, D] C2C12 cells. 24R,25(OH)₂D₃ prevented apoptosis as measured by [A] BAX/BCL2 and [B] total p53 levels in wild-type HCC38 and HCC38 cells transfected with an empty control vector, but induced apoptosis in ERα-transfected HCC38 cells. In C2C12 muscle cells, 24R,25(OH)₂D₃ prevented or did not induce apoptosis in wild-type cells but induced apoptosis in ERα-transfected cells as measured by [C] BAX/BCL2 and had no effect on apoptosis as measured by [D] total p53 levels in wild-type cells. Silencing the ERα66 isoform reversed the pro-apoptotic effect of 24R,25(OH)₂D₃ on [E, F] MCF7 cells. 24R,25(OH)₂D₃ induced apoptosis as measured by [E] BAX/BCL2 and [F] total p53 levels in wild-type and scramble-control MCF7 cells, but had no effect on apoptosis in ERα66-knocked down MCF7 cells.
Figure 6.12 Silencing the ERα66 isoform reversed the pro-apoptotic effect of 24R,25(OH)2D3 on MCF7 cells.

24R,25(OH)2D3 significantly increased phosphorylated p53 levels in wild-type MCF7 cells and qualitatively increased phosphorylated p53 in scramble-control MCF7 cells. In MCF7 cells knocked down for ERα66, 24R,25(OH)2D3 qualitatively decreased average phosphorylated p53.
Discussion

Vitamin D metabolites may have anti-tumorigenic properties in breast cancer; however, many vitamin D3 metabolites are highly calcemic and have narrow therapeutic windows (34). Previously we identified an alternative endogenous vitamin D3 metabolite, 24R,25(OH)2D3, which has anti-tumorigenic and pro-apoptotic effects in the ERα66-positive breast cancer cell lines MCF7 and T47D (21) but, unlike other vitamin D3 metabolites, 24R,25(OH)2D3 is not calcemic at therapeutic doses (21,35). However, recent studies have shown that vitamin D supplementation may not be beneficial for all cancer patients (5,6), and some studies have shown that patients with estrogen receptor positive (ERα66+) breast cancer benefit from vitamin D supplements, but patients with estrogen receptor negative (ERα66-) breast cancer do not (4,7,36). The dependence of the estrogen receptor and its isoforms on the tumorigenicity of vitamin D metabolites is an understudied area of breast cancer research, and more investigation into the crosstalk between estrogen receptors and vitamin D metabolites is necessary to refine treatment protocols and vitamin D supplement regimens.

In this paper, the effects of 24R,25(OH)2D3 were investigated in vivo in an HCC38 xenograft orthotopic mammary fat pad model of breast cancer in an immuno-compromised NSG mouse, and in vitro in cell monolayer cultures of HCC38. The effects of 24R,25(OH)2D3 on models of HCC38 (ERα66-) breast cancer were contrasted with the effects of 24R,25(OH)2D3 in MCF7 (ERα66+); the ERα66 receptor was overexpressed and silenced in HCC38, MCF7, and a third unrelated cell line, C2C12, to understand how the expression of ERα66 modulates the effect of 24R,25(OH)2D3. Although 24R,25(OH)2D3 was not calcemic in vivo, treatment with 24R,25(OH)2D3 increased tumor
burden and decreased animal survival in mice with HCC38 (ERα66-) xenografts. In vitro, 24R,25(OH)2D3 specifically decreased apoptosis and increased proliferation, metastasis, and migration in HCC38 (ERα66-) cells. This pro-tumorigenic effect is in direct contrast to the pro-apoptotic and anti-metastatic effects of 24R,25(OH)2D3 on MCF7 cells (ERα66+). Overexpressing and silencing ERα66 found that in cells that do not express ERα66, 24R,25(OH)2D3 does not stimulate apoptosis and may be tumorigenic. In cells that express ERα66, 24R,25(OH)2D3 is pro-apoptotic and anti-tumorigenic. Our results suggest that ERα66 directly modulates the apoptotic effects of 24R,25(OH)2D3.

Our previous work showed that 24R,25(OH)2D3 is anti-tumorigenic in mammary fat pad xenografts of MCF7 (ERα66+) in NOD.Cg-PrkdcscidIl2rgtm1Wjl/SzJ (NSG) mice. Here, we use a similar in vivo model to test the effects of 24R,25(OH)2D3 on tumor burden in mammary fat pad xenografts of HCC38 (ERα66-) in NSG mice. These models allowed us to study the effects of 24R,25(OH)2D3 directly on the tumor, rather than the direct and indirect effects of 24R,25(OH)2D3 on the tumor and concomitant immunity. However, it is important to note that 24R,25(OH)2D3 has been shown to modulate the immune system, and clinically 24R,25(OH)2D3 could also indirectly affect tumor invasion and metastasis. Other studies have shown that both 1,25(OH)2D3 and 24R,25(OH)2D3 can alter the production of pro-and anti-inflammatory cytokines (16,17). 1α,25(OH)2D3 reduced the production of tumor necrosis factor α (TNFα) and interleukin-10 (IL-6) by tumor-associated monocytes (17). In our lab, we have shown that 24R,25(OH)2D3 reduced serum levels of TNFα, IL-6, interleukin-1α, interleukin-1β, and other pro-inflammatory factors in Sprague-Dawley rats with surgically transected anterior crucial ligaments (16). While studies in this paper with immune compromised animals have shown that
24R,25(OH)$_2$D$_3$ is capable of directly modulating tumor growth in vivo, it is possible that 24R,25(OH)$_2$D$_3$ may also alter the immune-regulation of carcinomas in immunocompetent animals, and more studies are required to explore this possibility further.

Previous results by our lab and others have shown that 24R,25(OH)$_2$D$_3$ has anti-carcinogenic properties, particularly in models of breast, lung, and glandular stomach carcinomas (18,19,21). However, high expression of 24-hydroxylase (CYP24A1), the enzyme responsible for metabolizing 25(OH)D$_3$ into 24R,25(OH)$_2$D$_3$, is associated with increased tumor aggression in several cancers including breast cancer (18–20,37). 24R,25(OH)$_2$D$_3$ has been shown to act both directly on several cell types such as bone, cartilage, kidney, and cancer cells, and indirectly by increasing the production of 1α-hydroxylated vitamin D$_3$ metabolites in vivo (38,39). Physiologically, 24R,25(OH)$_2$D$_3$ increases the expression of 1α-hydroxylase (CYP27B1) and 1α,25(OH)$_2$D$_3$, in turn, increases the expression of 24-hydroxylase (CYP24A1) (40). While HCC38 cells express both CYP24A1 and CYP27B1, our results and others have shown that MCF7 cells express CYP27B1 but do not express CYP24A1 (37,41). 1α,25(OH)$_2$D$_3$, a product of CYP27B1 activity, has been shown to have anti-tumorigenic activity in MCF7 cells in vivo and in vitro (42,43). CYP24A1 is a commonly mutated gene in cancers (44), and further study is needed to determine whether the tumorigenicity of 24R,25(OH)$_2$D$_3$ is related to native CYP24A1 expression within the tumor. It is possible that the MCF7 cells respond anti-tumorigenically to 24R,25(OH)$_2$D$_3$ because they are CYP24A1 negative, and this point should be explored further in future studies. Previous studies in our lab show that MCF7 cells respond with the greatest fold-change to 10$^{-9}$M 24R,25(OH)$_2$D$_3$ (21), while in
this paper we show that HCC38 cells are more sensitive to $10^{-7}$M 24R,25(OH)$_2$D$_3$. This, coupled with the lack of CYP24A1 expression in MCF7, may indicate that MCF7 cells are more sensitive to 24R,25(OH)$_2$D$_3$ than HCC38 cells. This sensitivity may be a result of an increase in CYP27B1 and 1α-hydroxylated vitamin D$_3$ metabolite. Furthermore, our results with 24R,25(OH)$_2$D$_3$ on MCF7 and HCC38 cells may not be reflective of the direct effects of 24R,25(OH)$_2$D$_3$, but could be a result of shifting proportions of other vitamin D$_3$ metabolites caused by an influx of 24R,25(OH)$_2$D$_3$.

However, our data on the effect of 24R,25(OH)$_2$D$_3$ on phospholipase D (PLD) and protein kinase C (PKC) activity in breast cancer cells suggest that 24R,25(OH)$_2$D$_3$ can directly affect breast cancer cells. In male rat resting zone chondrocytes, the rapid actions of 24R,25(OH)$_2$D$_3$ are mediated by PLD, while the rapid actions of both 17β-estradiol and 1α,25(OH)$_2$D$_3$ are mediated by protein kinase C (PKC) (45). Our data show a similar effect, with 24R,25(OH)$_2$D$_3$ rapidly changing PLD activity in both HCC38 and MCF7, but not affecting PKC activity in HCC38 cells at the same time point. 24R,25(OH)$_2$D$_3$ slightly increased PKC activity in MCF7 cells at high doses, but this effect had a smaller fold change than the effect size of 24R,25(OH)$_2$D$_3$ on PLD activity in MCF7 cells. The rapid effect of 24R,25(OH)$_2$D$_3$ on PLD rather than PKC activity in HCC38 and MCF7 cells suggests that in vitro, 24R,25(OH)$_2$D$_3$ acts directly on both HCC38 cells and MCF7 cells rather than stimulating the production of another vitamin D$_3$ metabolite which is responsible for the effects of 24R,25(OH)$_2$D$_3$ on apoptosis, epithelial-to-mesenchymal transition, migration, and metastatic markers in these cells.

The connection between 24R,25(OH)$_2$D$_3$ and ER$\alpha$66 has implications for many different cell types and tissues. Firstly, 24R,25(OH)$_2$D$_3$ has an established
chondroprotective effect and may prevent bone resorption by limiting calcium efflux in cartilage and bone cells (46). Post-menopausal women have a higher incidence of osteoarthritis than men of similar ages (47,48), and the extremely low levels of circulating estradiol in post-menopausal women have been implicated in this discrepancy (48,49); estradiol has a protective effect on bone, bone resorption, cartilage (48,50). Treatment with 24R,25(OH)₂D₃ has been proposed as a potential therapeutic for osteoarthritis patients, as 24R,25(OH)₂D₃ prevents oxidative damage and apoptosis in chondrocytes (16). However, all previous research on the chondroprotective effects of 24R,25(OH)₂D₃ has previously been done in male chondrocytes (16), who have higher circulating estradiol than post-menopausal women and lower expression of ERα66 (48,51,52). Further research on 24R,25(OH)₂D₃ and ERα66 should be done to ensure that the chondroprotective effects of 24R,25(OH)₂D₃ are clinically applicable to both men and women, and male and female chondrocytes may be an appropriate model to further investigate crosstalk between 24R,25(OH)₂D₃ and ERα.

Secondly, 24R,25(OH)₂D₃ is a naturally occurring ubiquitous metabolite of vitamin D₃. Serum 24R,25(OH)₂D₃ is present at higher levels in the blood than 1α,25(OH)₂D₃, the more commonly studied active metabolite of vitamin D₃ (53). Women with breast cancer have been reported to take vitamin D₃ supplements at a higher rate than the general population (4), and some of that supplemental vitamin D₃ is undoubtedly getting metabolized into 24R,25(OH)₂D₃ (54). Further investigation into the benefits of taking vitamin D₃ supplements for patients with ERα66+ and ERα66- breast cancer needs to be conducted, as our results imply that women with ERα66- breast cancer may be aggravating their tumors by taking excess vitamin D₃.
Conclusion

In summary, 24R,25(OH)\textsubscript{2}D\textsubscript{3} modulates apoptosis and tumorigenicity in breast cancer cells through an ER\textalpha\textsubscript{66}-associated pathway. In ER\textalpha\textsubscript{66}-negative breast tumors, 24R,25(OH)\textsubscript{2}D\textsubscript{3} increases tumor burden, migration, and metastasis, and may decrease survival. In ER\textalpha\textsubscript{66}-positive tumors, 24R,25(OH)\textsubscript{2}D\textsubscript{3} induces apoptosis and reduces tumor burden. Our results show that overexpressing ER\textalpha\textsubscript{66} in wild-type ER\textalpha\textsubscript{66}-negative HCC38 cells reverses the anti-apoptotic effect of 24R,25(OH)\textsubscript{2}D\textsubscript{3} and causes treatment with 24R,25(OH)\textsubscript{2}D\textsubscript{3} to induce apoptosis; likewise, silencing ER\textalpha\textsubscript{66} in wild-type ER\textalpha\textsubscript{66}-positive MCF7 cells eliminates the pro-apoptotic effect of 24R,25(OH)\textsubscript{2}D\textsubscript{3}. Our results with C2C12, an unrelated cell line, showed that the regulation of the 24R,25(OH)\textsubscript{2}D\textsubscript{3} effect by ER\textalpha\textsubscript{66} is not limited to breast cancer cells; rather, treatment with 24R,25(OH)\textsubscript{2}D\textsubscript{3} induced apoptosis in ER\textalpha\textsubscript{66}-overexpressing C2C12 cells. Our data show that ER\textalpha\textsubscript{66} is a modulator of the apoptotic effect of 24R,25(OH)\textsubscript{2}D\textsubscript{3} in a diverse array of cell lines, and should inform diverse research areas such as the implications of vitamin D supplementation in breast cancer patients and the potential of 24R,25(OH)\textsubscript{2}D\textsubscript{3} as a therapeutic in male vs. female osteoarthritis patients.
Chapter 7.

Conclusions and Future Perspectives

Although high serum 25(OH)D$_3$ is often associated with improved prognosis and reduced risk of tumor recurrence in breast cancer patients, double-blind clinical trials investigating the efficacy of vitamin D$_3$ supplementation in cancer have been inconclusive. This is in part because breast cancer is a multifaceted disease with genetic and phenotypic variations. As such, the idea of making blanket treatment recommendations on vitamin D$_3$ supplementation may be as impractical as suggesting a single, defined cure for all types of breast cancer. Preliminary research on vitamin D$_3$ supplementation in breast cancer suggests that ER+ patients may benefit from vitamin D$_3$ supplementation, but ER- patients may not. This thesis demonstrates that 24R,25(OH)$_2$D$_3$ is functionally active in breast cancer cells and that the tumorigenicity of 24R,25(OH)$_2$D$_3$ is regulated in part by ERα66. Furthermore, this work suggests that crosstalk between 24R,25(OH)$_2$D$_3$ and ERα66 may not be cell-type specific, and the actions of 24R,25(OH)$_2$D$_3$ on apoptosis may have implications for other disease states.

Like other steroid hormones, 24R,25(OH)$_2$D$_3$ has been found to initiate multiple rapid signaling pathways from the cell membrane in chondrocytes, and a membrane receptor has been implicated, which remains the subject of active investigation. Here, we show evidence that the rapid actions of 24R,25(OH)$_2$D$_3$ are conserved across cell types, with 24R,25(OH)$_2$D$_3$ regulating proliferation and apoptosis in MCF7, HCC38, and C2C12 cells – three unrelated cell lines from different species, tissues, and with different ERα profiles. Furthermore, the regulation of proliferation and apoptosis by 24R,25(OH)$_2$D$_3$ was
specific to the ‘R’ enantiomer. 24S,25(OH)2D3 did not have an effect on apoptosis in either MCF7 or HCC38 cells.

Our results also demonstrated that in MCF7 and HCC38 cells, 24R,25(OH)2D3 stimulates PLD at 9 minutes. Extensive studies in chondrocytes have shown that 24R,25(OH)2D3 also stimulates PLD after 9 minutes15. In proliferative chondrocytes, 24R,25(OH)2D3 has been shown to prevent apoptosis through a PLD-dependent mechanism by promoting p53 degradation via ubiquitination. This is similar to the mechanism we observed in HCC38 breast cancer cells, where 24R,25(OH)2D3 prevented apoptosis by decreasing p53 production. This suggests that 24R,25(OH)2D3 regulates apoptosis in proliferating chondrocytes and ERα66-negative breast cancer cells through the same PLD, p53-dependent mechanism.

Many of the second messengers of the rapid signaling pathway of 24R,25(OH)2D3, such as PLD and PKC, are also implicated in 17β-estradiol (E2) signaling through the membrane receptor ERα36. Our results show that E2 signaling through ERα36 is tumorigenic in laryngeal cancer as well as breast cancer, suggesting that E2 signaling through ERα36 might also regulate apoptosis in other disease states. We also show that, as in breast cancer, ERα66-, ERα36+ laryngeal cancer is more aggressive than its ERα66+, ERα36+ counterparts.

Many of the rapid signaling pathways activated by E2 binding to ERα36 in laryngeal cancer are also activated by 24R,25(OH)2D3 in breast cancer. Our results suggest that the rapid actions of 24R,25(OH)2D3 in MCF7 (ERα66+, ERα36+) breast cancer cells may be modulated by a membrane-mediated, caveolae-localized pathway employing a palmitoylated nuclear receptor to stimulate PLD with downstream regulatory effects on
Modulation of 24R,25(OH)₂D₃ Tumorigenicity by ERα Isoforms in E₂-Responsive Cancer • Spring 2019

Anjali Verma • Ph.D. Dissertation • Biomedical Engineering • Virginia Commonwealth University 204

apoptosis, proliferation, migration, and metastasis. In MCF7 (ERα66+, ERα36+), 24R,25(OH)₂D₃ confers a chemoprotective effect against breast cancer cells in vitro and in vivo to prevent tumorigenesis and reduce metastasis.

Our work with MCF7 (ERα66+, ERα36+) showed that the chemoprotective effects of 24R,25(OH)₂D₃ were dependent upon activation of isoforms of ERα in this cell line. This suggested that ERα isoforms may modulate 24R,25(OH)₂D₃ in breast cancer cells. The effects of 24R,25(OH)₂D₃ on tumorigenicity were examined in a second cell line, HCC38, which does not express ERα66 (ERα66-, ERα36+). 24R,25(OH)₂D₃ was tumorigenic in HCC38 (ERα66-, ERα36+), and this tumorigenicity was initiated by a PLD-dependent signaling mechanism. Overexpressing ERα66 in HCC38 (ERα66-, ERα36+) reversed the tumorigenic effects of 24R,25(OH)₂D₃, suggesting that ERα66 may modulate the rapid actions of 24R,25(OH)₂D₃. We also overexpressed ERα66 and ERα36 in an unrelated cell line, C2C12, and found that 24R,25(OH)₂D₃ increased apoptosis in C2C12 cells that overexpressed ERα66 and ERα36. And wild-type MCF7 (ERα66+, ERα36+) and ERα66-silenced MCF7 (ERα66-, ERα36+) were tested for 24R,25(OH)₂D₃-induced apoptosis, and we found that in wild-type and scramble-transfected MCF7 cells, 24R,25(OH)₂D₃ induced apoptosis; however in ERα66-silenced MCF7 cells, 24R,25(OH)₂D₃ did not stimulate apoptosis. This demonstrates that the pro-apoptotic effect of 24R,25(OH)₂D₃ is dependent upon the ERα66 isoform across cell lines.

The implications of this in the wake of similarities between 24R,25(OH)₂D₃ signaling in chondrocytes, MCF7, and HCC38 cells are interesting. The 24R,25(OH)₂D₃ signaling pathway was discovered and tested in male rat chondrocytes, where 24R,25(OH)₂D₃ was found to prevent apoptosis. Male rat chondrocytes express all
isoforms of ERα, including ERα66 and ERα36. However, female rat chondrocytes express roughly equal amounts of ERα66 in the plasma membrane, caveolae, and nucleus. Male rat chondrocytes expressed more ERα66 in the plasma membrane and caveolae fraction than in the nucleus. Furthermore, ERα36 was found at similar levels in the plasma membrane and nucleus of female rat chondrocytes. However male rat chondrocytes only expressed ERα36 in the nucleus, and did not express detectable levels of ERα36 in the plasma membrane or the caveolae. It is worth noting that ERβ levels were the same in female and male rat chondrocytes. Similarly, MCF7 cells express high levels of ERα isoforms in the plasma membrane and nucleus, while HCC38 cells only express ERα isoforms in the membrane. Perhaps the pro-apoptotic effects of 24R,25(OH)2D3 are regulated not by ERα66 itself, but by an abundance of nuclear ERα isoforms rather than an abundance of membrane ERα isoforms.

This work provides a foundation for the further investigation of 24R,25(OH)2D3 as a modulator of tumorigenicity in breast cancer, and suggests that vitamin D3 supplementation should only be considered for patients with ER+, or ERα66+, breast cancer. Our lab has begun developing a means of assessing 24R,25(OH)2D3 metabolite expression by HPLC. We plan to use this technique to measure the local production of 24R,25(OH)2D3 in breast tissue. Later on, we plan to assess 24R,25(OH)2D3 metabolite levels in banked serum samples from breast cancer patients. A meta-analysis can be conducted to assess correlations between circulating 24R,25(OH)2D3 metabolite expression and ERα66 and ERα36 expression in breast cancer.

Preliminary results from these studies also suggest that 24R,25(OH)2D3 can alter markers of osteoclast activation in breast tumors. In MCF7 (ERα66+, ERα36+)
24R,25(OH)\(_2\)D\(_3\) reduced markers of osteoclast activation and in HCC38 (ER\(\alpha\)66-, ER\(\alpha\)36+) 24R,25(OH)\(_2\)D\(_3\) enhanced markers of osteoclast activation and osteoclast activity in HCC38. Stage 4 ER+ breast cancer commonly metastasizes to the long bone and spine, and 5-year survival rates for patients with late-stage bone metastasis are around 22%. 24R,25(OH)\(_2\)D\(_3\) may have potential as a therapeutic for these patients. Our lab has established a model of breast tumor osteolysis in murine femurs, and we are planning to examine the effect of 24R,25(OH)\(_2\)D\(_3\) on models of ER+ and ER-breast cancer metastasis in mice. If these pre-clinical studies show that 24R,25(OH)\(_2\)D\(_3\) is efficacious against ER+ breast cancer metastasis, we would suggest a clinical trial on the use of 24R,25(OH)\(_2\)D\(_3\) supplementation in breast cancer patients.
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Vita

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